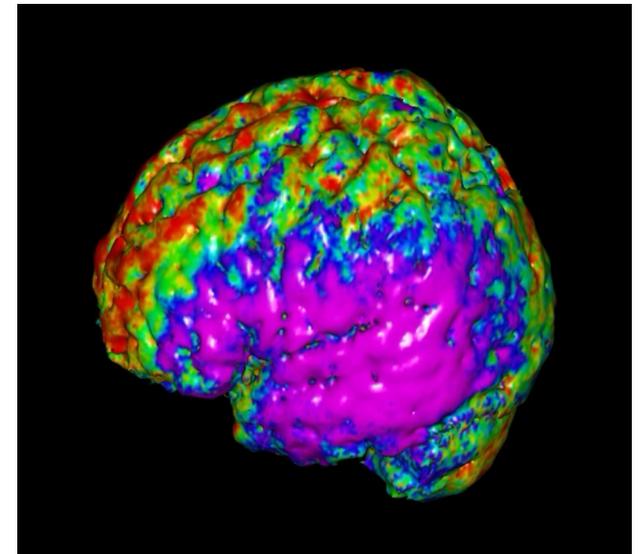


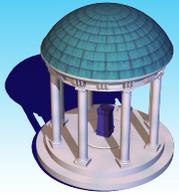
Statistical Issues in Imaging Studies

Hongtu Zhu, Ph.D.

**Department of Biostatistics and Biomedical
Research Imaging Center**

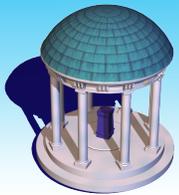
University of North Carolina at Chapel Hill





UNC Biostatistics and Imaging Analysis Lab

- 2 Faculty Members**
- 10 Ph.d Students**
- 5 Postdoctor Fellows**
- 2 Visitor Scholars**
- 4 Past members**



UNC Biostatistics and Imaging Analysis Lab

UNC Gillings School of Global Public Health



THE UNIVERSITY
of NORTH CAROLINA
at CHAPEL HILL

UNC BIostatISTICS AND IMAGING ANALYSIS LAB (BIA)

[HOME](#)

[PEOPLE](#)

[GRANTS](#)

[PUBLICATIONS](#)

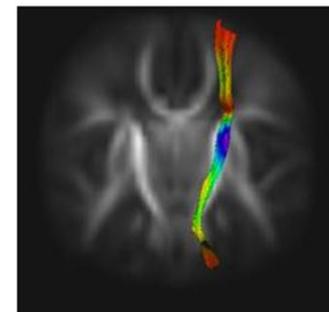
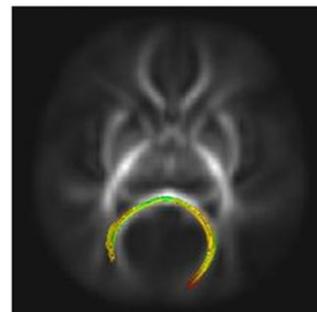
[SOFTWARE](#)

[COLLABORATION](#)

[JOURNAL CLUB](#)

[GROUP MEETING](#)

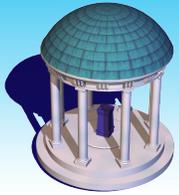
[CONTACT](#)



About Us

We have diverse interest in solving methodological issues in statistics. Our past and present statistical projects include diagnostic measures, stochastic approximation algorithm, structural equation models, mixed effect models, spline regression, missing data problems, variable selections, empirical likelihood, mixture models and regression tree.

We have developed methods and software for the analysis of the data from a state-of-the art magnetic resonance imaging (MRI) technique including MRI, functional MRI, and diffusion tensor image. We have developed and enhanced tools in data mining, Monte Carlo method, statistical modeling, and applied them to scientific problems to understand the function and structure of the brain. Our collaborators and we work closely to study healthy and neurologically disordered children and adults.



UNC Biostatistics and Imaging Analysis Lab

Sequence Evaluation

Sequence Optimization

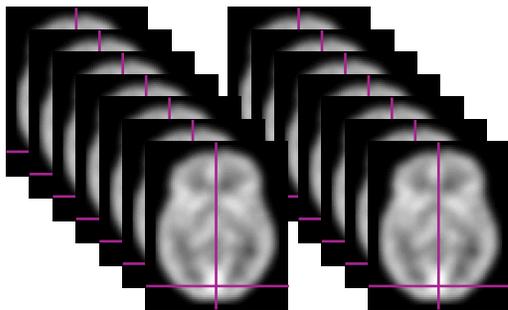
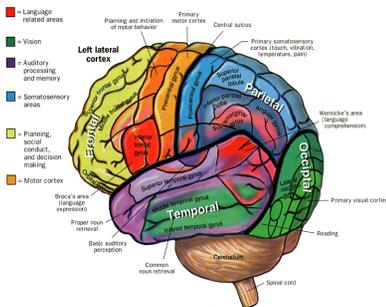
Reconstruction

Smoothing

Population Analysis

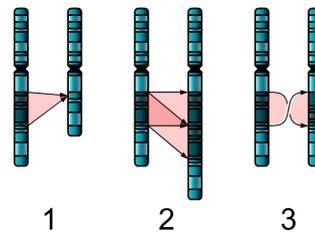
Data Mining

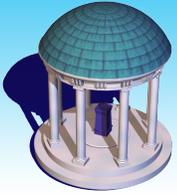
Imaging Genetics



group 1

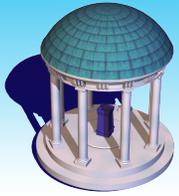
group 2





Outline

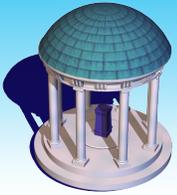
- **Evaluating** Imaging Sequence Accuracy
- Imaging Sequence **Optimization**
- **Reconstructing** Diffusion Tensor Images
- **Smoothing** Diffusion Tensor Field
- **Analyzing** Tract-based Diffusion Tensor Statistics
- **Multiscale Adaptive Regression** Models
- **Brain Connectivity** Analysis



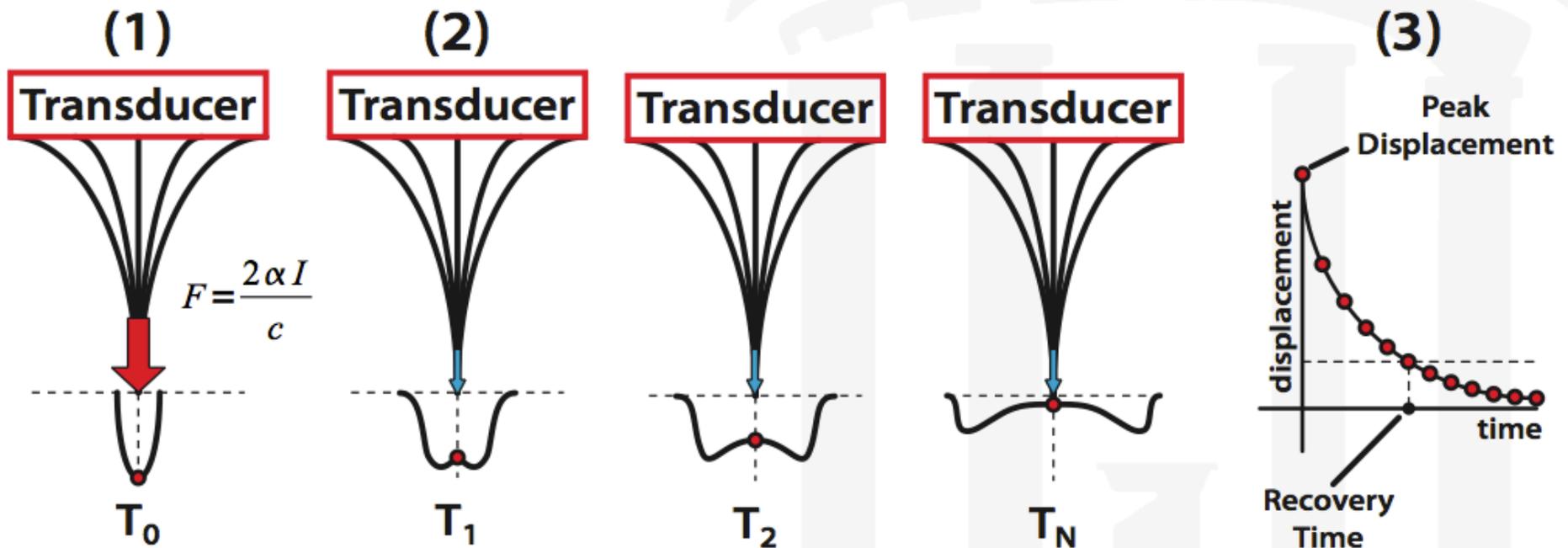
Evaluating Imaging Sequence Accuracy

Statistical Methods in Diagnostic Medicine

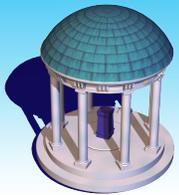
**ARFI Beam Sequence Performance as Evaluated
by Trained Readers: Plaque Detection**
PI. Caterina M. Gallippi



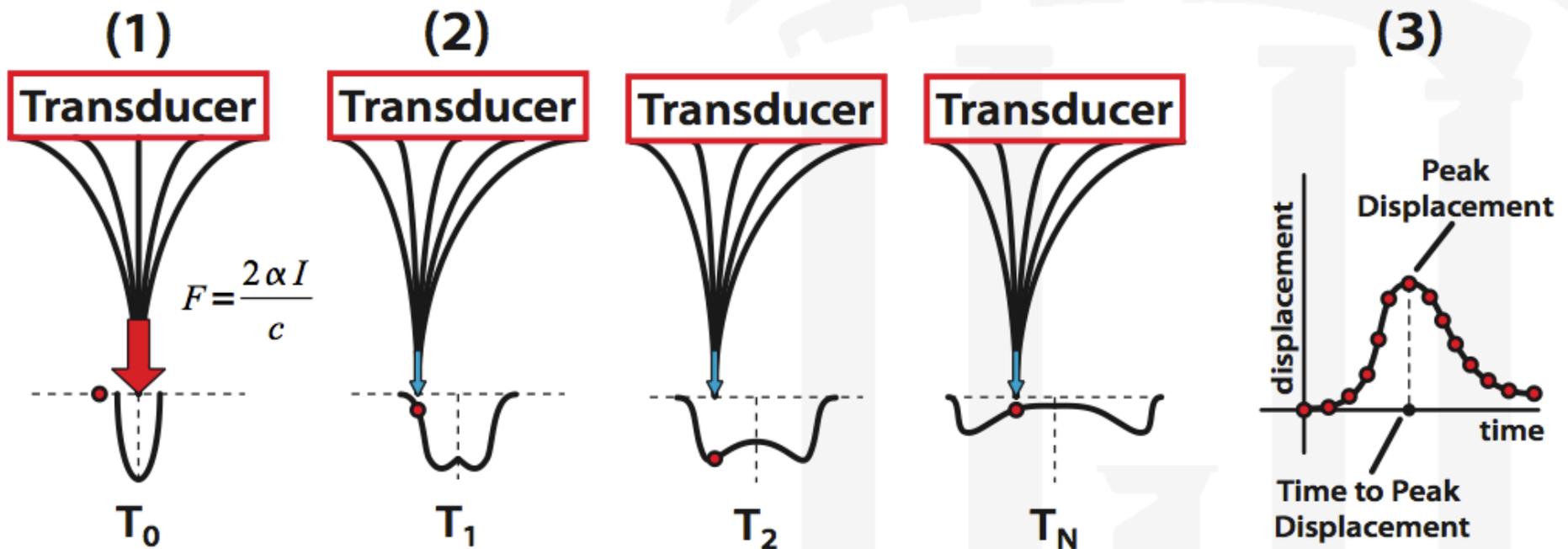
Methods of Acoustic Radiation Force Impulse (ARFI) Ultrasound



- (1) ARFI Excitation Pulse ($\sim 70\mu\text{s}$) induces axial displacement**
- (2) Conventional B-Mode pulses track induced displacement**
- (3) Displacements are calculated to create a displacement profile for every pixel within the image.**



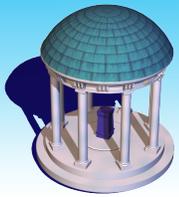
Methods of Shear Wave Elasticity Imaging (SWEI)



(1) ARFI Excitation Pulse ($\sim 70\mu\text{s}$) induces axial displacement

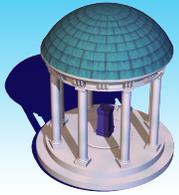
(2) Tracking Away From Region of Excitation Tracks ARFI-induced Shear Waves

(3) Displacement Profiles Are Created



Hypothesis

Select ARFI/SWEI beam sequences will yield higher sensitivity and specificity for atherosclerotic plaque detection in peripheral arteries.



General Methods: Beam Sequences

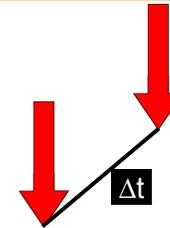
- 3 Types of Excitation



Single F/1.5
(SP1.5)

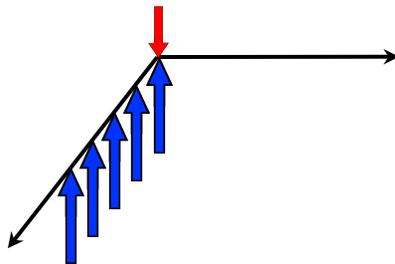


Single F/3
(SP3)

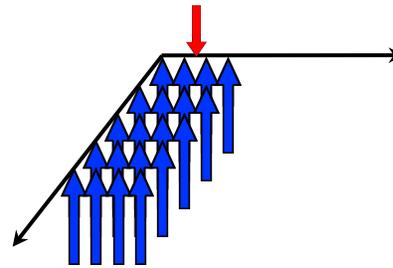


Double F/1.5
(DP)

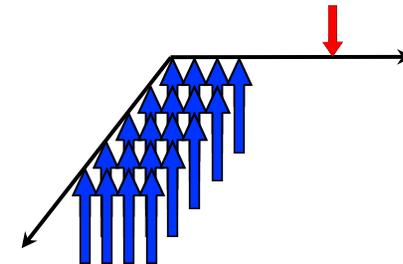
- 3 Types of Tracking



Single A-line RX
(SRx)

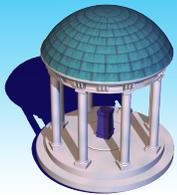


4:1 Parallel RX
(ParRx)



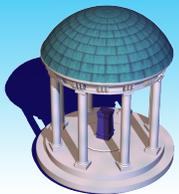
SWEI

- Combine for 9 Total Sequences

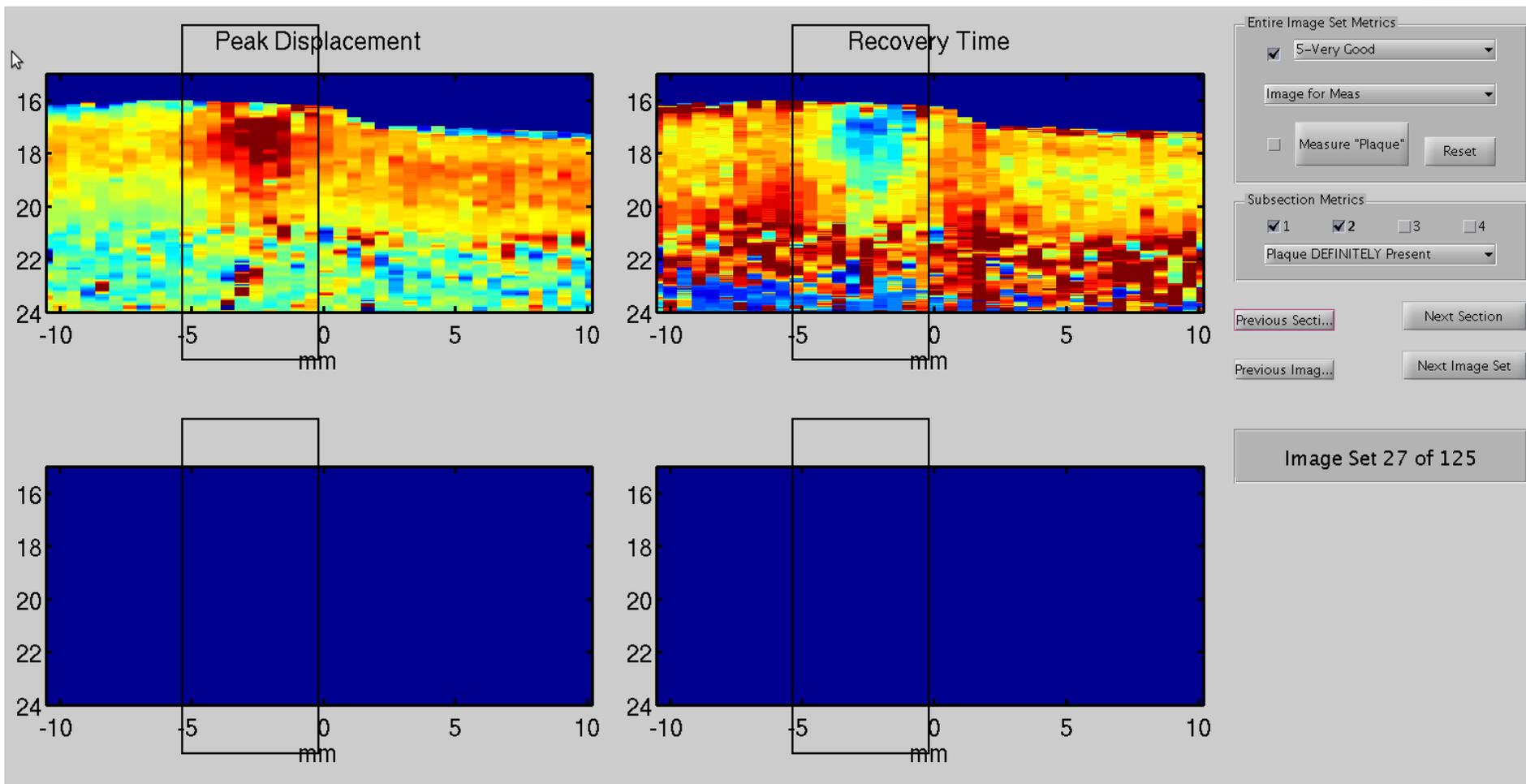


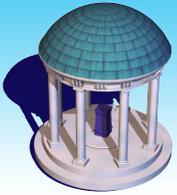
Reader Study

- **Automatically Generated Images**
 - ◆ **Lumen masking & Color scaling**
- **12 Trained Readers**
 - ◆ **Various levels of experience with ARFI**
 - ◆ **Only 6 evaluated each Image Set**
- **Validation**
 - ◆ **Phantom results compared with known truth**
 - ◆ **Ex vivo results compared with pathologist rating of spatially matched histology**
- **Statistics**
 - ◆ **Latent Variable Models to compare ordinal responses**
 - ◆ **Generated receiver operating characteristic (ROC) curves**
 - ◆ **Calculated mean area under the curve (AUC)**



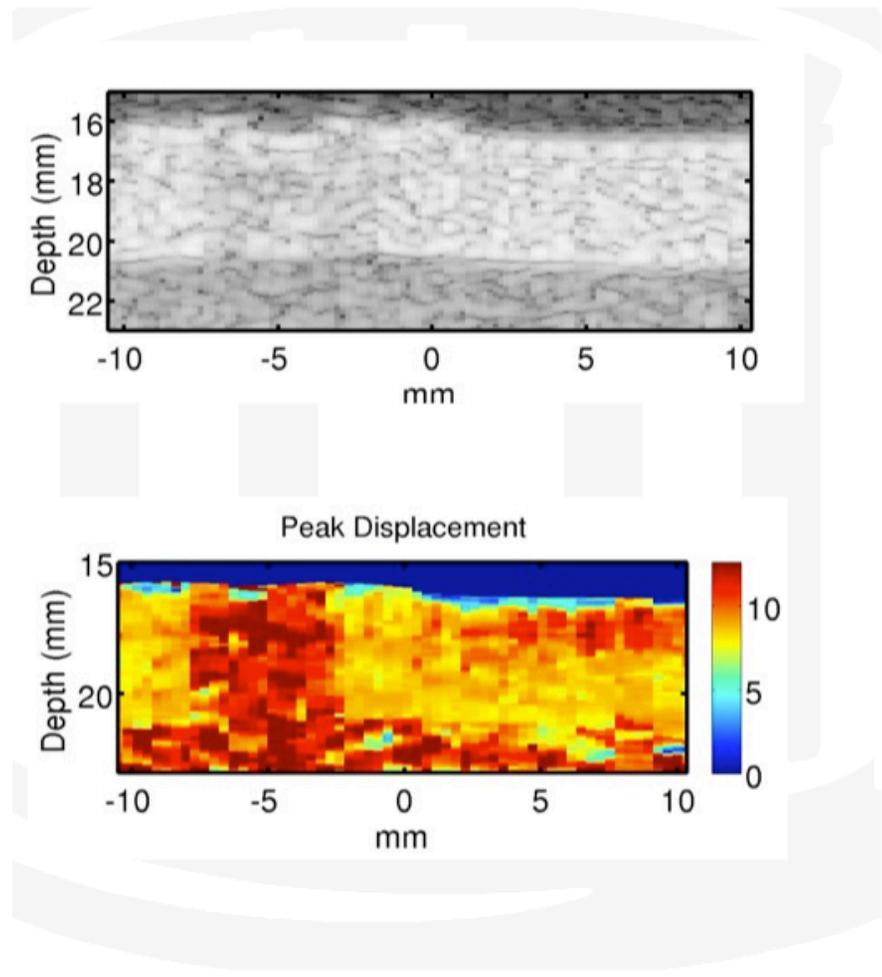
Reader GUI

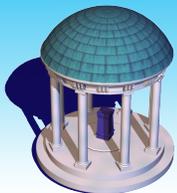




Custom Phantom Structure

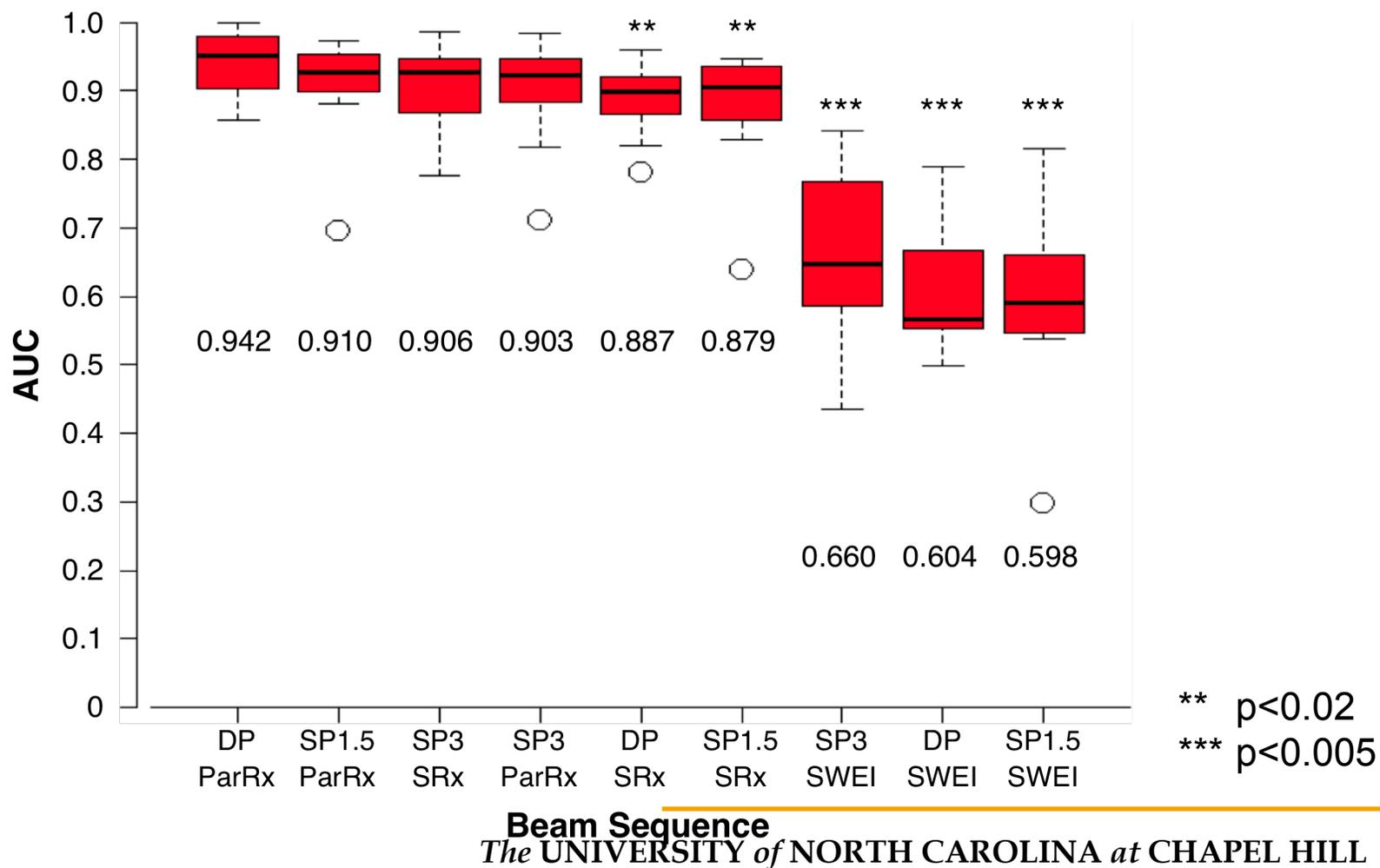
- ~4mm Layer
- Hard or Soft Inclusions
 - ◆ ~110 kPa & ~190 kPa
- 2.5 or 5mm Width
- Imaging in 3 Locations
 - ◆ Centered (0mm Offset)
 - ◆ -3mm Lateral Offset
 - ◆ -6mm Lateral Offset
- 2 Acquisitions
- 250 Total Image Sets

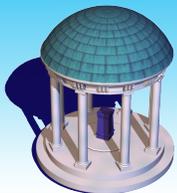




Results: Phantom, All Locations

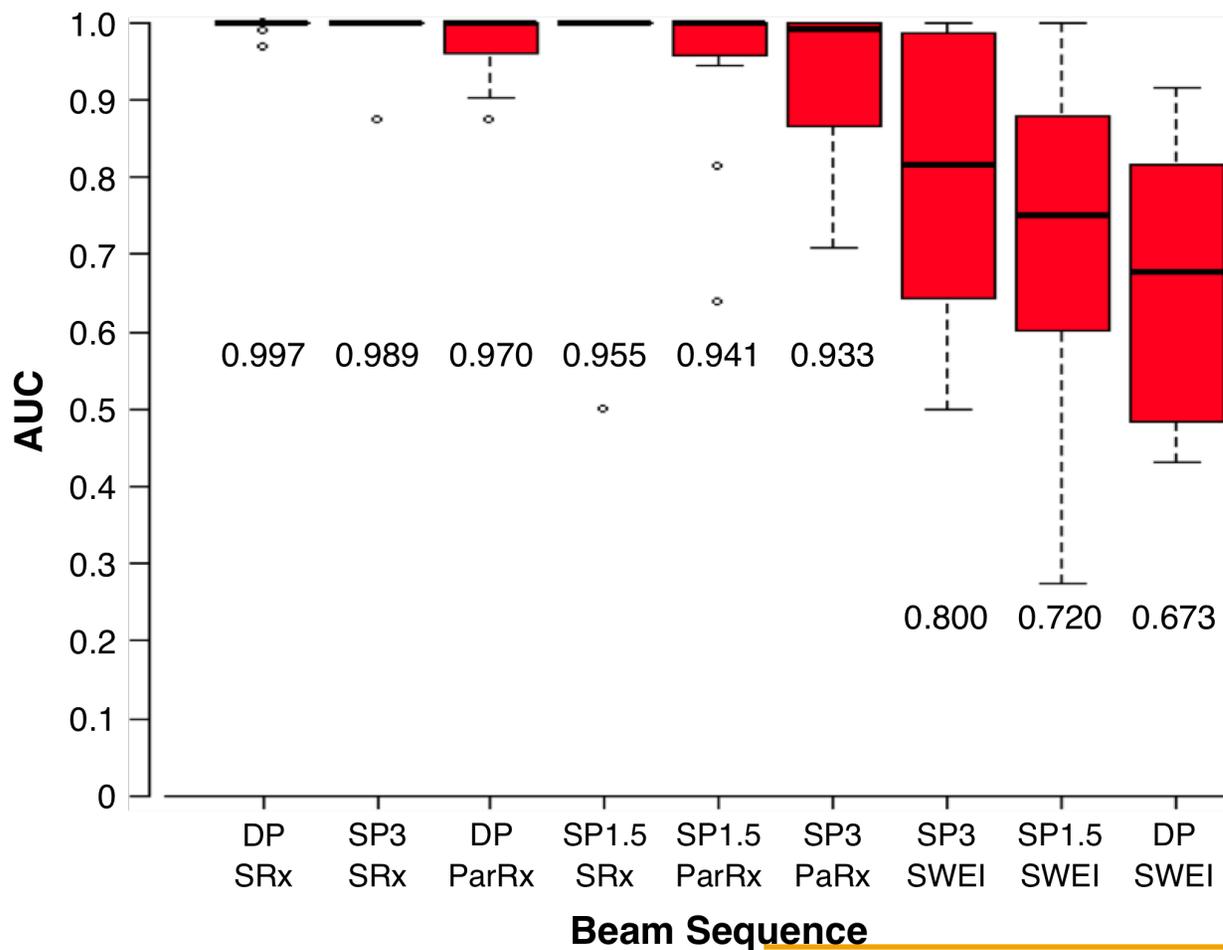
Area Under ROC Curve vs. Beam Sequence

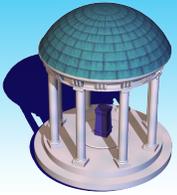




Results: Phantom, -6 mm Lateral Offset

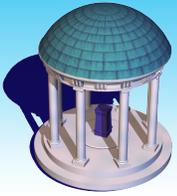
Area Under ROC Curve vs. Beam Sequence





Conclusions

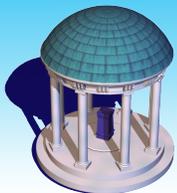
- **Robust Method for Statistically Comparing Beam Sequences**
- **Plaque Detection was better when tracking in ROE**
 - ◆ **Both in Phantoms and Ex Vivo**
 - ◆ **Even when accounting for optimal positioning**
- **Ranking of sequence performance remained consistent between phantom and ex vivo studies**
 - ◆ **SP1.5-SRx**
 - ◆ **SP3-SRx**
 - ◆ **SP3-ParRx**
 - ◆ **DP-SRx**
 - ◆ **DP-ParRx**
 - ◆ **SP3-SWEI**
 - ◆ **SP1.5-SWEI**



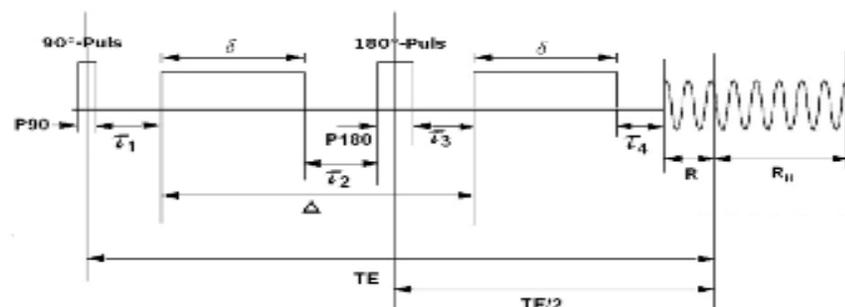
Imaging Sequence **Optimization**

Experimental Design

How to design an **optimal** imaging acquisition scheme to achieve the best signal-to-noise ratio for a given scan time?

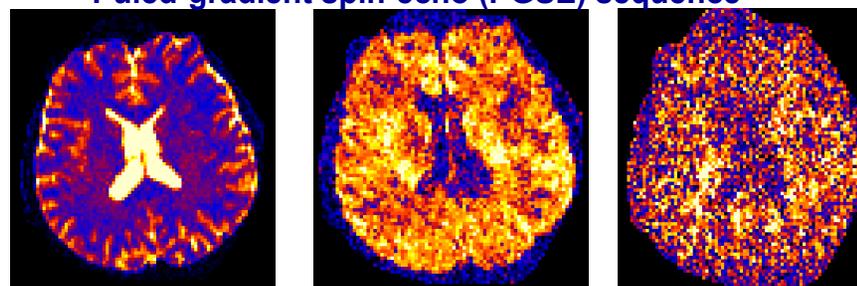


Acquisition Scheme (Imaging Parameters)

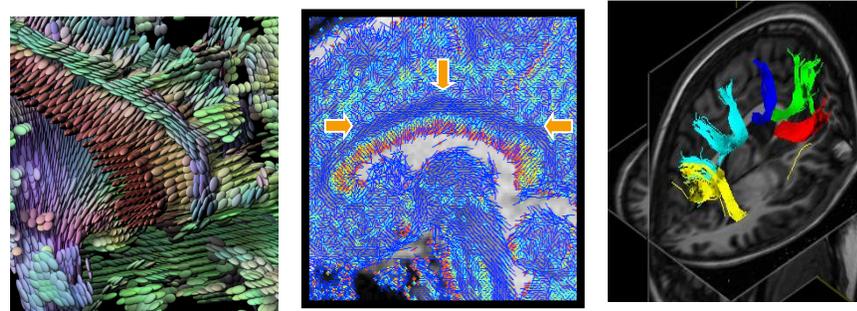


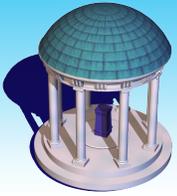
Puled-gradient spin-echo (PGSE) sequence

Noisy Images



Images Reconstruction





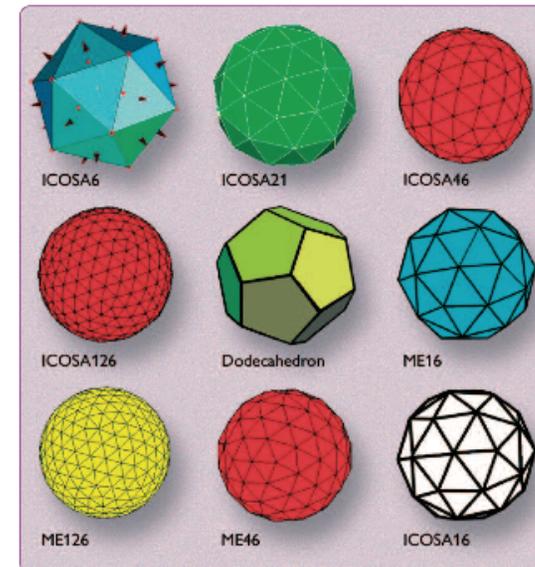
STATISTICAL MODEL

**Gradient Orientations &
b factors**

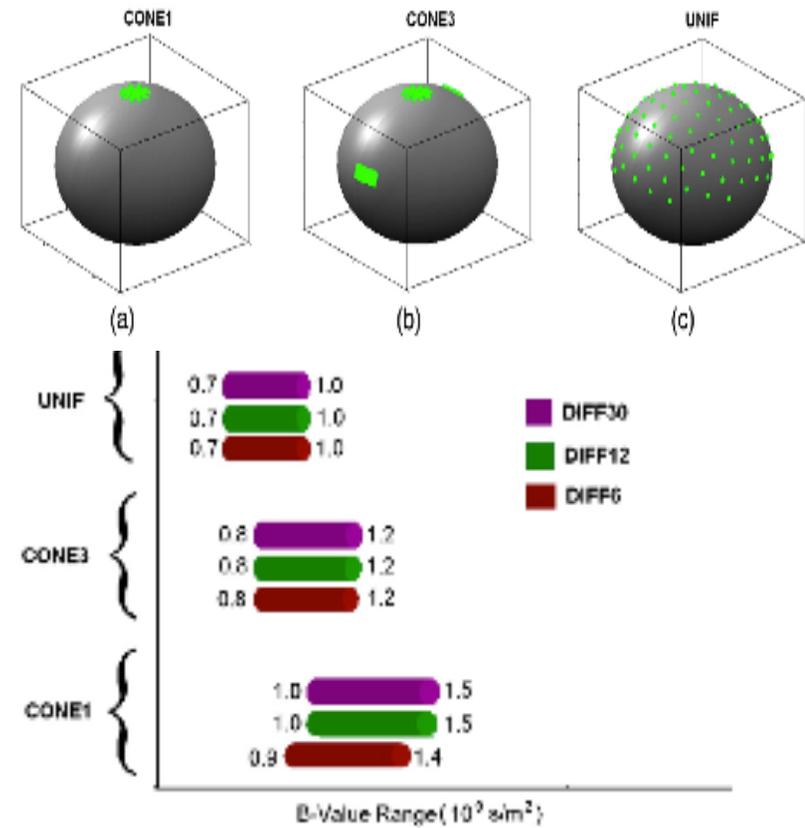
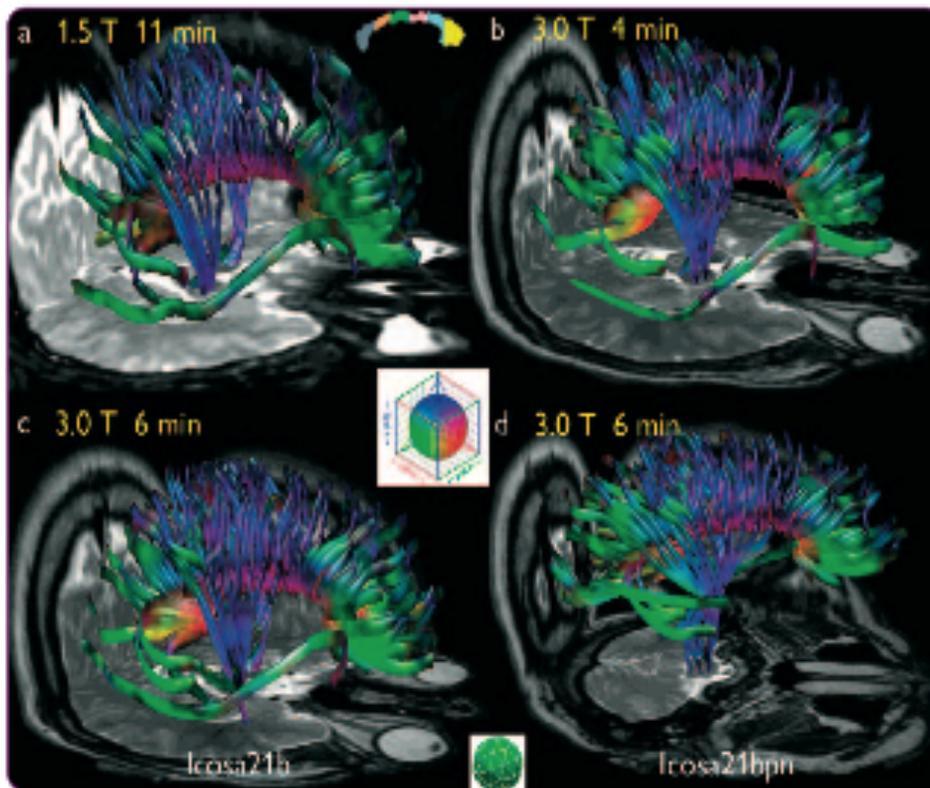
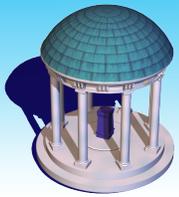
Design Criterion

Global Optimization

$$S \approx S_0 e^{-bg^T Dg} = f(x, \theta)$$
$$p(S, b, g | S_0, D)$$

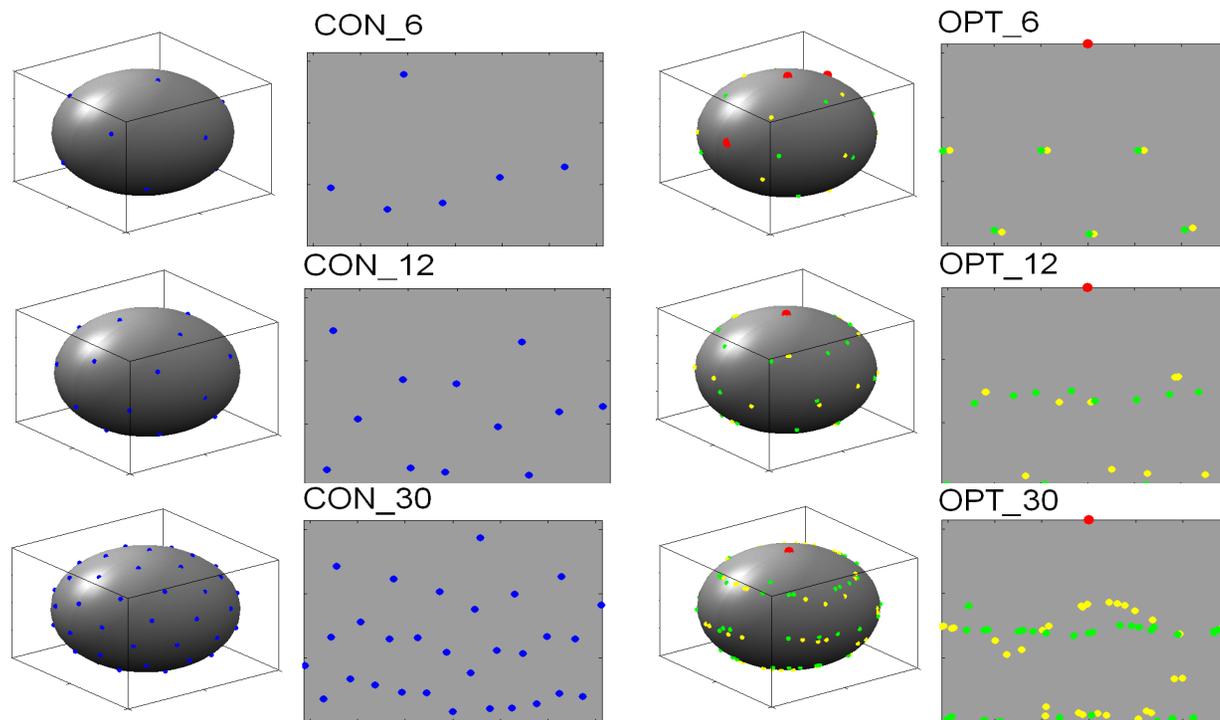
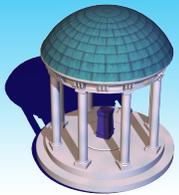


Gradient directions (Hasan & Narayana, 2005, MedicaMundi)



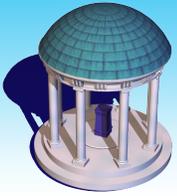
Hasan & Narayana (2005), *MedicaMundi*.

Gao, Zhu, Lin. (2008). *Neuroimage*.



Conventional gradient schemes (a) and optimized schemes (b) based on LS and WLS estimation for uniform fiber case.

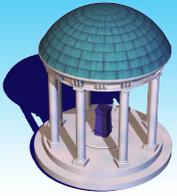
Gao, Zhu, Lin. (2008). *Neuroimage*.



Reconstructing Diffusion Tensor Images

Estimation Theory

How to obtain **accurate** estimates of diffusion tensor and its derived quantities?



Data

$$\{(S_i(v), b_i, g_i) : i = 1, \dots, n; v \in V\}$$

**Rician Regression or
Log-linear Model**

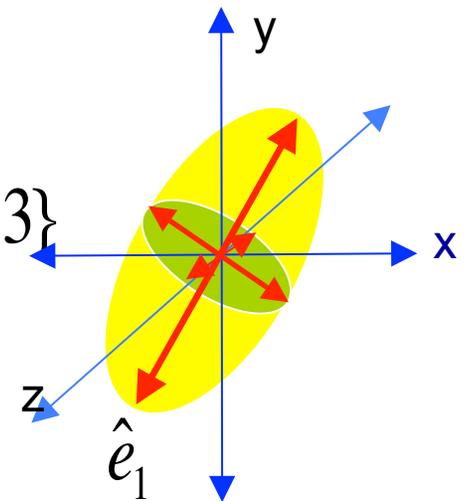
$$S_i(v) = S_0(v) \exp(-b_i g_i^T D(v) g_i) + \text{noise}$$

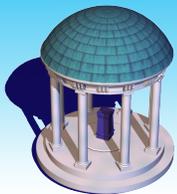
**Estimated Diffusion
Tensor**

$$\hat{D}(v)$$

**Estimated Eigenvalues or
Eigenvectors**

$$\{(\hat{\lambda}_k, \hat{e}_k) : k = 1, 2, 3\}$$





Sorting Bias

$$\hat{D}(v) = \hat{\lambda}_1 \hat{e}_1 \hat{e}_1^T + \hat{\lambda}_2 \hat{e}_2 \hat{e}_2^T + \hat{\lambda}_3 \hat{e}_3 \hat{e}_3^T$$

$$P(\hat{\lambda}_1 > \hat{\lambda}_2 > \hat{\lambda}_3) = 1$$

$$E(\hat{\lambda}_1) > \lambda_1$$

$$E(\hat{\lambda}_3) < \lambda_3$$

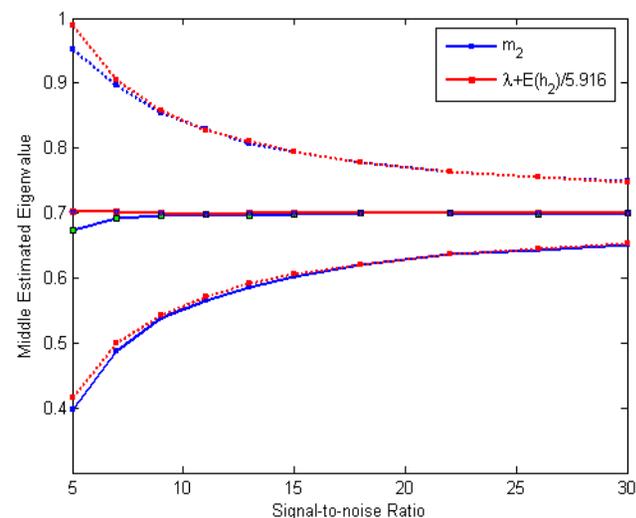
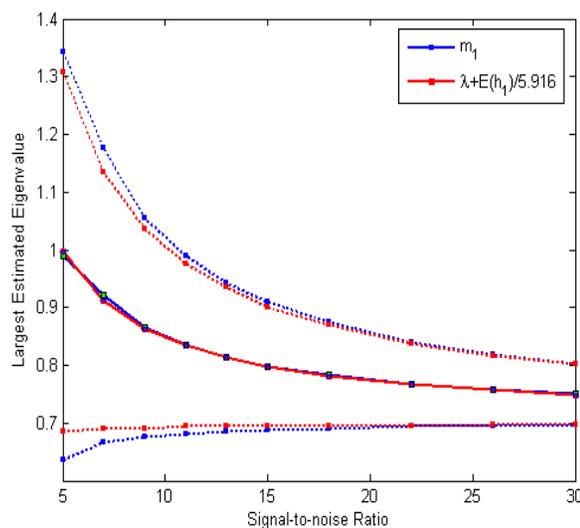
$$E(\hat{\lambda}_2) \approx \lambda_2$$

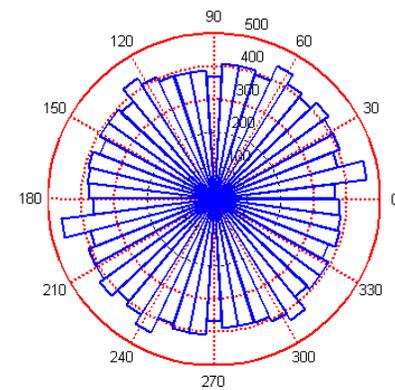
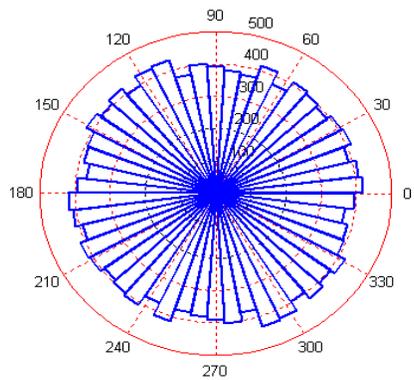
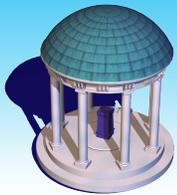
True Diffusion Tensor

$$D(v) = \lambda_1 e_1 e_1^T + \lambda_2 e_2 e_2^T + \lambda_3 e_3 e_3^T$$

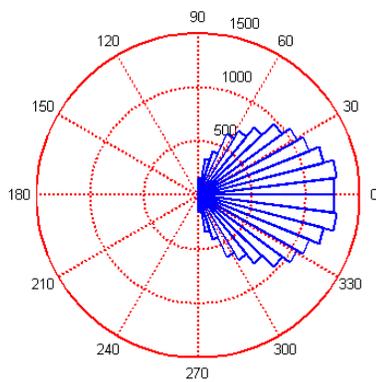
$$\lambda_1 \geq \lambda_2 \geq \lambda_3$$

D=diag(0.7, 0.7, 0.7)

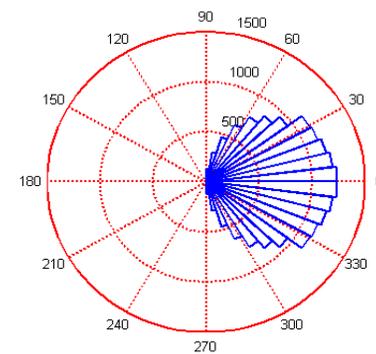


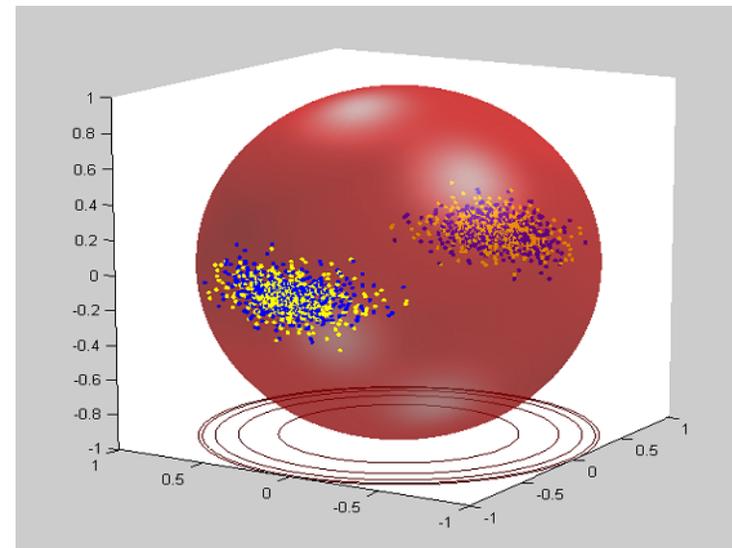
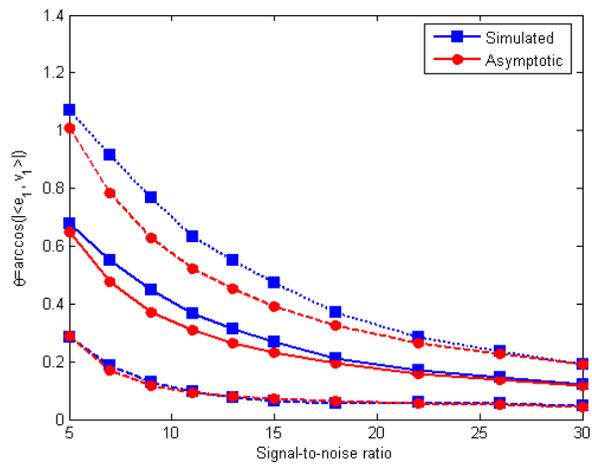
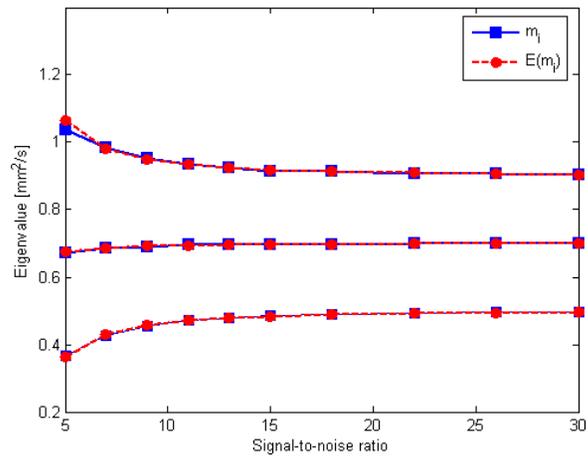
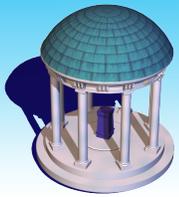


$\theta \in [0, 2\pi]$ and $\phi \in [0, \pi]$ are associated with the spherical coordinate $(1, \theta, \phi)$ of \mathbf{e}_1 .



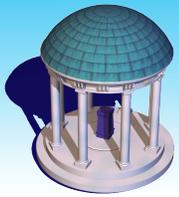
$$SNR = S_0 / \sigma = 22$$



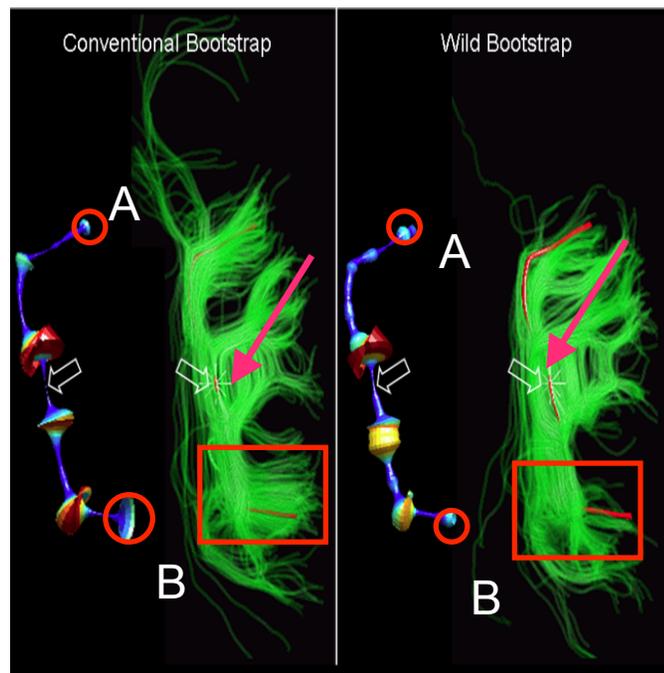


$$SNR = S_0 / \sigma = 22$$

$$D = \text{diag}[0.9, 0.7, 0.5]$$

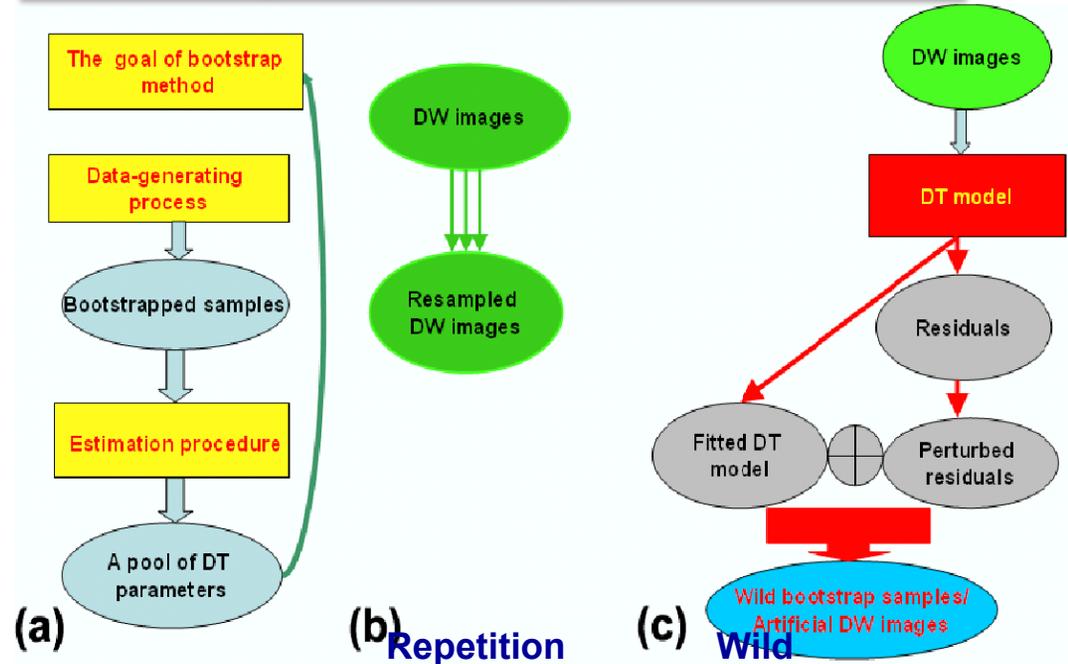


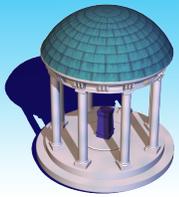
Quantifying Uncertainty by using bootstrap methods



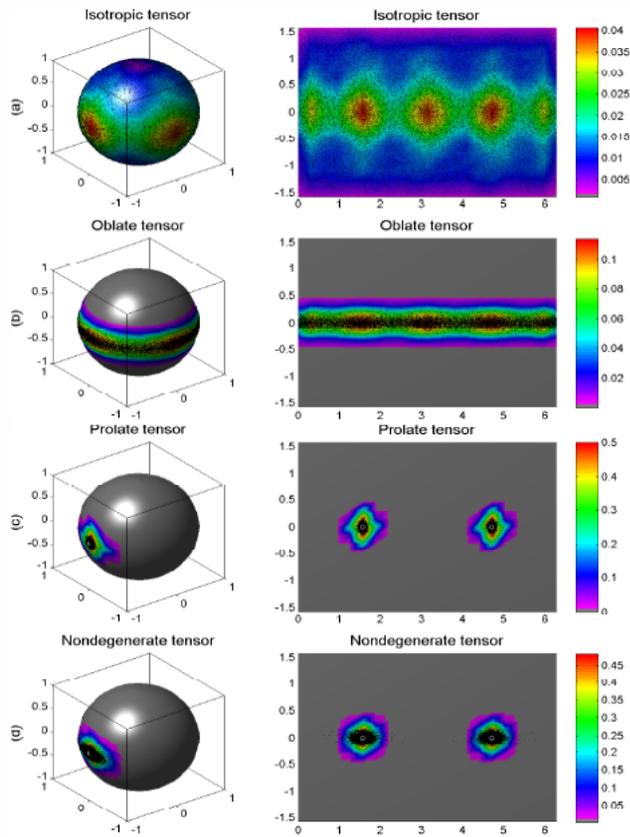
Repetition
Bootstrap
Wild

Yuan, Zhu, Ibrahim, Lin, Peterson. (2008). IEEE TMI

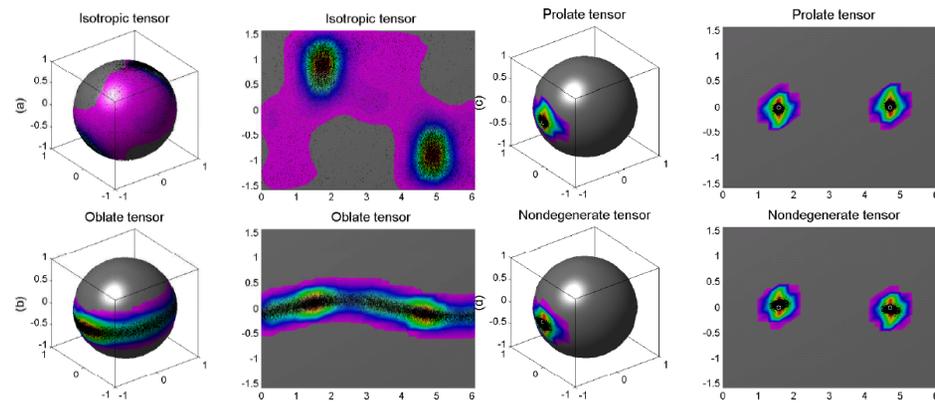




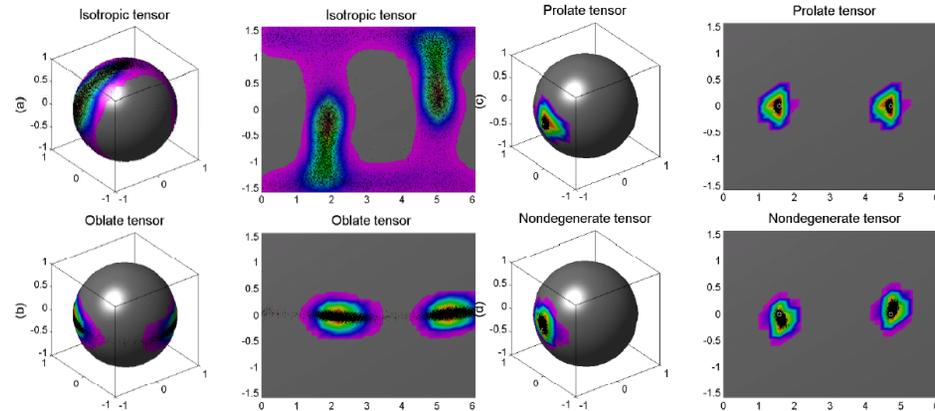
Gold Standard

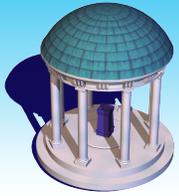


Repetition Bootstrap



Wild Bootstrap

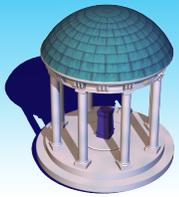




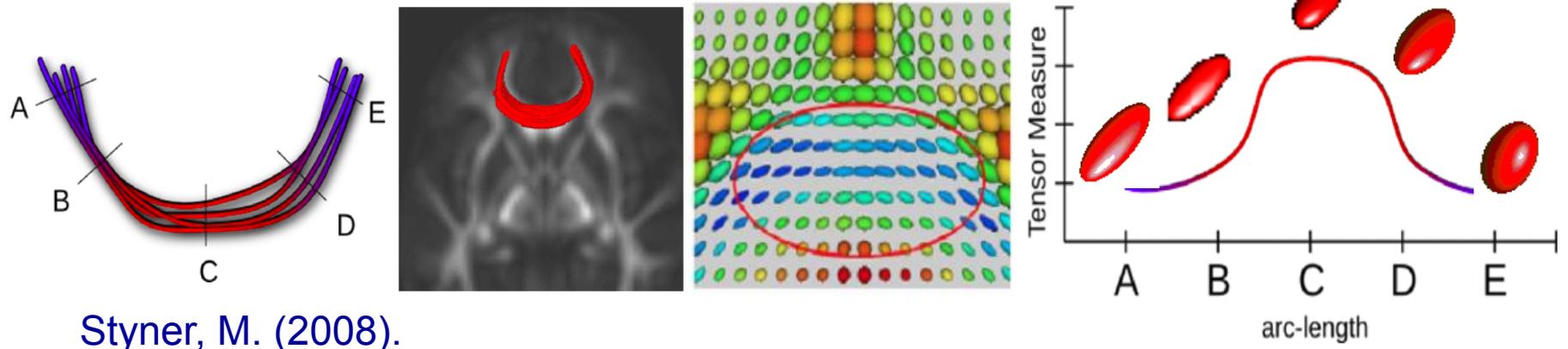
Smoothing Diffusion Tensor Field

Nonparametric Regression

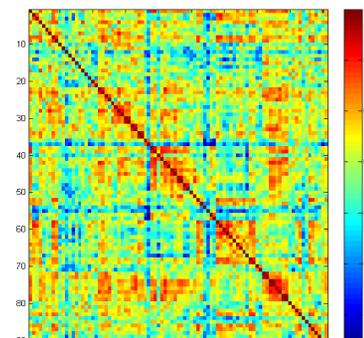
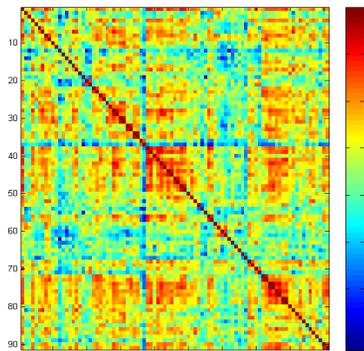
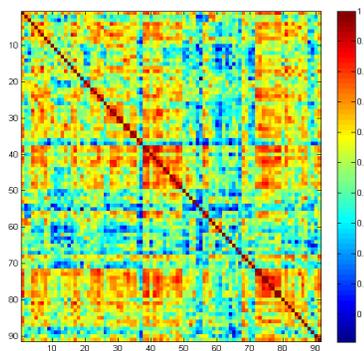
How to **smooth** diffusion tensor
along fiber tracts or in 3D volume?

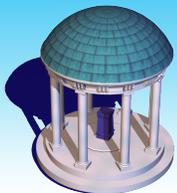


Data $(x_1, S_1), \dots, (x_n, S_n)$



Styner, M. (2008).





$$M = \text{Sym}(m)^+$$

Inner product $\langle\langle Y_D, Z_D \rangle\rangle$

Geodesic

Riemannian exponential/logarithm maps

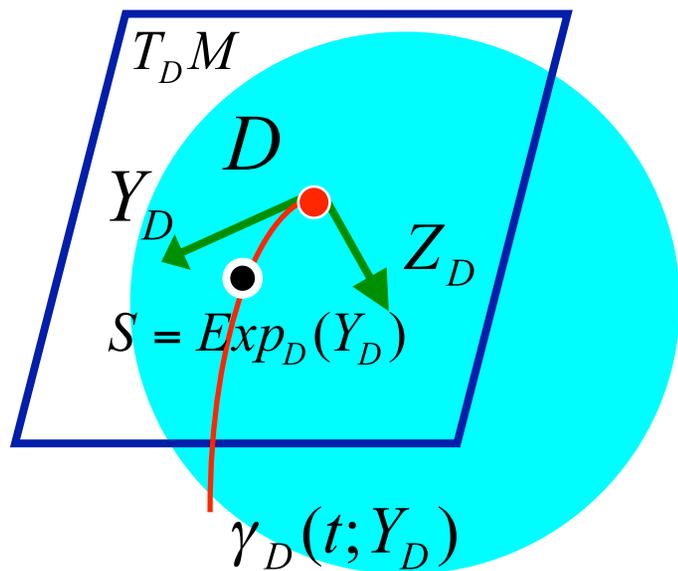
- Affine invariant metric

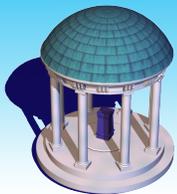
$$\langle\langle Y_D, Z_D \rangle\rangle_{D,R} = \text{tr}(Y_D D^{-1} Z_D D^{-1})$$

- Log-Euclidean metric

$$\langle\langle Y_D, Z_D \rangle\rangle_{D,L} = \text{tr}(R_D(Y_D)R_D(Z_D))$$

$$R_D : T_D M \rightarrow T_{I_m} M$$

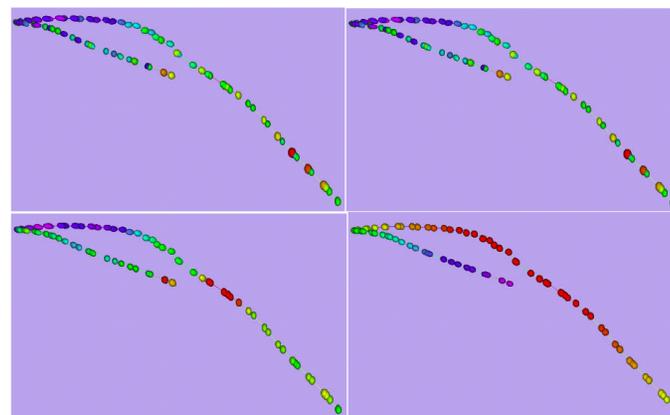
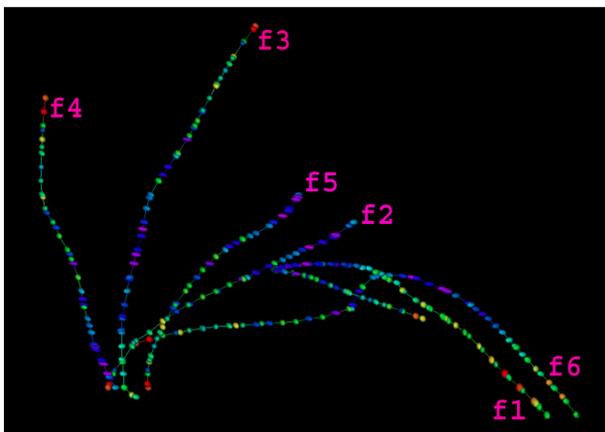


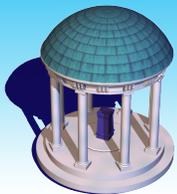


Local polynomial kernel regression to nonparametrically estimate an intrinsic mean of S given x .

Local linear regression performs better than local constant regression.

Statistical inferences depend on a specific inner product defined on the tangent space.





Local Polynomial Kernel Regression

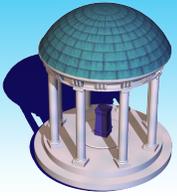
$$\text{Log}_{D(x_0)}(D(x)) \in T_{D(x_0)} \text{Sym}^+(m)$$

$$\phi_{D(x_0)}(\cdot) : T_{D(x_0)} \text{Sym}^+(m) \rightarrow T_{I_m} \text{Sym}^+(m)$$

$$Y(x) = \phi_{D(x_0)}(\text{Log}_{D(x_0)}(D(x)))$$

$$\text{Log}_{D(x_0)}(D(x)) = \phi_{D(x_0)}^{-1}(Y(x)) \approx \phi_{D(x_0)}^{-1}(Y(x_0) + \sum_{k=1}^K Y^{(k)}(x_0)(x - x_0)^k)$$

$$D(x) = \text{Exp}_{D(x_0)}(\phi_{D(x_0)}^{-1}(Y(x))) \approx \text{Exp}_{D(x_0)}(\phi_{D(x_0)}^{-1}(\sum_{k=1}^K Y^{(k)}(x_0)(x - x_0)^k))$$



Simulation Studies

Data model

$$S_i = C(x_i) \exp(E_i) C(x_i), \quad \begin{array}{l} E_i \sim MN(0, \Omega) \\ x_i \sim N(0, 0.25) \end{array}$$
$$D(x) = C(x)^2$$

$$C(x) = \begin{pmatrix} -0.1x & 0.2x & \sin(x) \\ 0.2x & 0.6x & -0.4x \\ \sin(x) & -0.4x & 0.5x \end{pmatrix}$$

Correlation

$$\Sigma_1 = \begin{pmatrix} 0.3 & 0.049 & 0.052 \\ 0.049 & 0.2 & 0.0424 \\ 0.052 & 0.0424 & 0.1 \end{pmatrix} \quad \Sigma_2 = 2\Sigma_1, \quad \Sigma_3 = 4\Sigma_1, \quad \Sigma_4 = 8\Sigma_1$$

Data $\{(x_i, S_i) : i = 1, \dots, n\}$ for $n = 50$ or 100

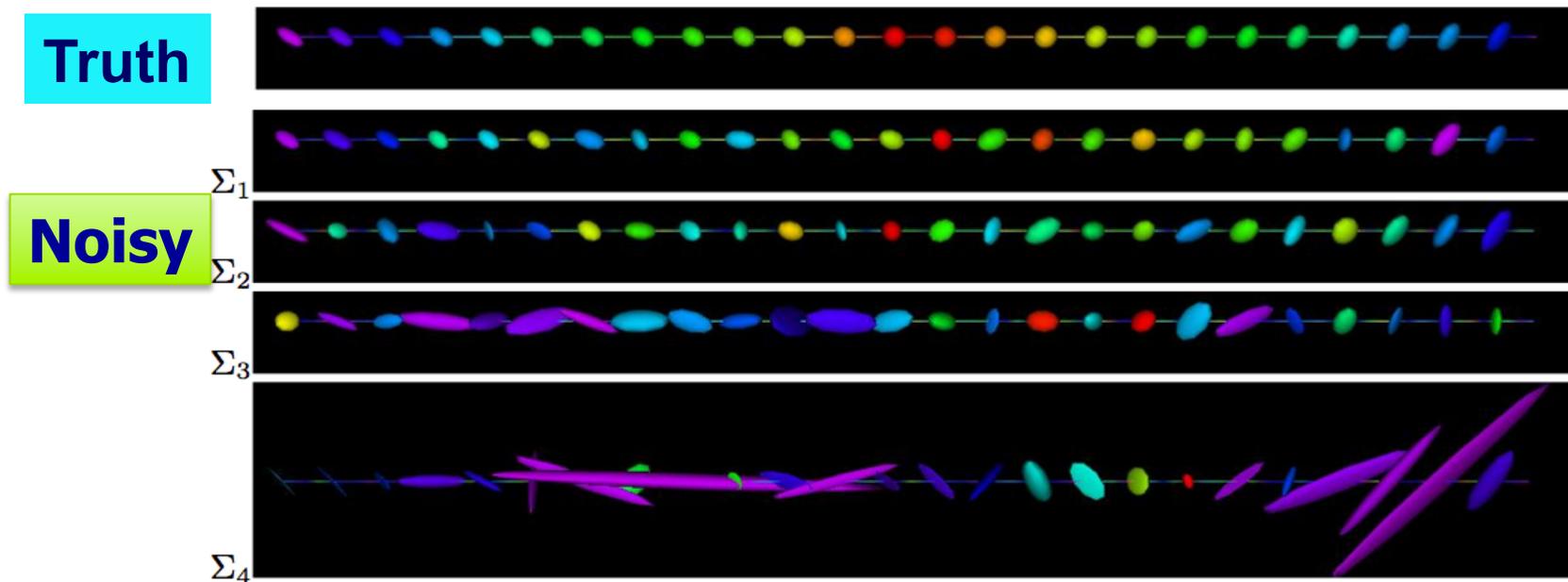
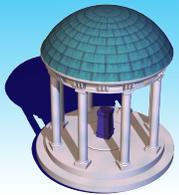
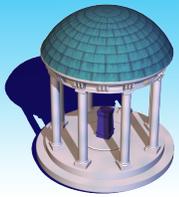


Fig. 1. Ellipsoidal representations of the true (the first row) and simulated SPD matrix data along the design points under the four different noise distributions (the second to the fifth rows: Σ_1 - Σ_4) colored with FA values.



Truth

Log-Euclidean
Local Linear

Riemannian
Local Linear

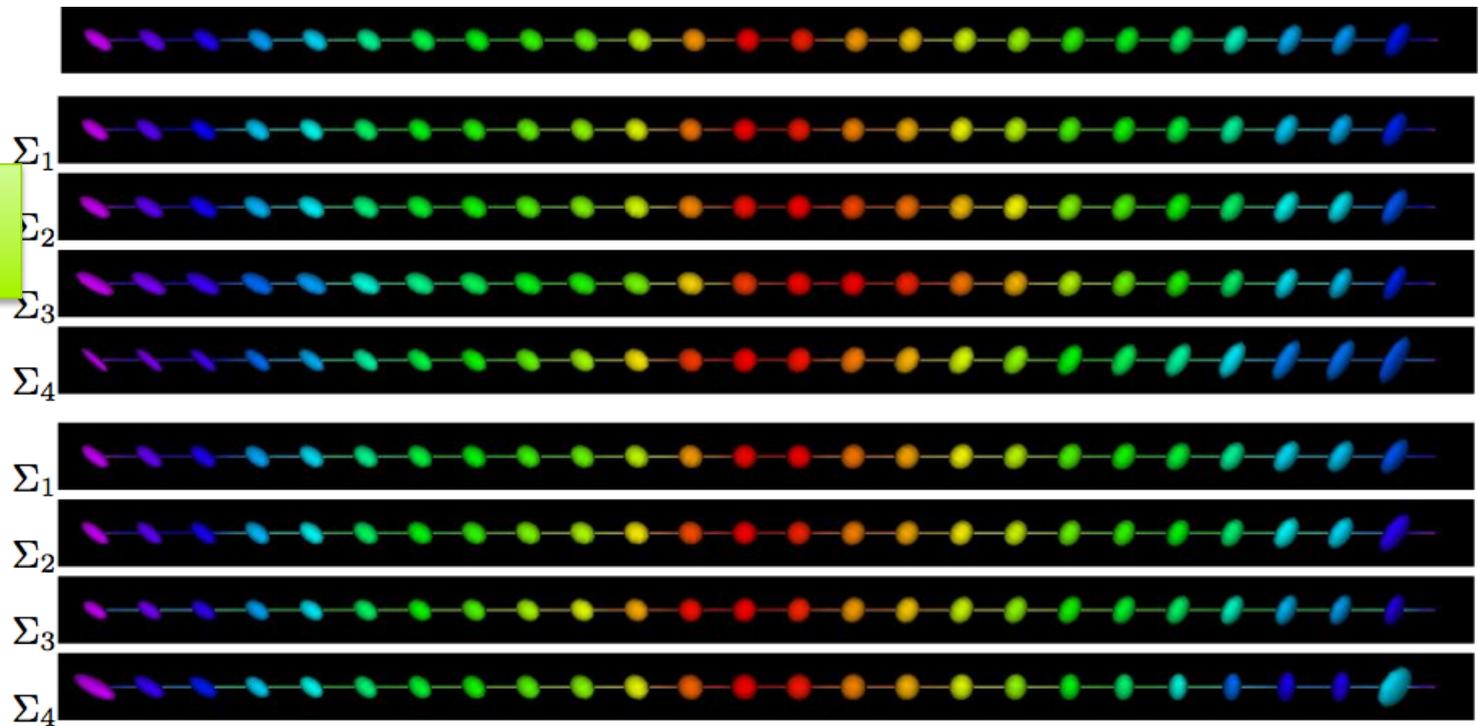
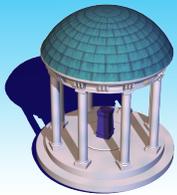


Fig. 2. Ellipsoidal representations of the true (the first row) and estimated SPD matrix data along the design points under the four different noise levels colored with FA values. The second to the fifth rows (Log-Euclidean metric): Σ_1 - Σ_4 , the sixth to the ninth rows (the Riemannian metric): Σ_1 - Σ_4 .



Simulation 1.

- Compare the performance of the **local linear** with the **local constant**
- Assess the performance using the Average Geodesic Distance (AGD) for each replication $j=1, \dots, N$ with N as the number of replications, denoted by

$$\text{AGD} = (nN)^{-1} \sum_{j=1}^N \sum_{i=1}^n d(\hat{D}_j(x_i), D(x_i))$$

where $\hat{D}_j(x_i)$ and $D(x_i)$ are, respectively, the estimated and true diffusion tensors at x_i

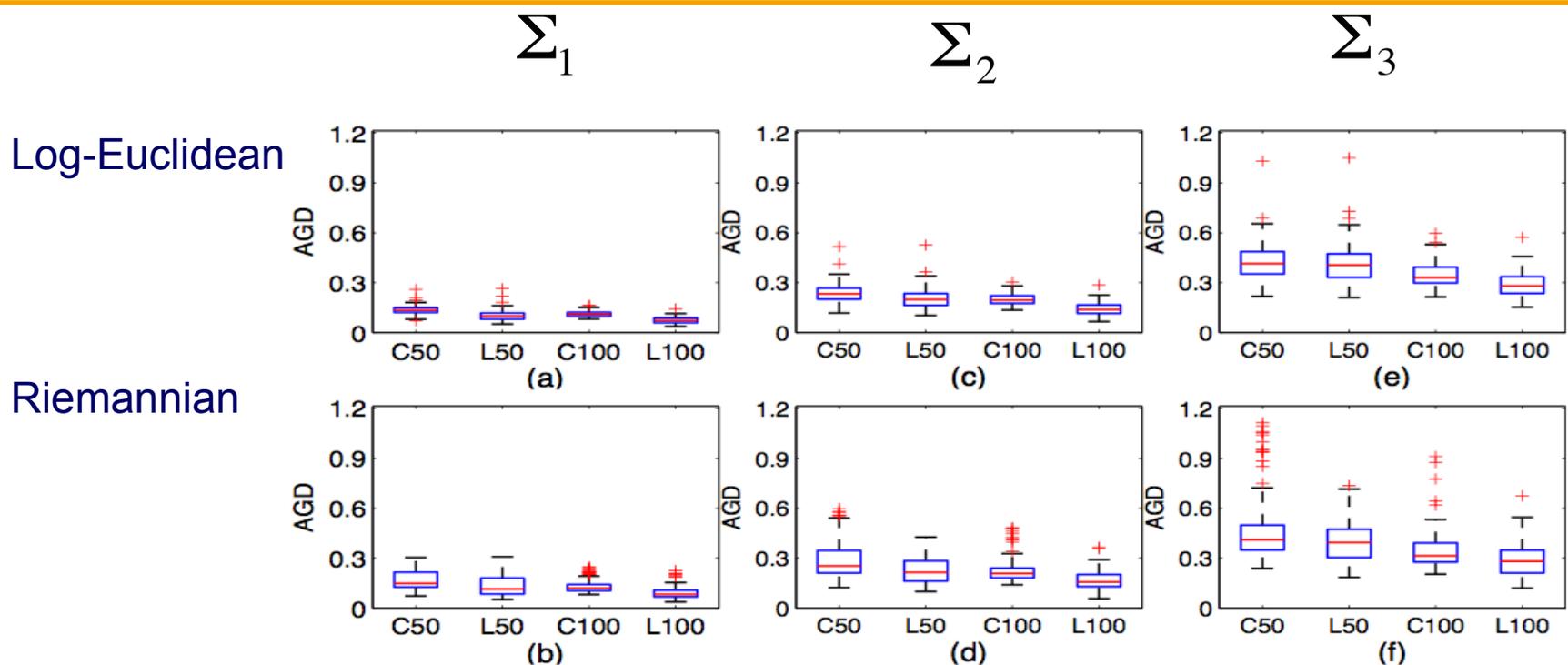
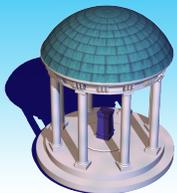


Fig. 3. Boxplots of the AGD using the intrinsic local constant and linear estimators under the log-Euclidean (the first row) and Riemannian (the second row) metrics based on 100 replications under the three covariance matrices (a)-(b) Σ_1 , (c)-(d) Σ_2 , and (e)-(f) Σ_3 . C50 and C100 represent the intrinsic local constant estimators at sample sizes 50 and 100, respectively. L50 and L100 represent the intrinsic local linear estimators at sample sizes 50 and 100, respectively.

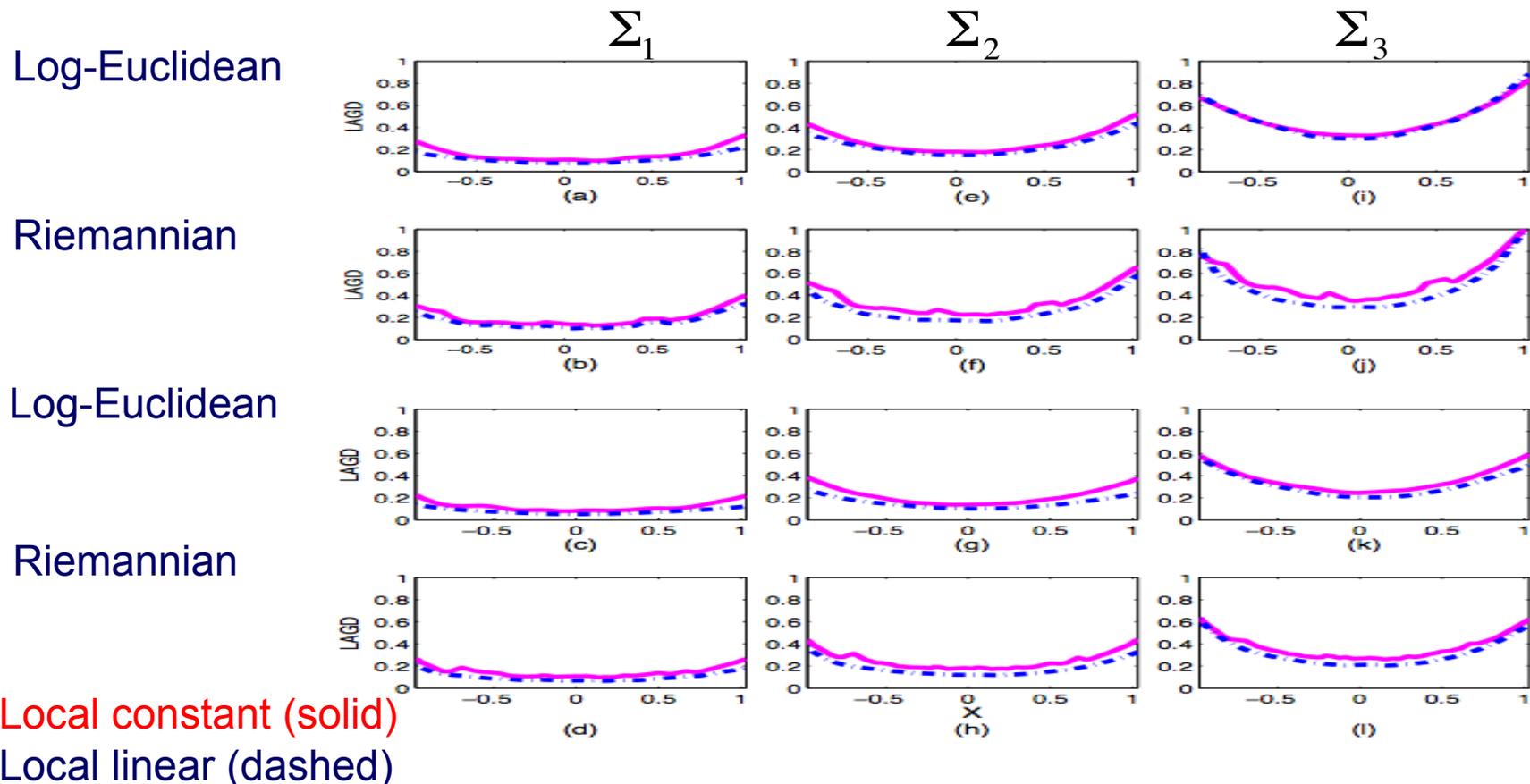
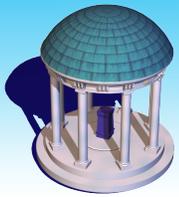
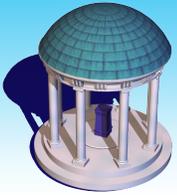


Fig. 4. The LAGD curves at each sample point using the intrinsic local constant (solid line) and linear (dash-dotted line) estimators under the three covariance matrices (a)-(d) Σ_1 , (e)-(h) Σ_2 , (i)-(l) Σ_3 for sample sizes 50 (the top two rows) and 100 (the bottom two rows). The first and third rows correspond to the log-Euclidean metric while the second and fourth rows correspond to the Riemannian metric.



Simulation 2. High noisy level

Compare the performance of the local linear under two metrics

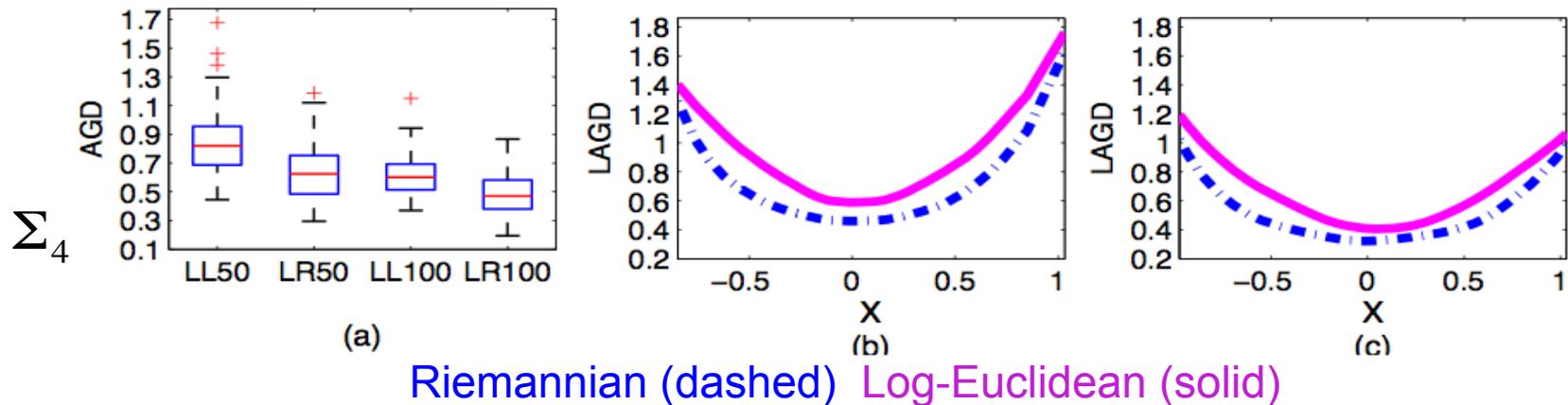
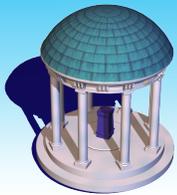


Fig. 5. (a) Boxplots of the AGD's using the linear regressions based on 100 replications under the covariance matrix Σ_4 , under the Log-Euclidean and Riemannian metrics, respectively. (b) and (c) LAGD curves at each sample point using the local linear regressions under the affine invariant (dash-dotted line) and Log-Euclidean (solid line) metrics under the the covariance matrix Σ_4 at sample size 50 (b) and 100 (c), respectively. LL50 (LR50) and LL100 (LR100), respectively, represent the local linear regressions under Log-Euclidean (Riemannian) metrics at sample sizes 50 and 100.



Simulation 3.

- Value of developing the LPK smoothing method
- Two different methods for smoothing FA values

M1. Calculate FA values from `noisy' SPDs and then use the local linear method to smooth the FA values

M2. Use the local linear method to smooth SPDs and then calculate FA values from the smoothed SPDs

- Calculate the Mean Absolute Deviation Error (MADE):

$$\text{MADE} = (nN)^{-1} \sum_{j=1}^N \sum_{i=1}^n |\hat{\text{FA}}_j(x_i) - \text{FA}_j(x_i)|$$

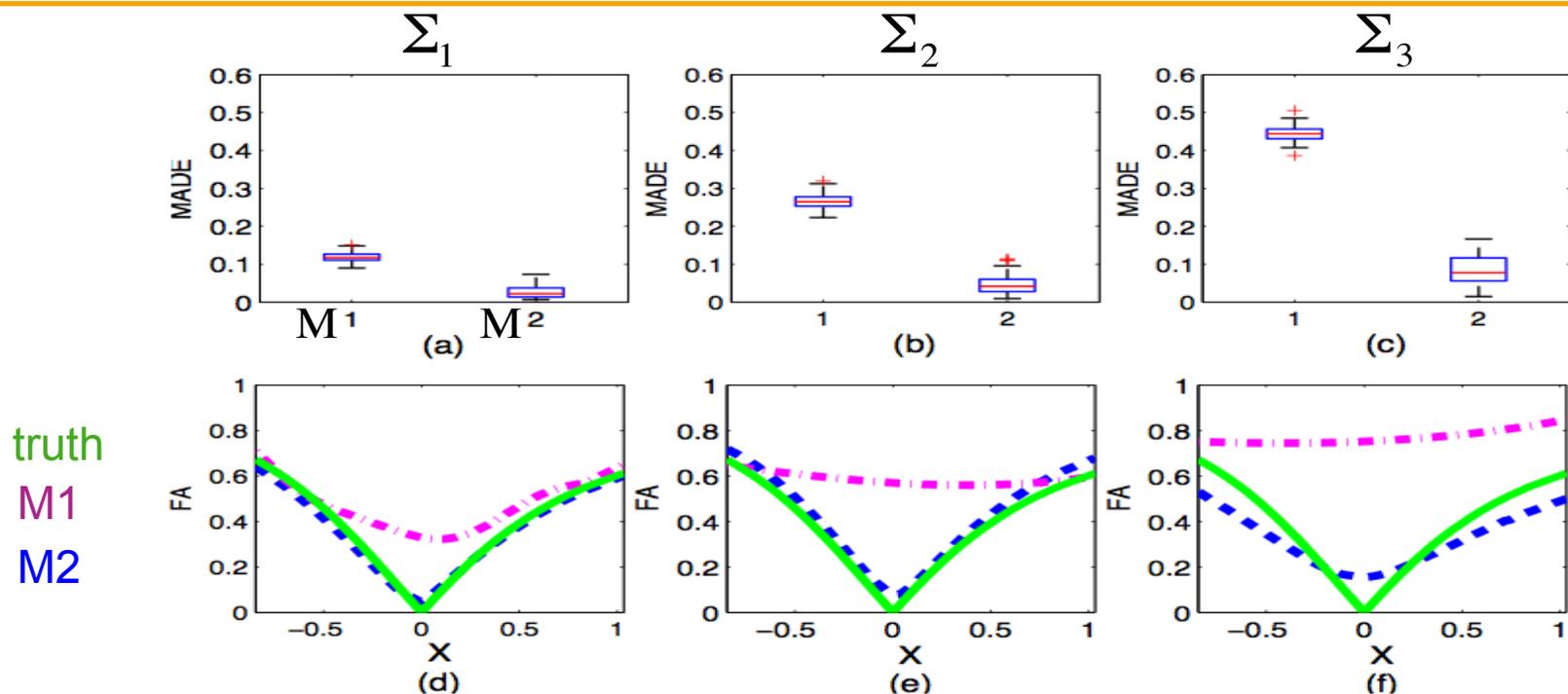
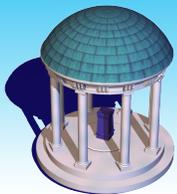
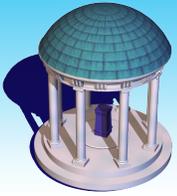


Fig. 6. Boxplot of the MADE's using the two smoothing methods based on 100 replications under the covariance matrices (a) Σ_1 , (b) Σ_2 , and (c) Σ_3 at sample size 50. Smoothed FA curves for the realizations with median MADE under the covariance matrices: (d) Σ_1 , (e) Σ_2 , and (f) Σ_3 . The true FA curve (the solid line), the estimated FA curve using the first method (the dash-dotted line) and the estimated FA curve using the second method (the dashed line). This shows that the more intrinsic approach is much better.



Smoothing DTs along a select tract

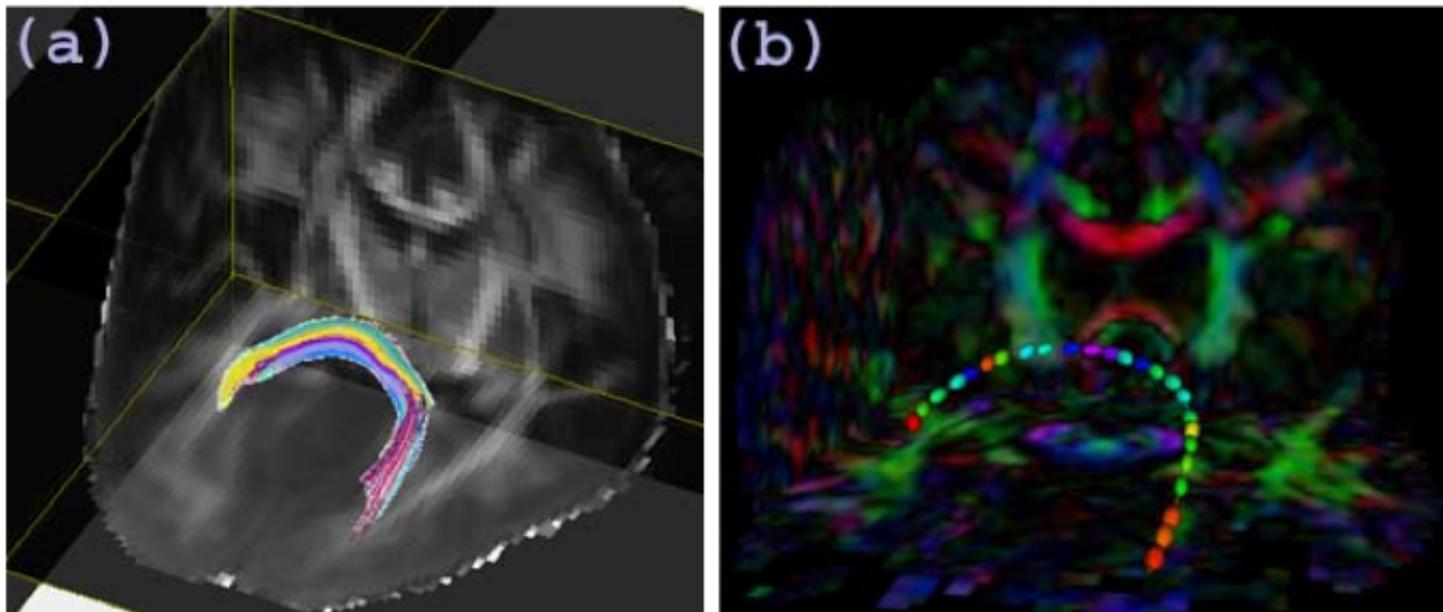


Fig. 7. (a) The splenium of the corpus callosum in the analysis of HIV DTI data. (b) The ellipsoidal representation of full tensors on the fiber tract from a selected subject.

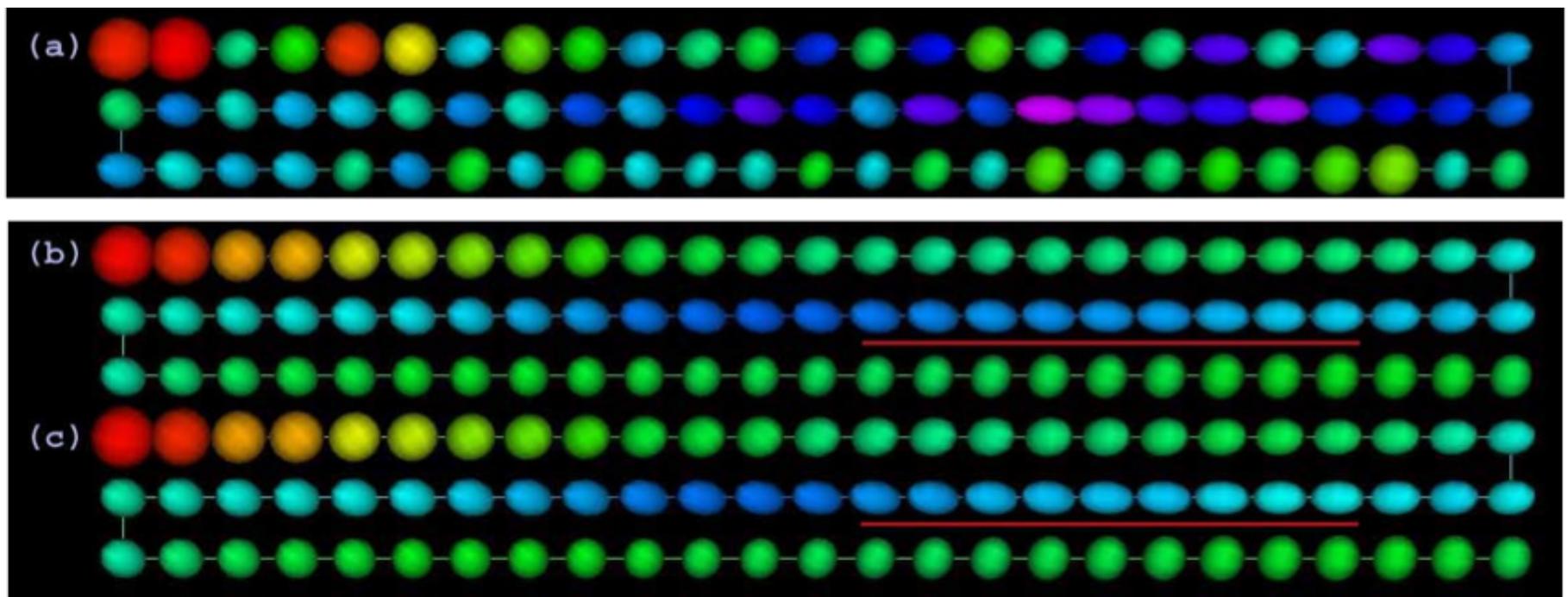
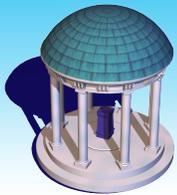
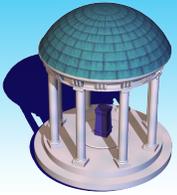


Fig. 8. (a) Ellipsoidal representations of the diffusion tensor data and estimated tensors using the intrinsic local linear regression under the (b)log-Euclidean and (c) Riemannian metrics along the fiber tract f1 colored with FA values. The estimated tensors in the middle right part (highlighted in the red line) are more anisotropic using the method under the Log-Euclidean metric.



Local Polynomial Kernel Regression for SPD

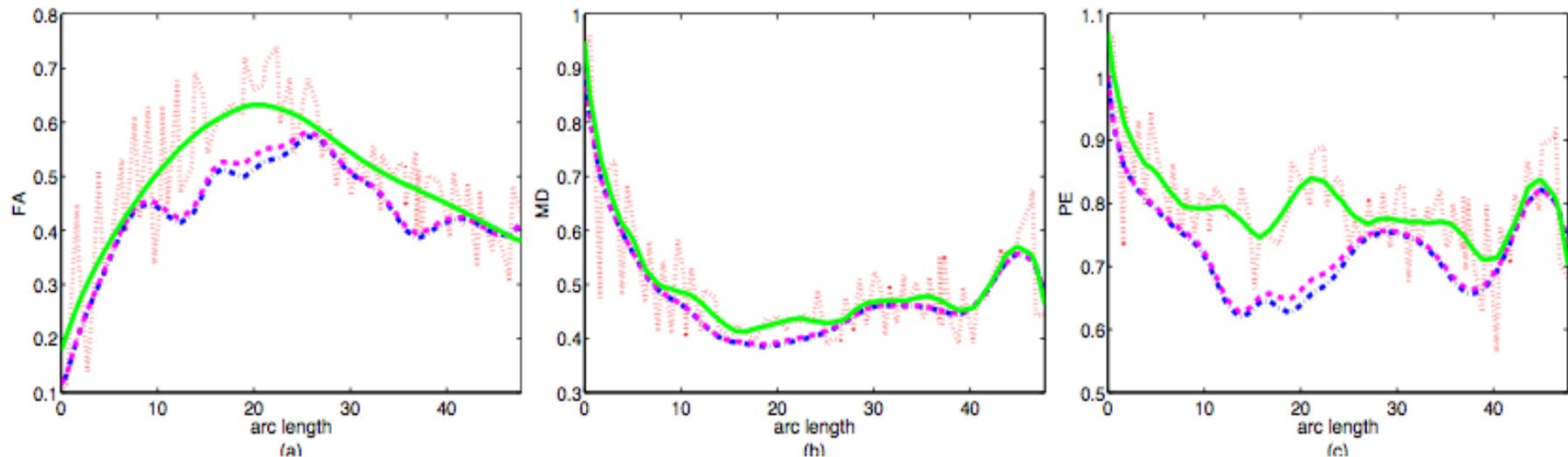
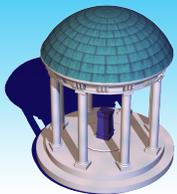


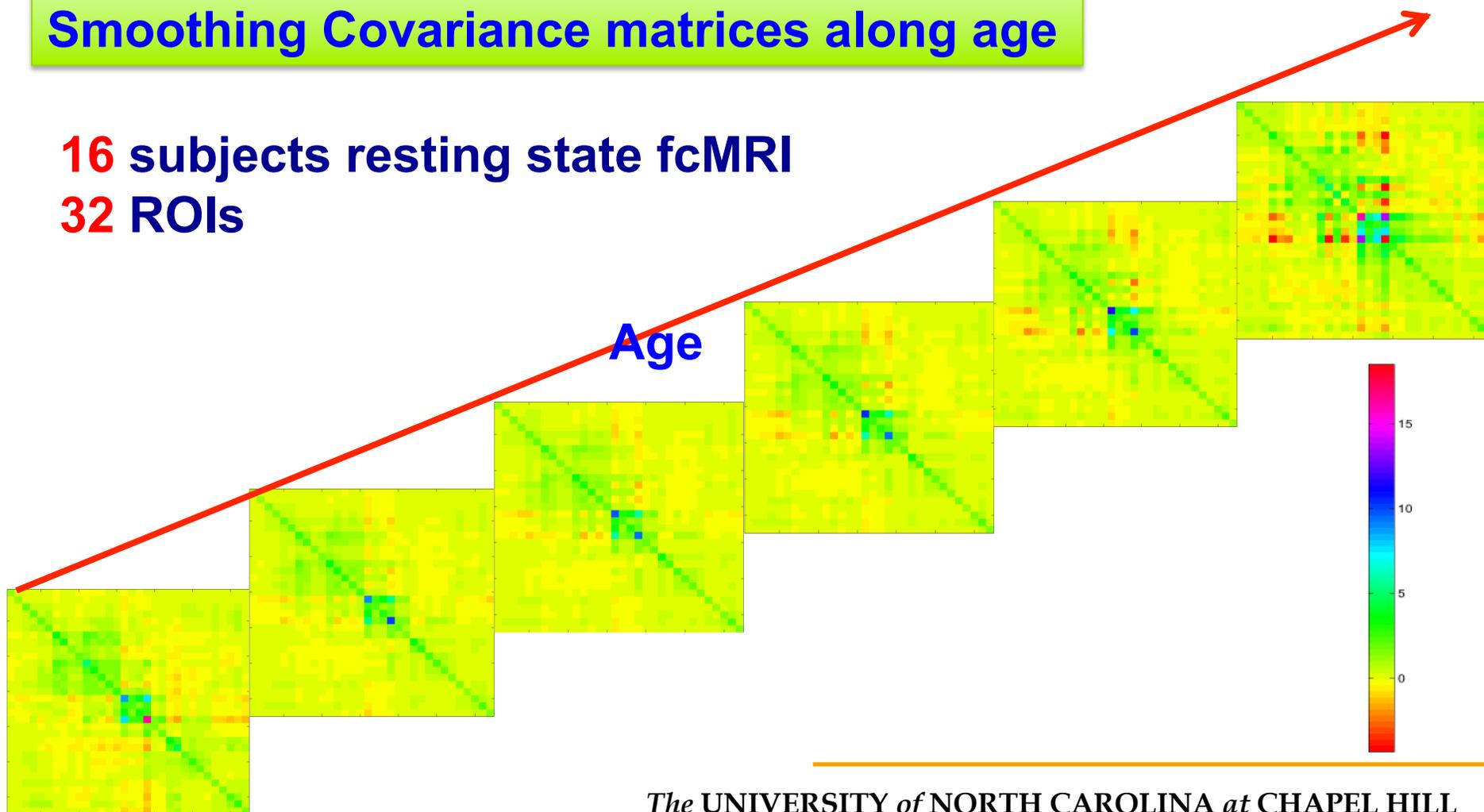
Fig. 9. (a) FA's , (b) MD's and (c) PE's derived from the raw tensor data (dot line) and estimated tensors using the intrinsic local linear regression under the Riemannian (dash-dot line) and log-Euclidean (dash line) metrics as the function of arc-length along the tract f1. Estimated FA function along the fiber tract f1 by using the standard local linear regression for scalars (solid line).

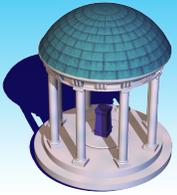


Smoothing Covariance matrices along age

16 subjects resting state fcMRI

32 ROIs

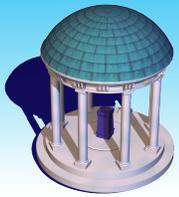




Analyzing Tract-based Diffusion Tensor Statistics

Multivariate Varying Coefficient Model

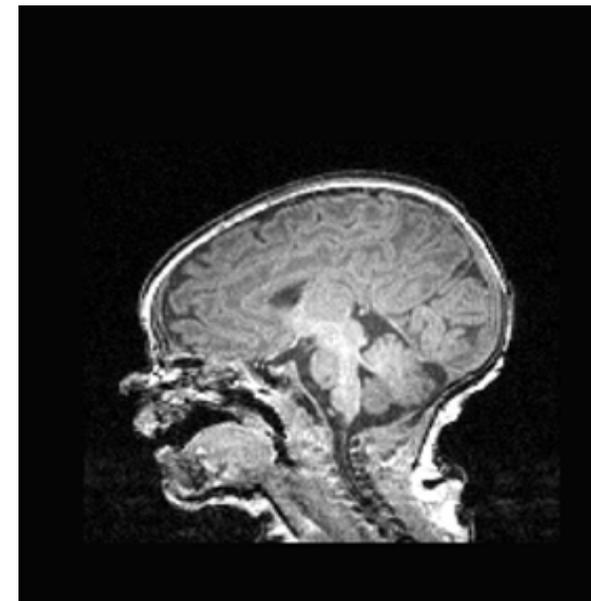
How to **compare** diffusion tensors or tensor quantities along fiber tracts?



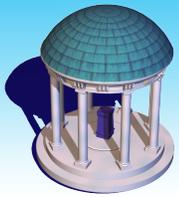
Neonatal Brain Development



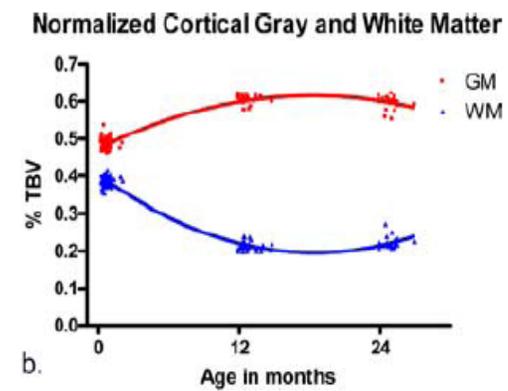
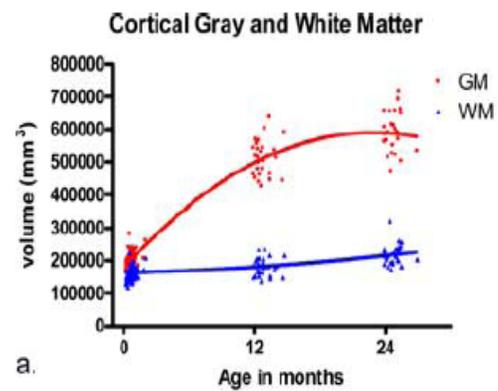
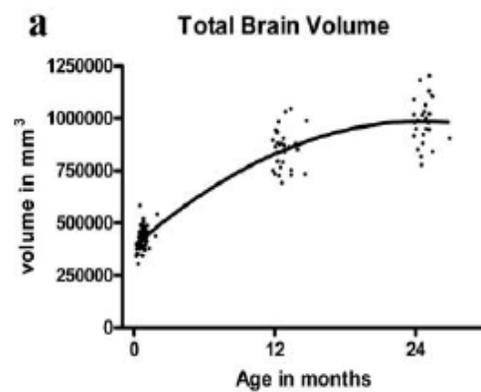
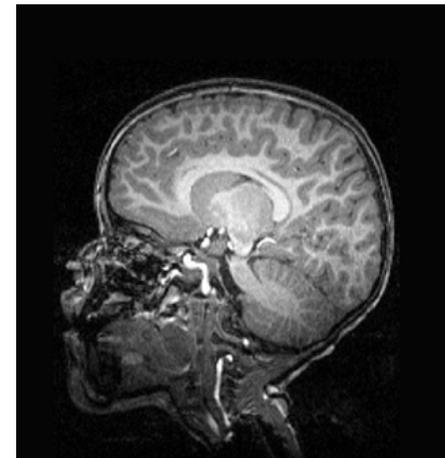
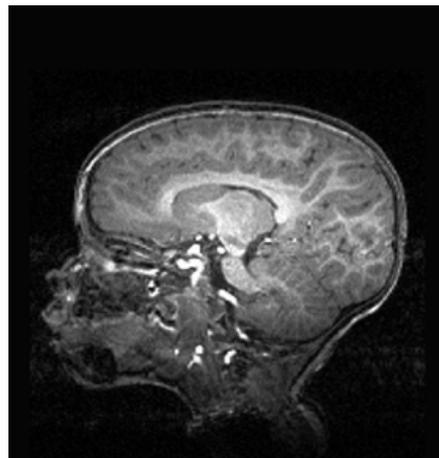
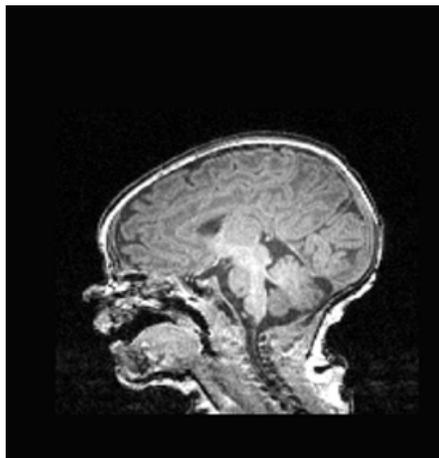
PI: John H. Gilmore

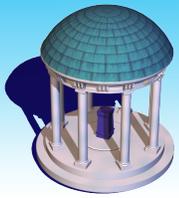


www.google.com

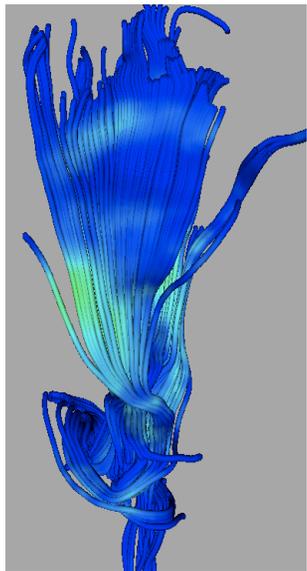


Early Brain Development

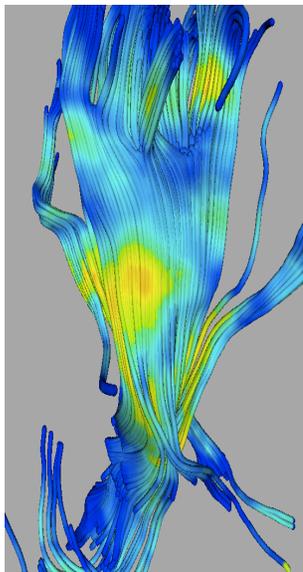




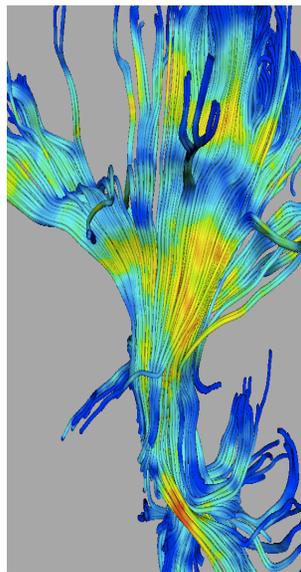
White Matter Maturation



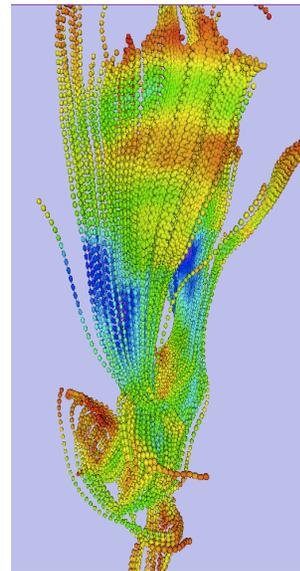
Week 2



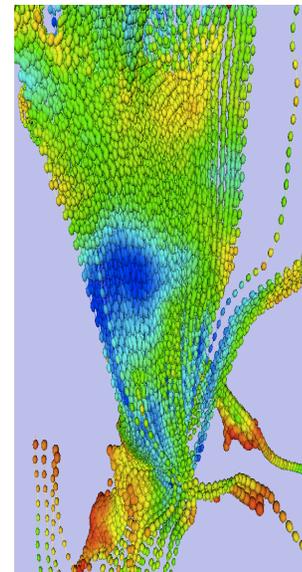
Year 1



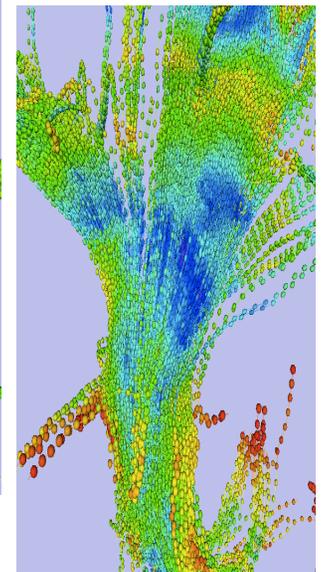
Year 2



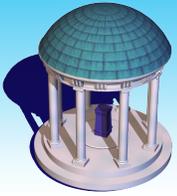
Week 2



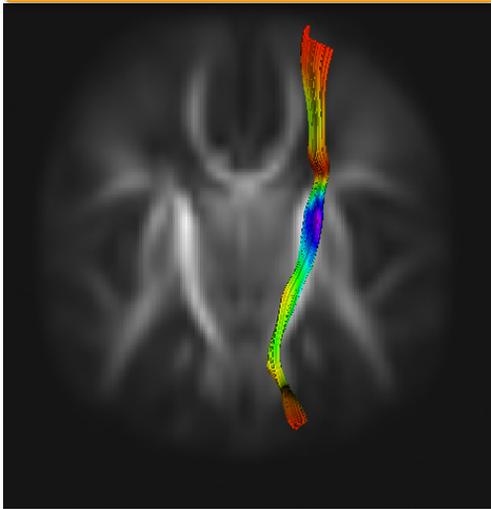
Year 1



Year 2



Sample Data



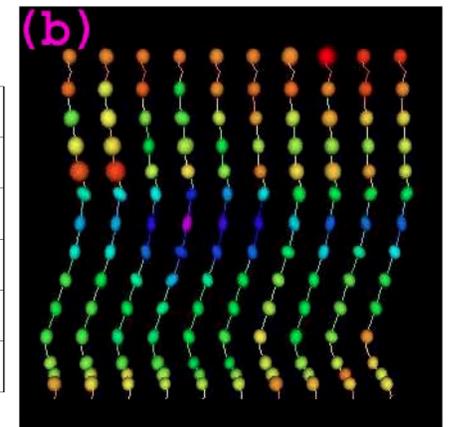
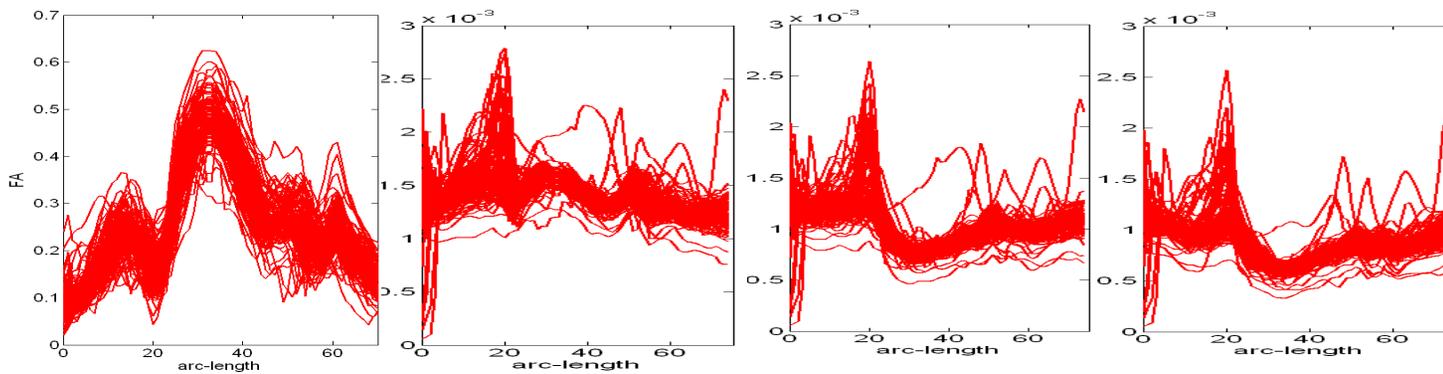
Right internal capsule: a collection of axons connecting the cerebral cortex and the brain stem

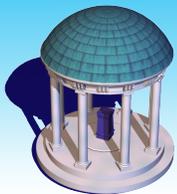
diffusion properties or diffusion tensors

$$Y_i(s_j) = (y_{i,1}(s_j), \dots, y_{i,m}(s_j))^T$$

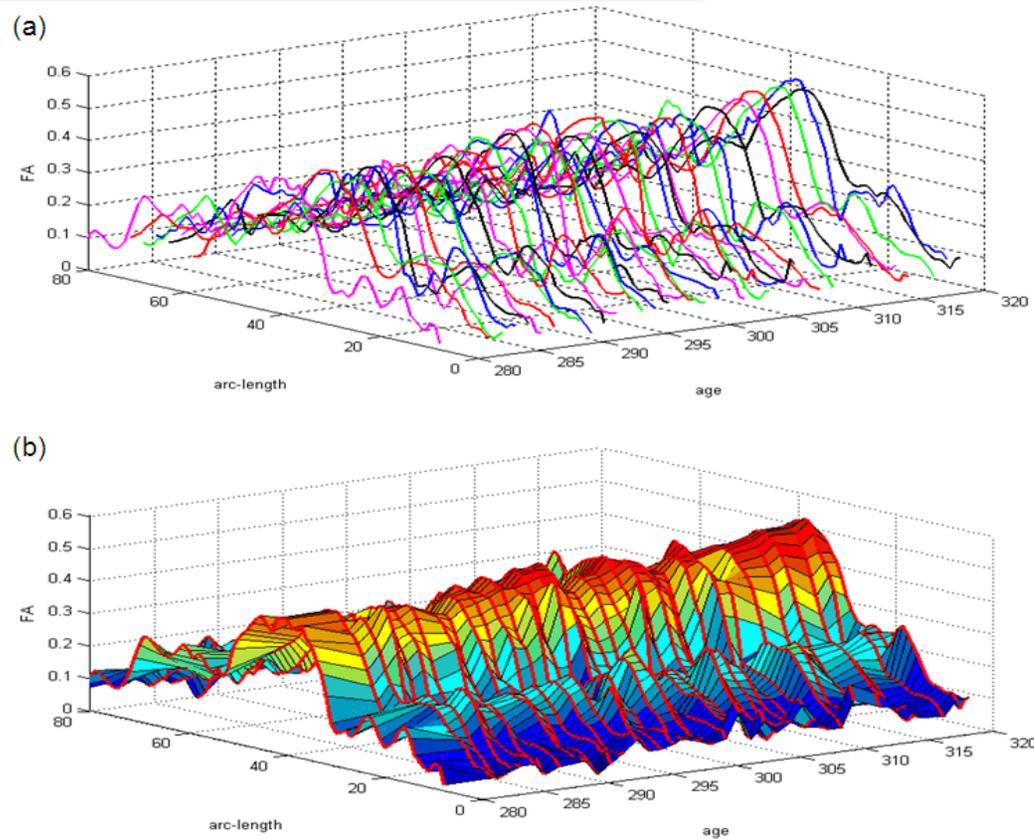
grids $\{s_1, \dots, s_{n_G}\}$

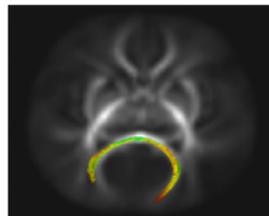
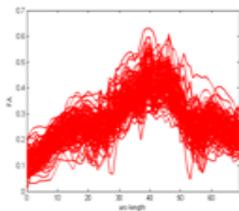
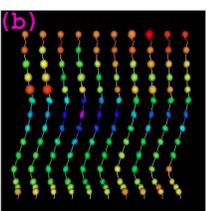
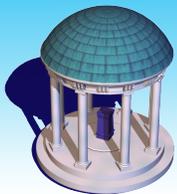
covariates x_1, \dots, x_n





Tract-based FA as a function of Age





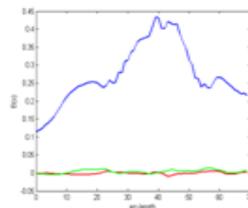
Varying Coefficient Model

Multivariate varying coefficient model

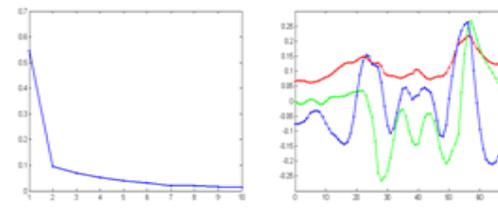
Weighted least square estimation

Functional principal component analysis

$$y_{i,k}(s) = \mathbf{x}_i^T B_k(s) + \eta_{i,k}(s) + \epsilon_{i,k}(s)$$



$$B_k(s_j) = B_k(s) + \dot{B}_k(s)(s_j - s)$$



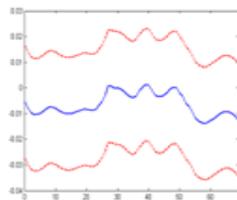
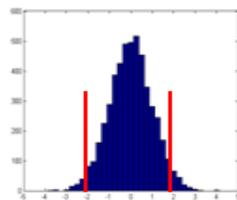
$$\hat{\Sigma}_{\eta,kk}(s, t) = \sum_{l=1} \hat{\lambda}_{k,l} \hat{\psi}_{k,l}(s) \hat{\psi}_{k,l}(t)$$

Resampling methods

Confidence bands

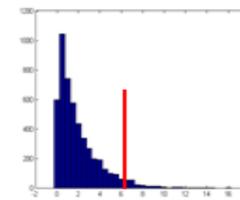
Hypothesis test

Resampling methods



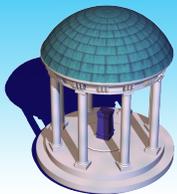
$$H_0 : C\text{vec}(B(s)) = \mathbf{b}_0(s)$$

$$H_1 : C\text{vec}(B(s)) \neq \mathbf{b}_0(s)$$



$$P(\sup_{s \in [0, L_0]} |G_{kl}(s)| \leq C_{kl}(\alpha)) = 1 - \alpha$$

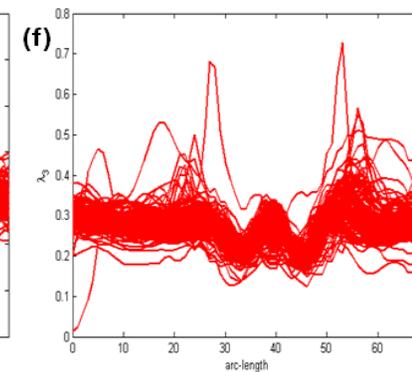
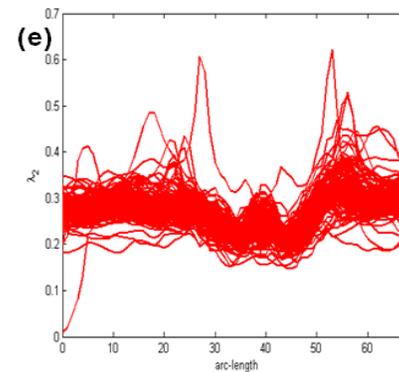
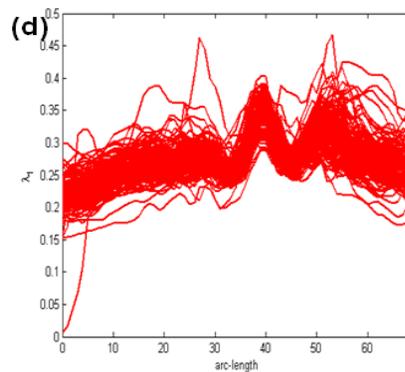
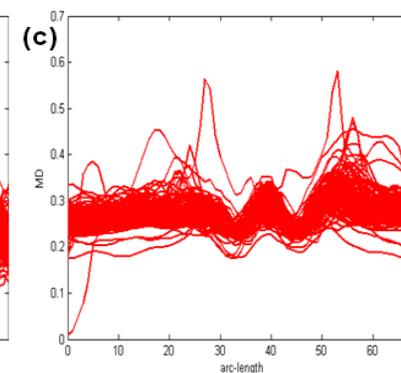
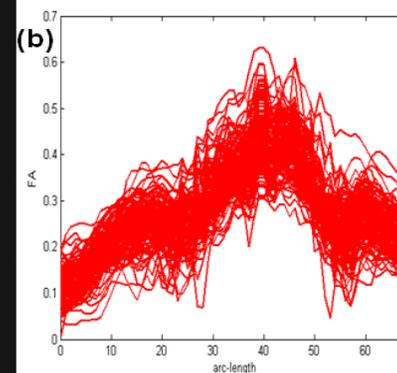
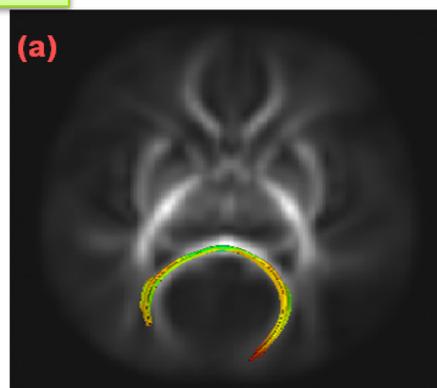
$$S_n = n \int_0^{L_0} \mathbf{d}(s)^T [C(\hat{\Sigma}_{\eta}(s, s) \otimes \hat{\Omega}_X^{-1}) C^T]^{-1} \mathbf{d}(s) ds \quad p = G^{-1} \sum_{g=1}^G 1(S_n^{(g)} \geq S_n)$$



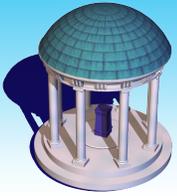
PI: John H. Gilmore

Subjects:
125 healthy infants
(75:53 M:F)
Gestational age
(298 \pm 17.6 days)

Aims:
Gender Effects
Age Effects

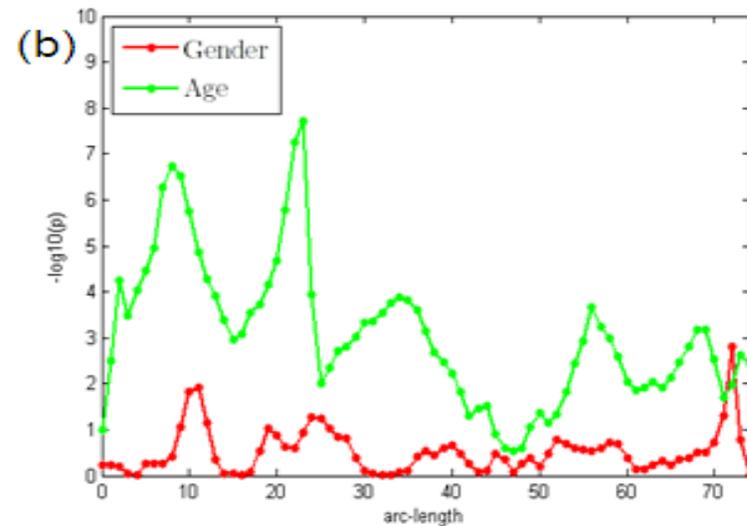
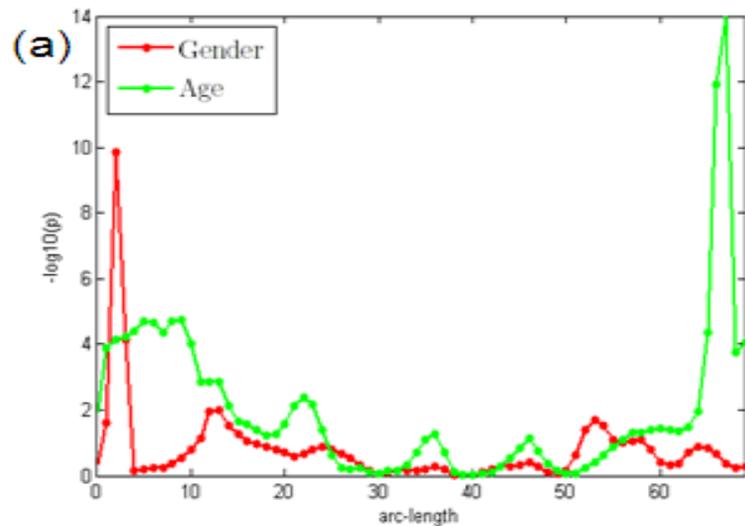


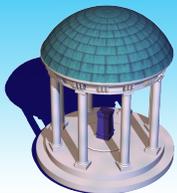
Splenium



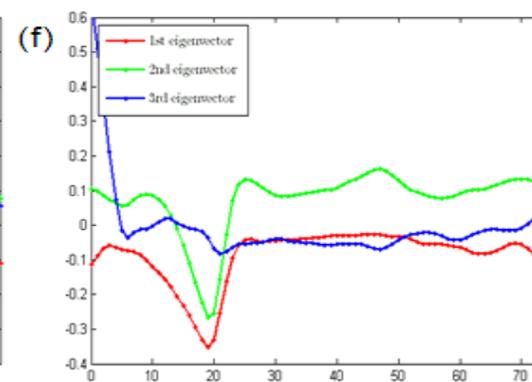
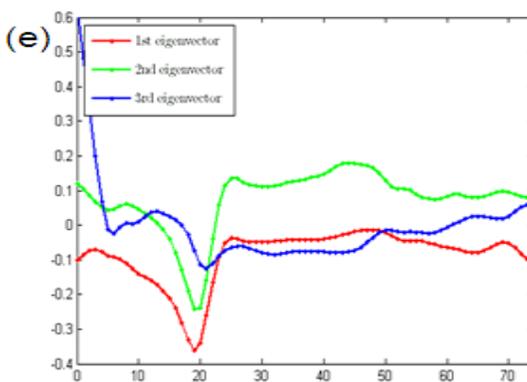
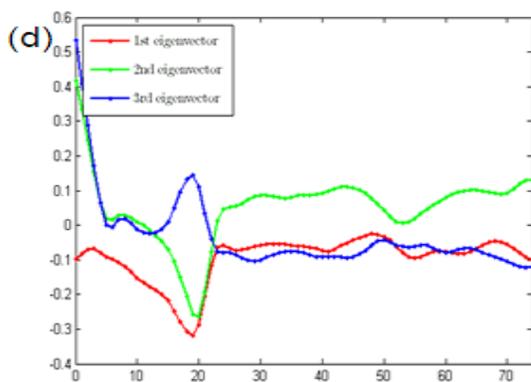
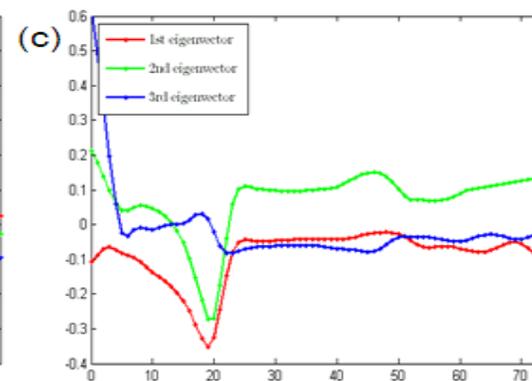
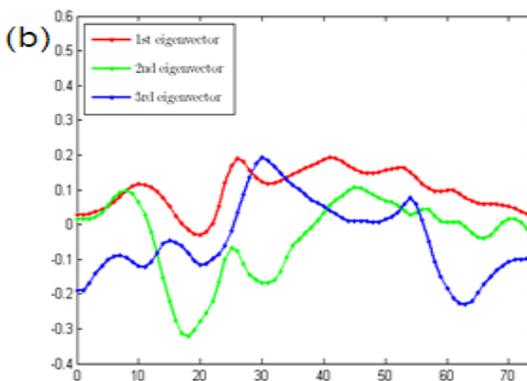
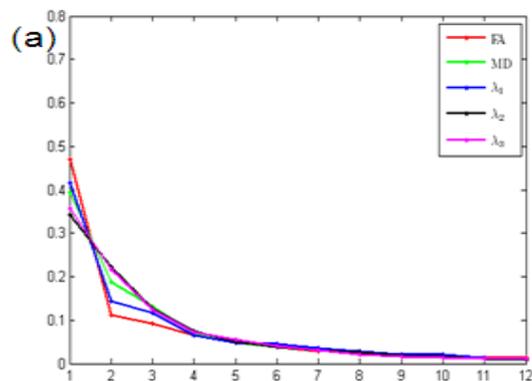
Global and Local p-values for Gender and Age Effects

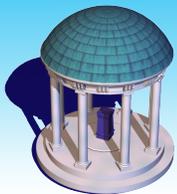
	Right Internal Capsule					Splenium				
	FA	MD	Lambda1	Lambda2	Lambda3	FA	MD	Lambda1	Lambda2	Lambda3
Gender	.169	.354	.241	.314	.376	.683	.063	.048	.057	.302
Age	<.001	<.001	<.001	<.001	<.001	.103	.008	.003	.007	.010



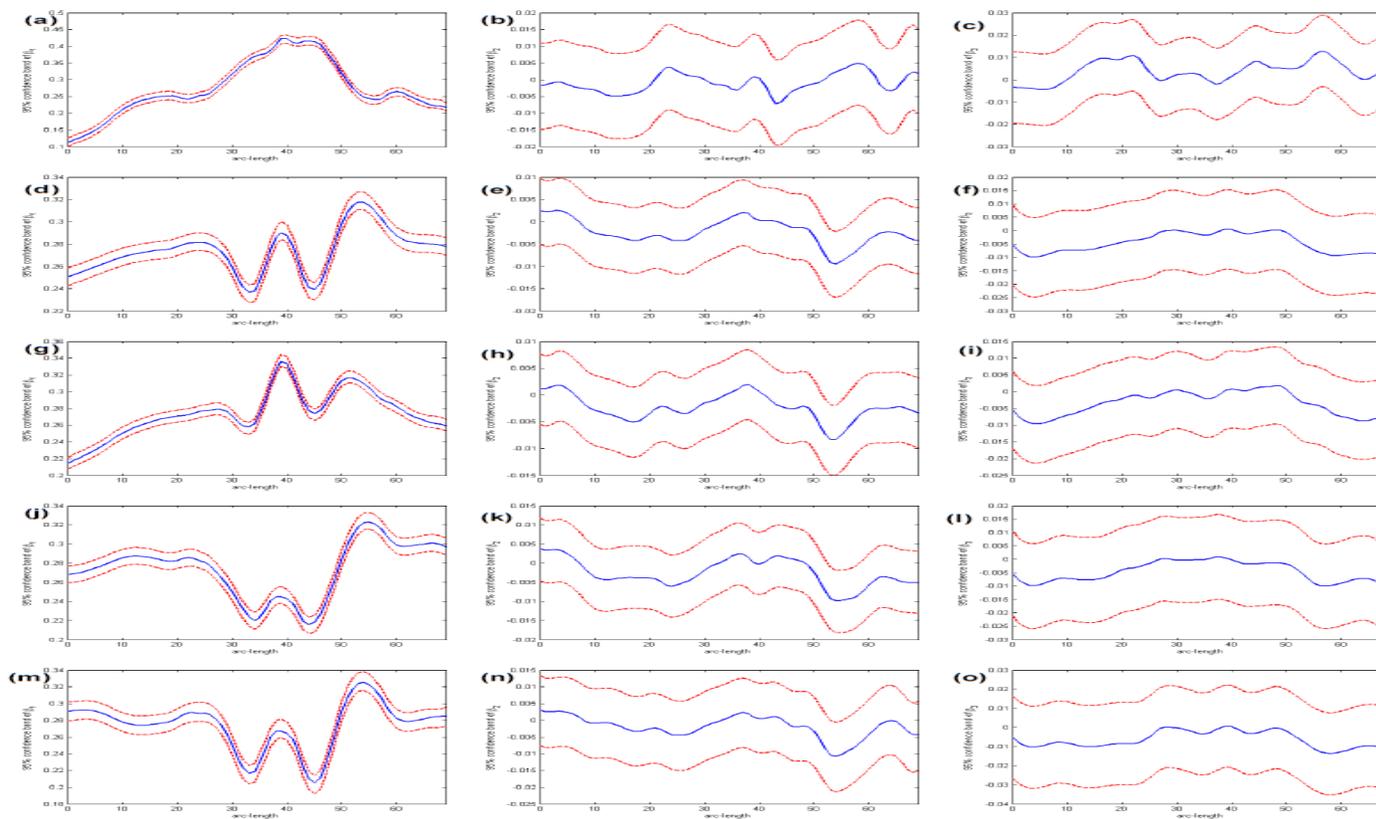


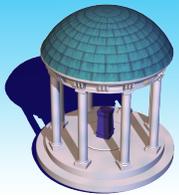
Functional Principal Components





Simultaneous Confidence Bands

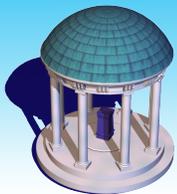




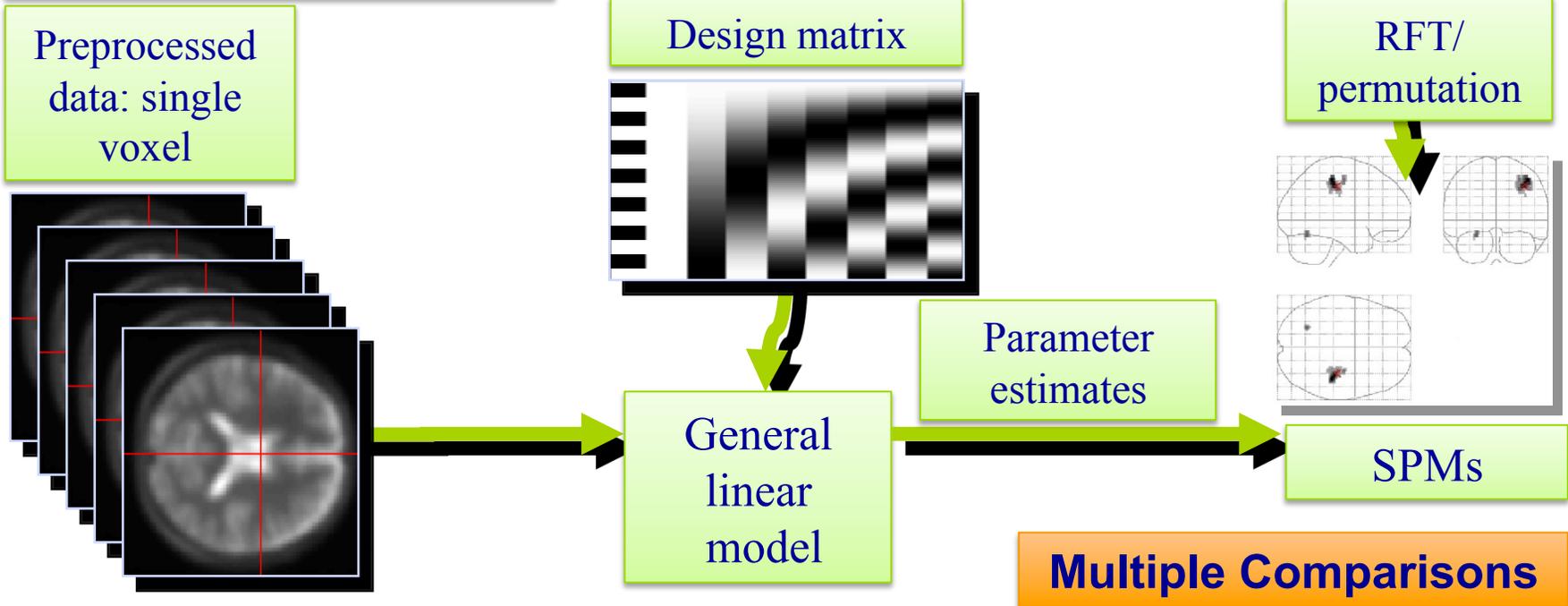
Multiscale Adaptive **Regression** Models

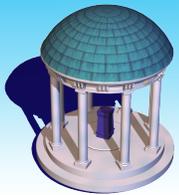
Regression Analysis and Nonparametric
Smoothing Methods

How to **spatially and adaptively** compare
imaging measures across subjects
in 3D volume or 2D surface?



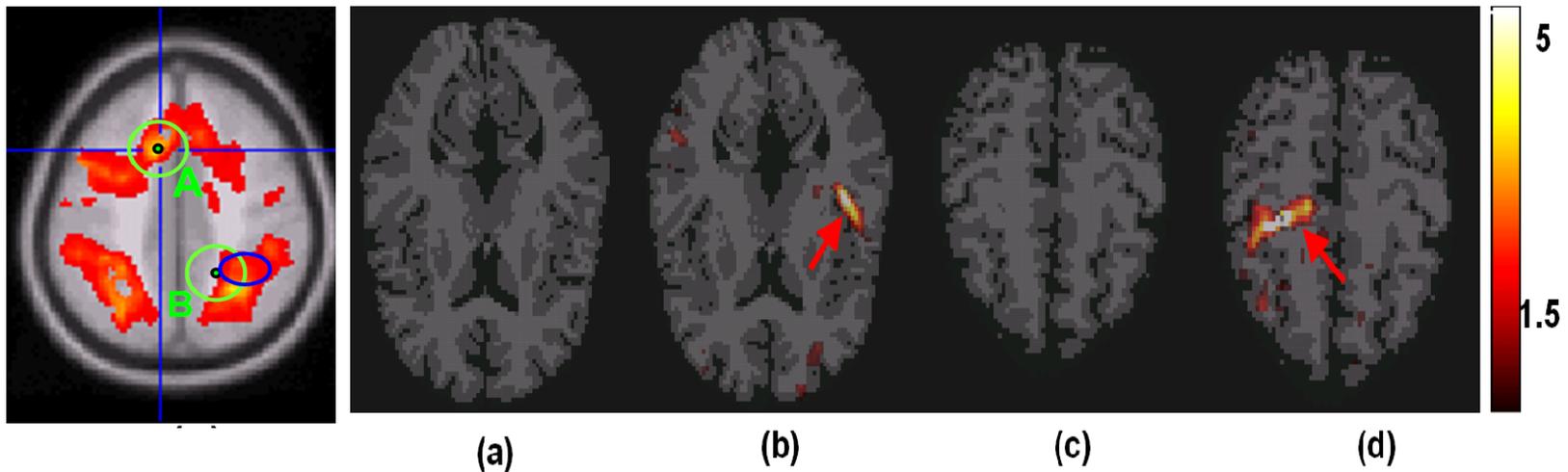
Univariate Statistics

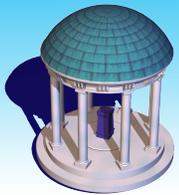




All voxels are treated as **independent units**.

Initial smoothing step before the voxel-wise approach often blurs the image data near the edges of activated regions.





Multiscale Adaptive Regression Model

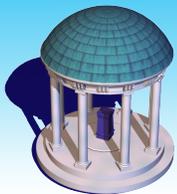
Learning Voxel Feature

Local Feature Adaptation

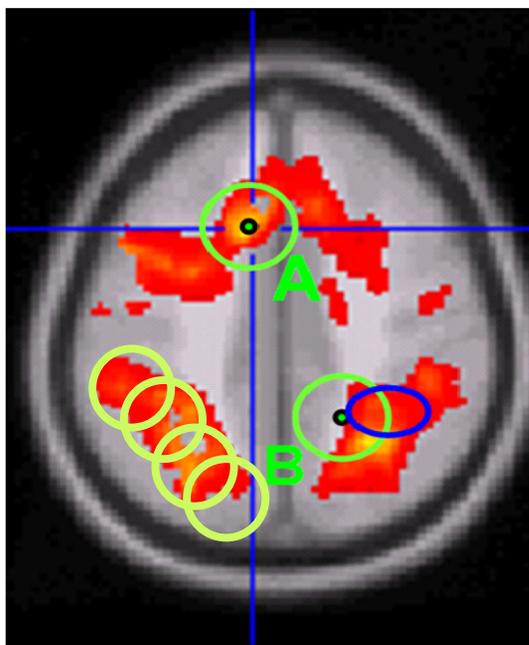
Adaptive Estimation and Testing

Automatic Stop

Nice Asymptotic Results

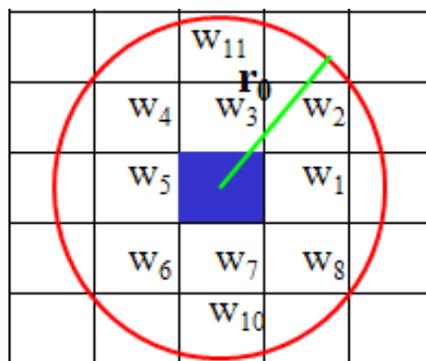


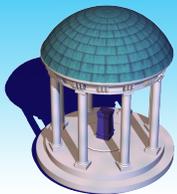
Identifying homogeneous regions



Drawing a sphere with radius r_0 at each voxel

Calculating the similarities between the current voxel and its neighboring voxels.



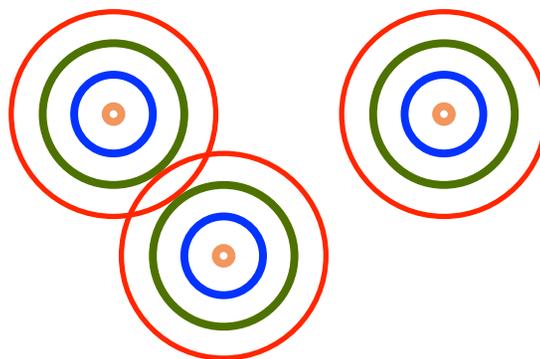


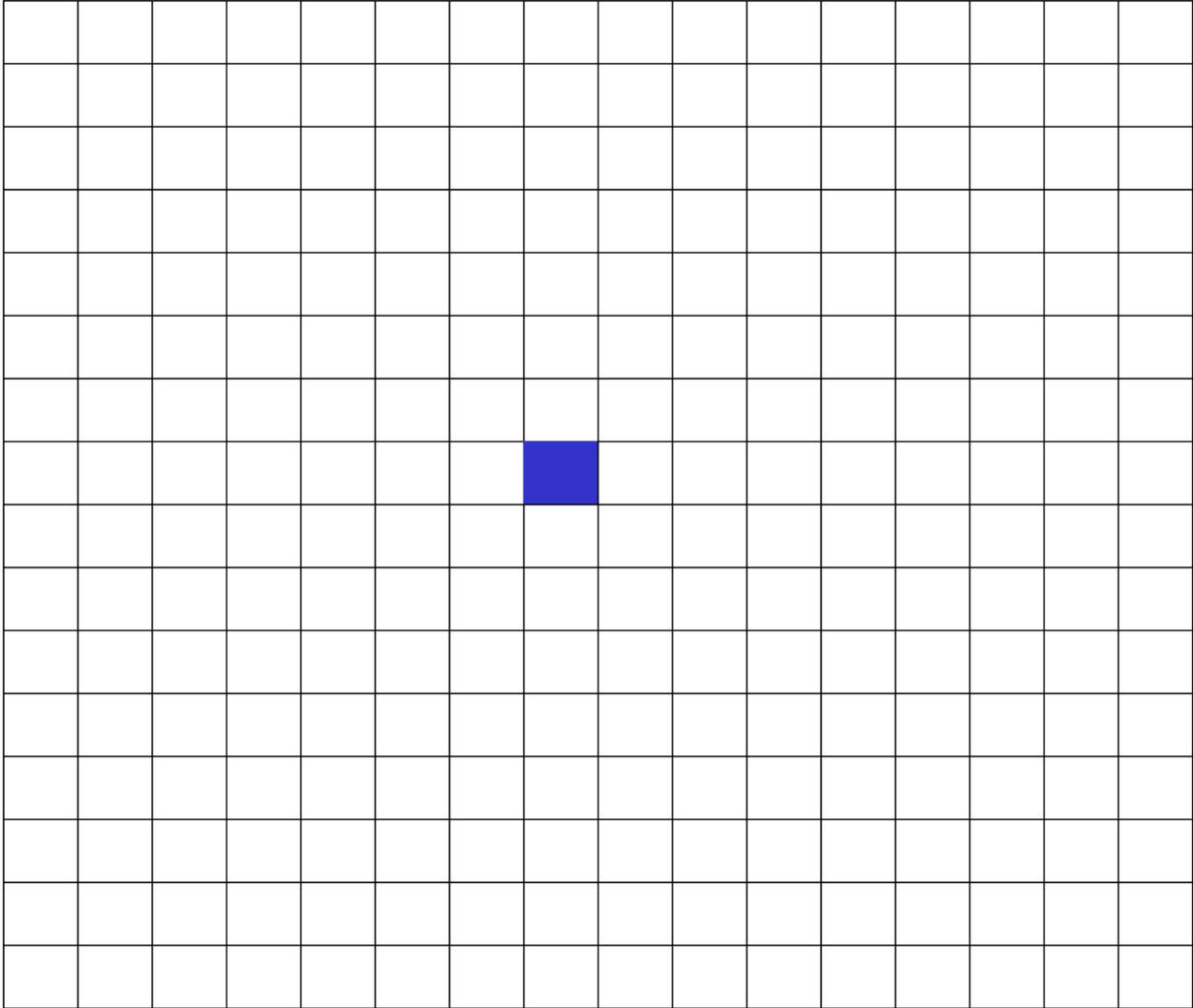
Being Hierarchical

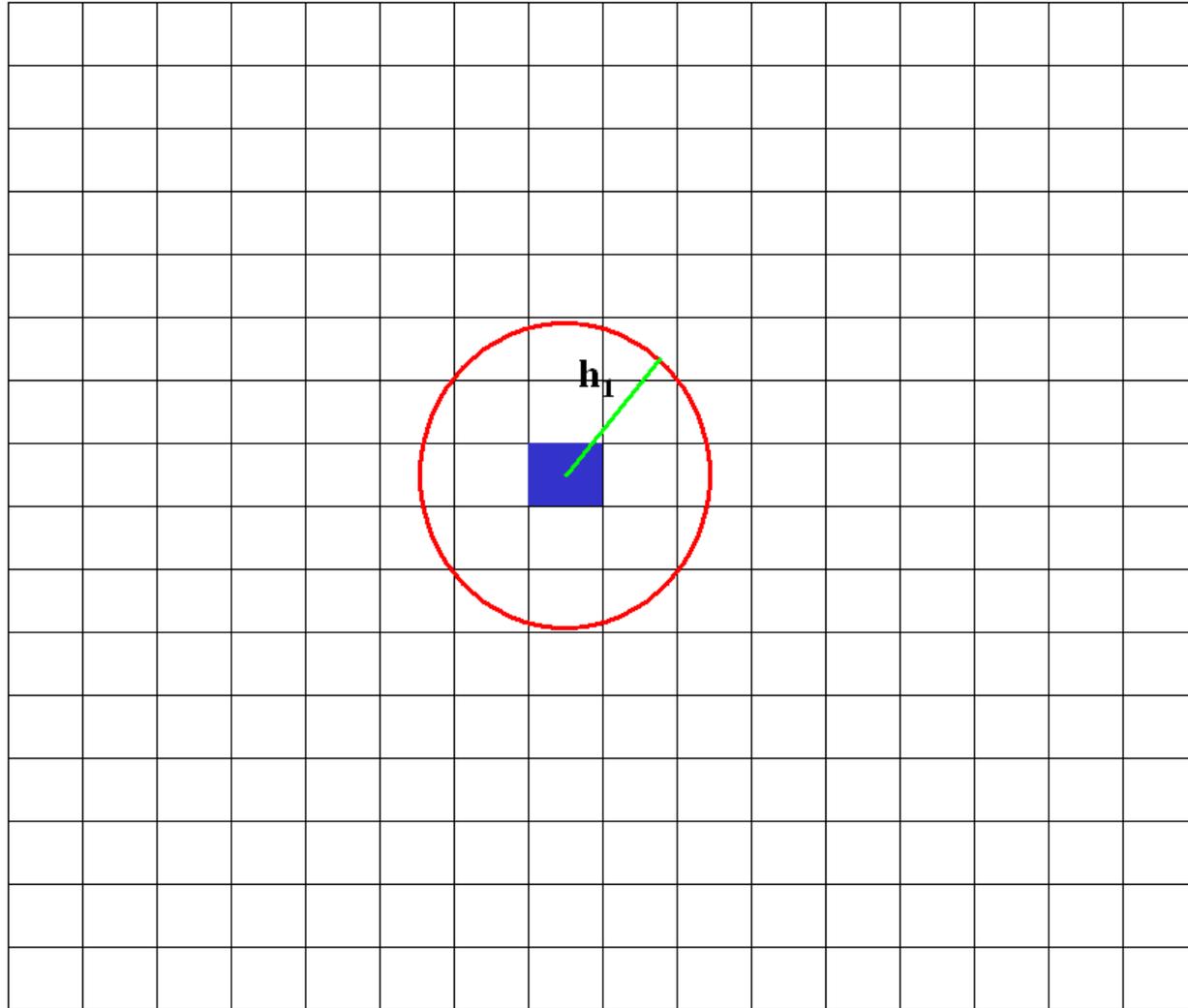
Drawing nested spheres with increasing radiuses at each voxel

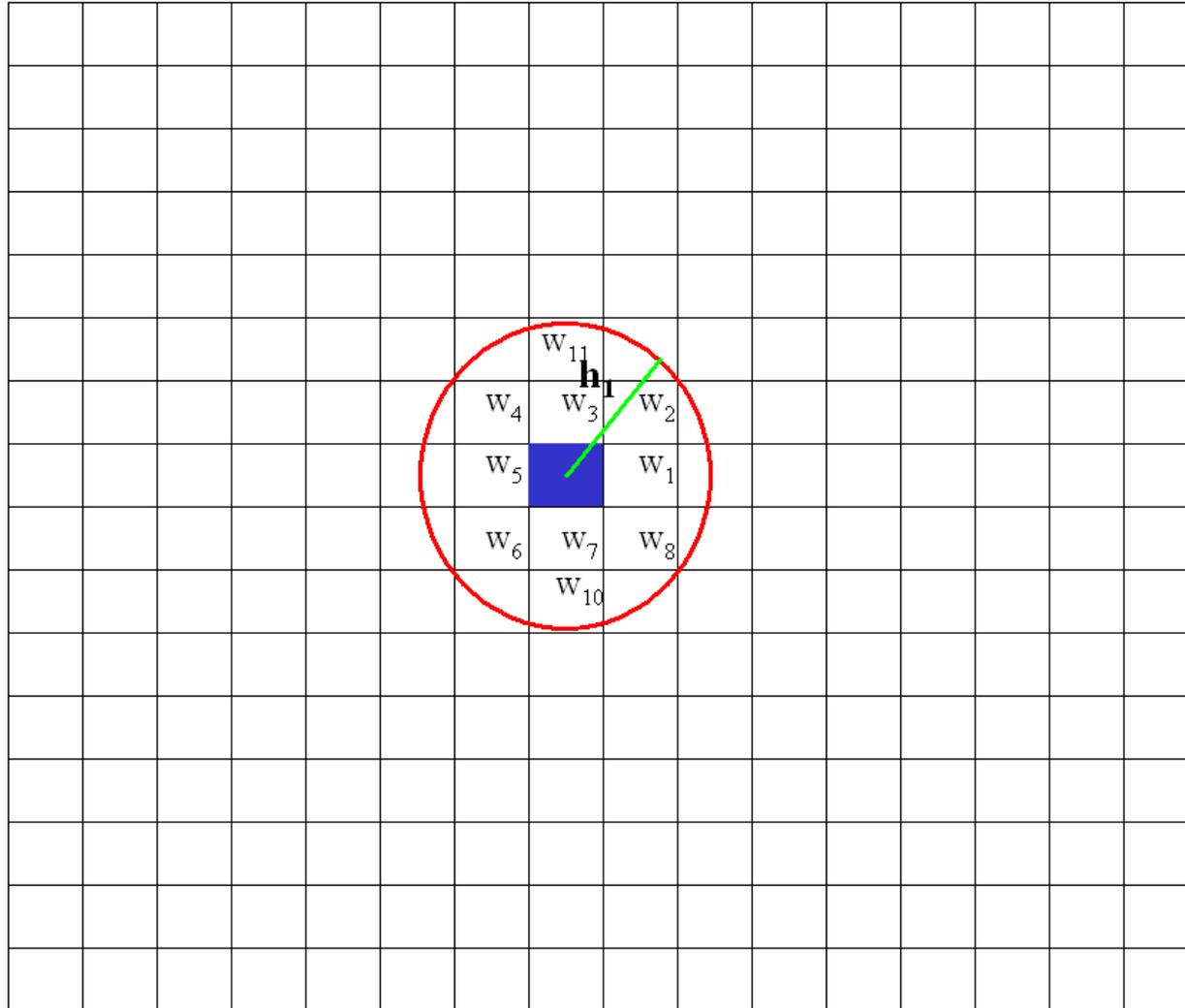


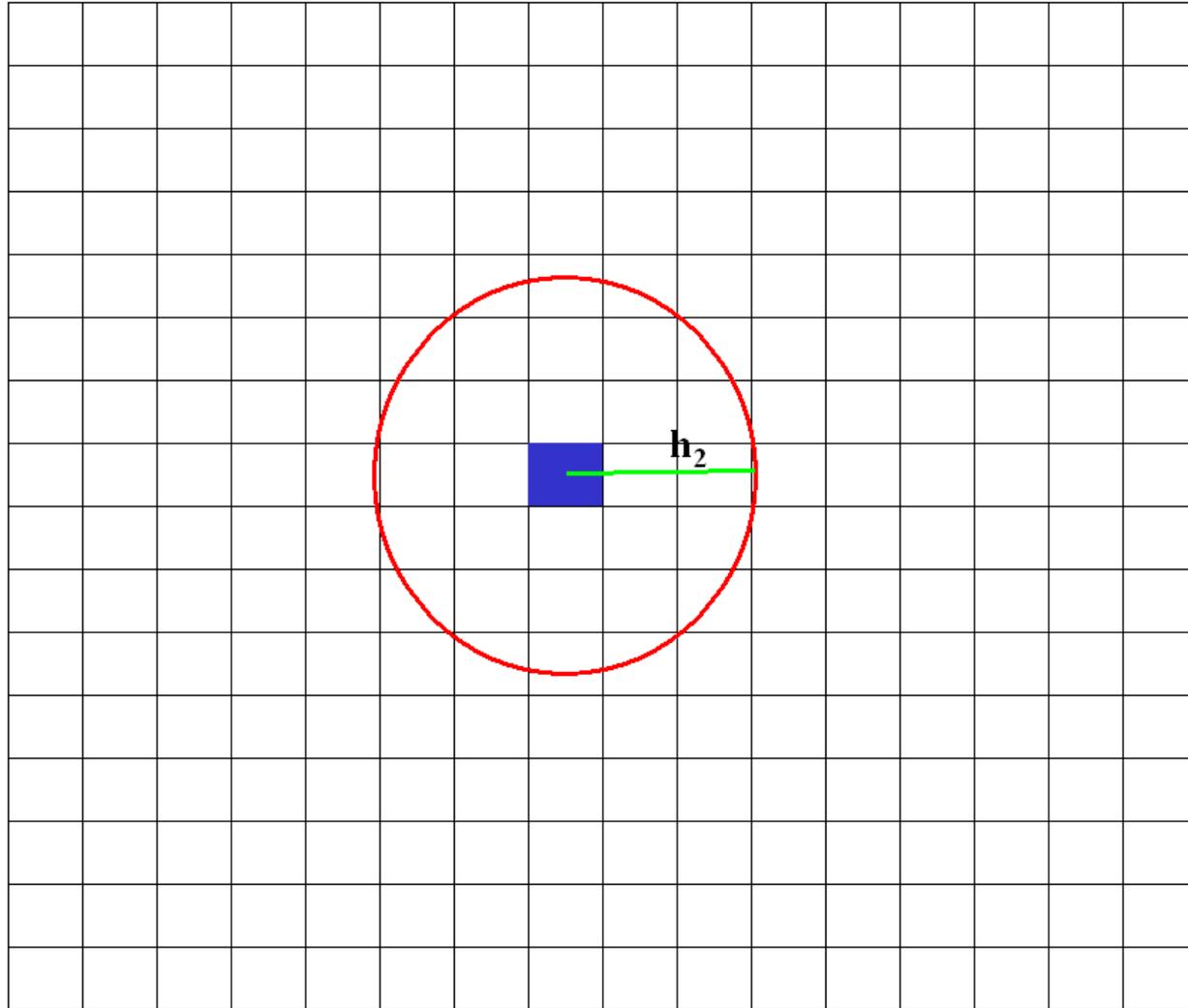
$$h_0 = 0 < h_1 < \dots < h_S = r_0$$

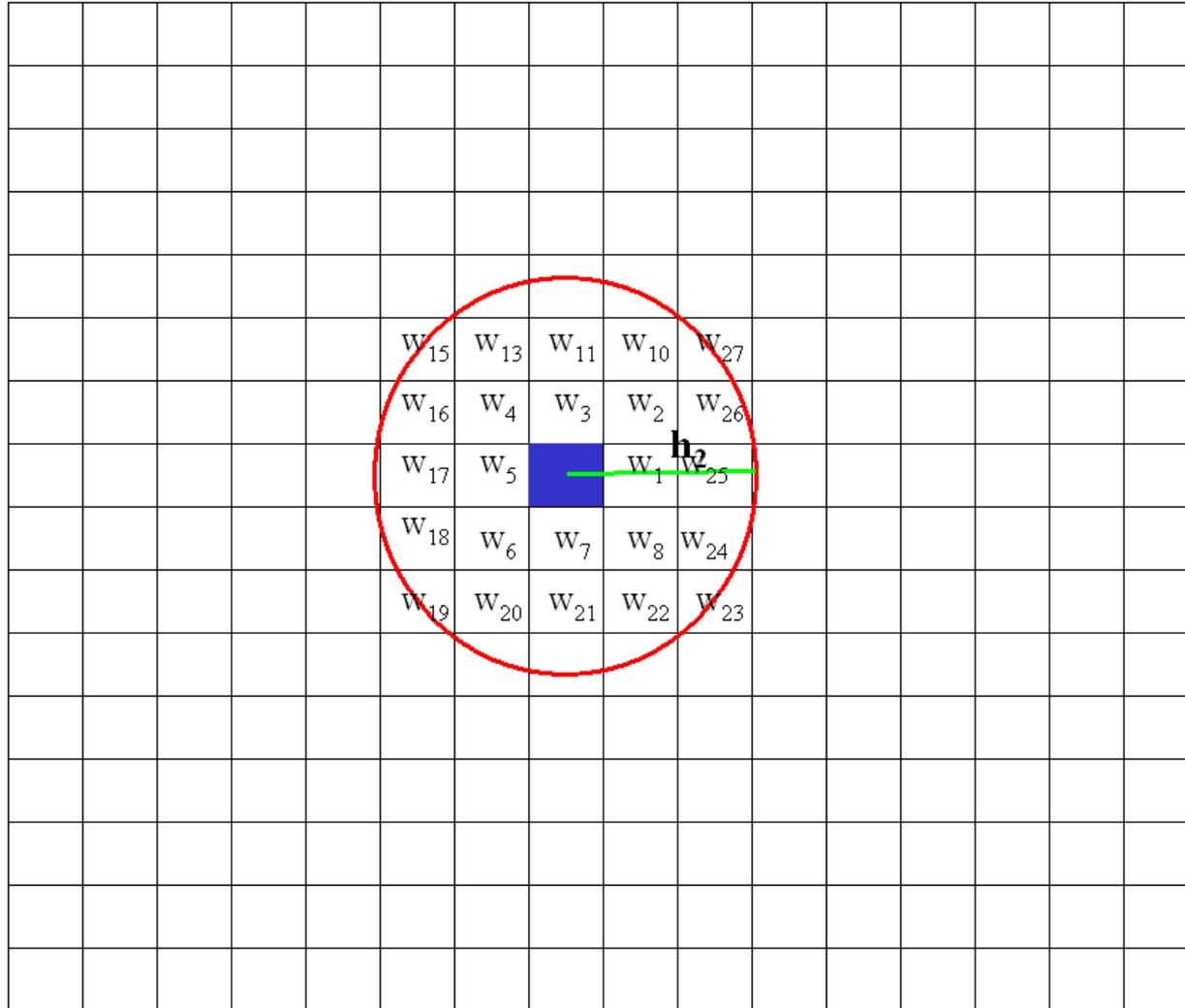


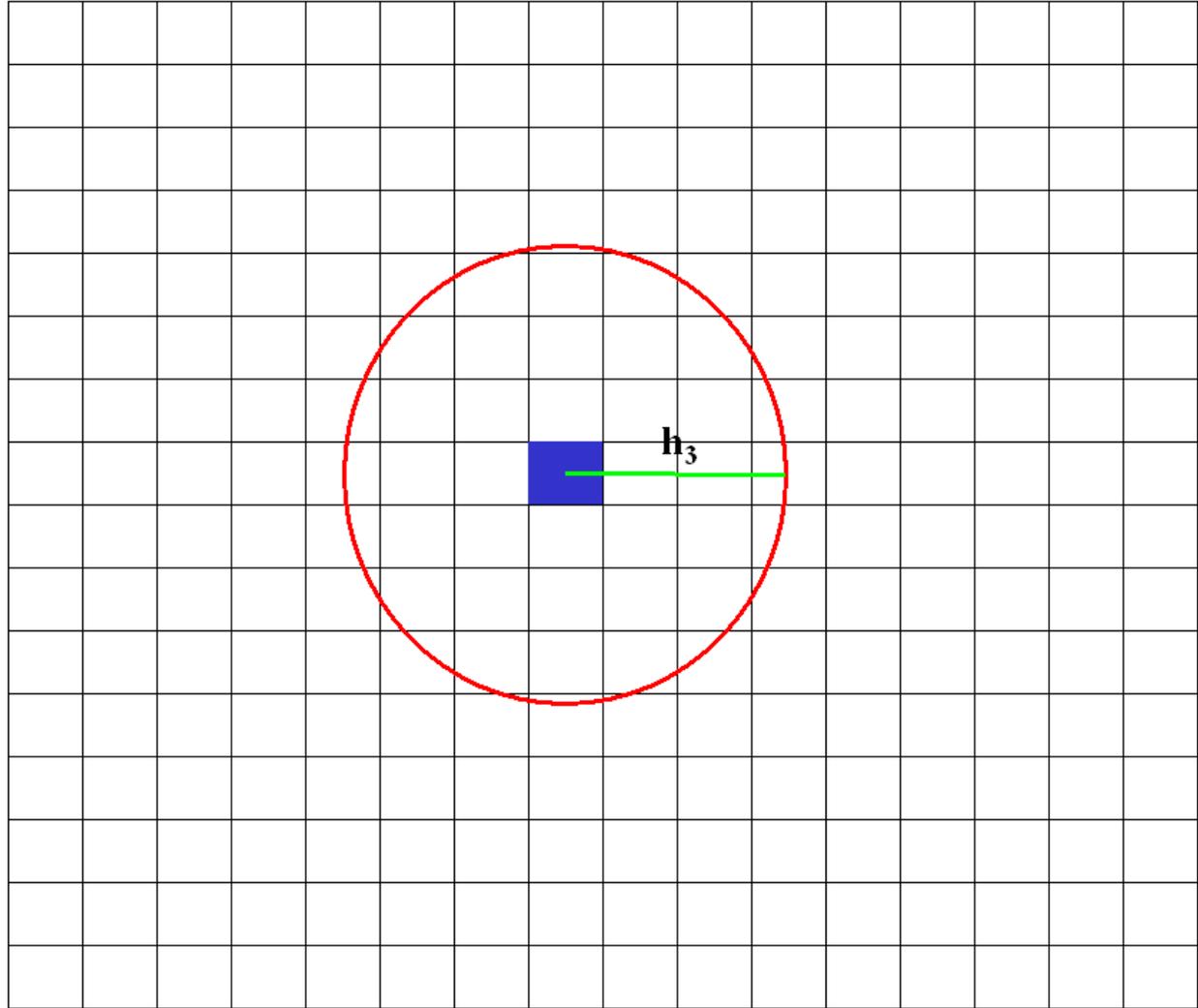


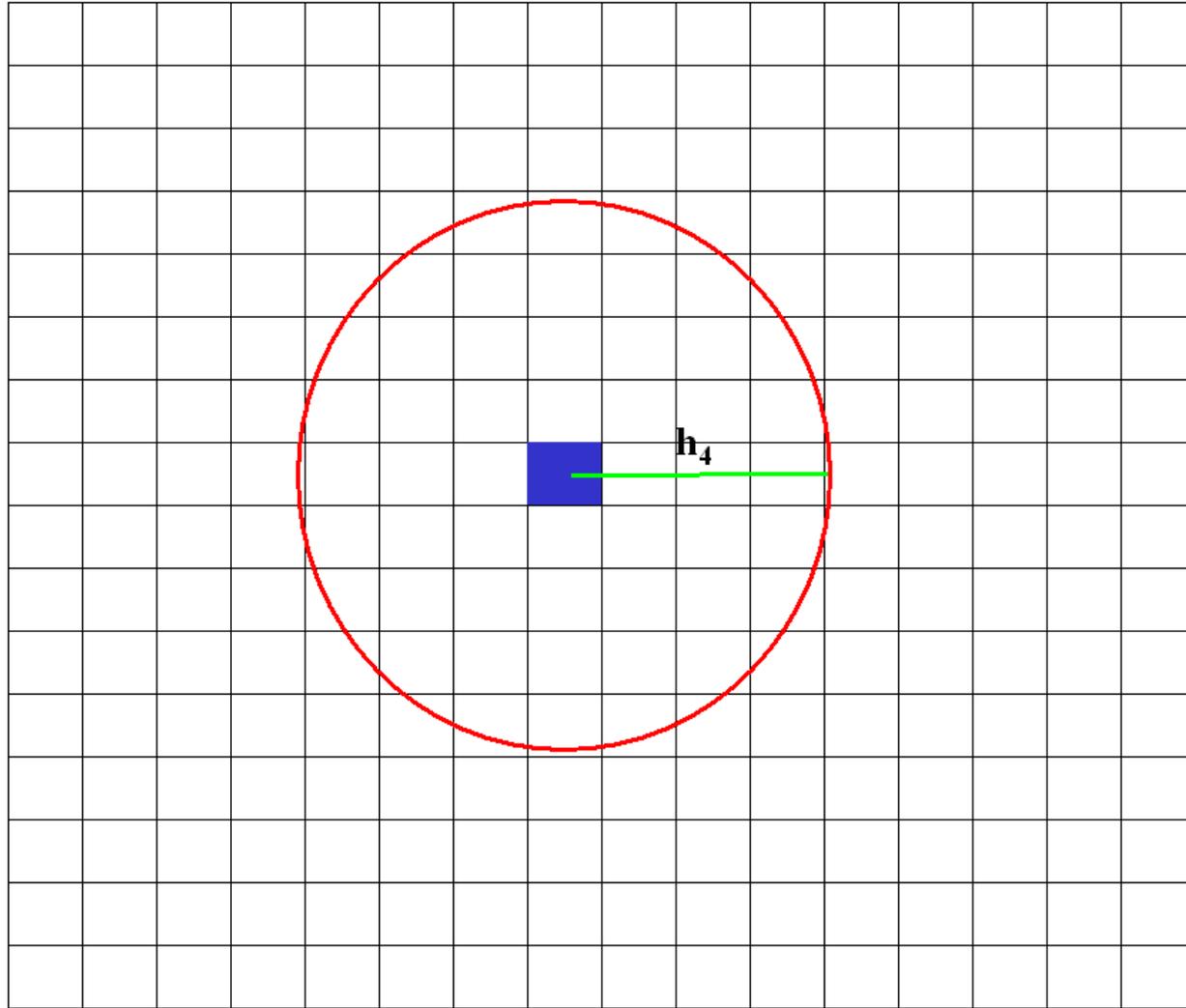


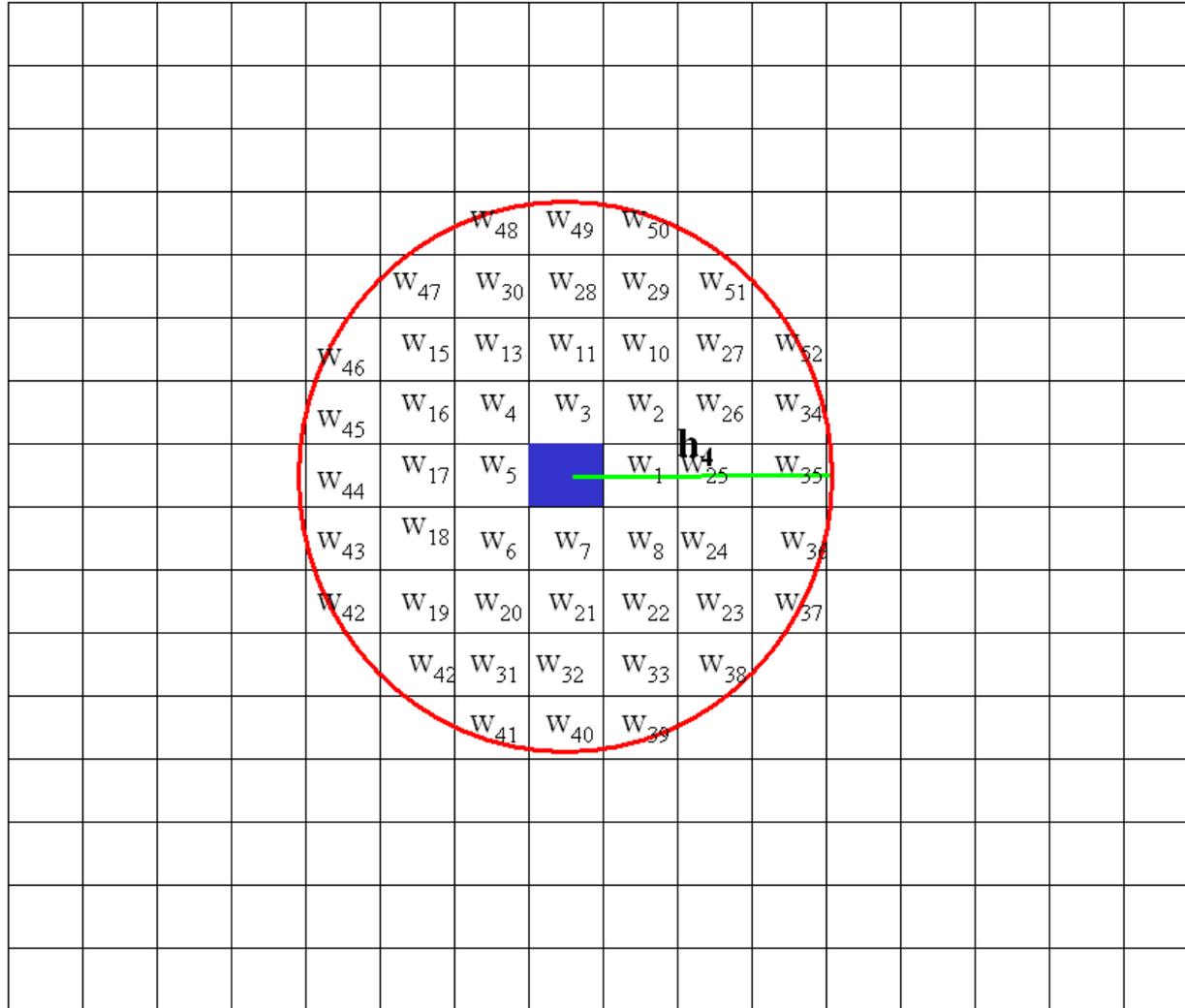


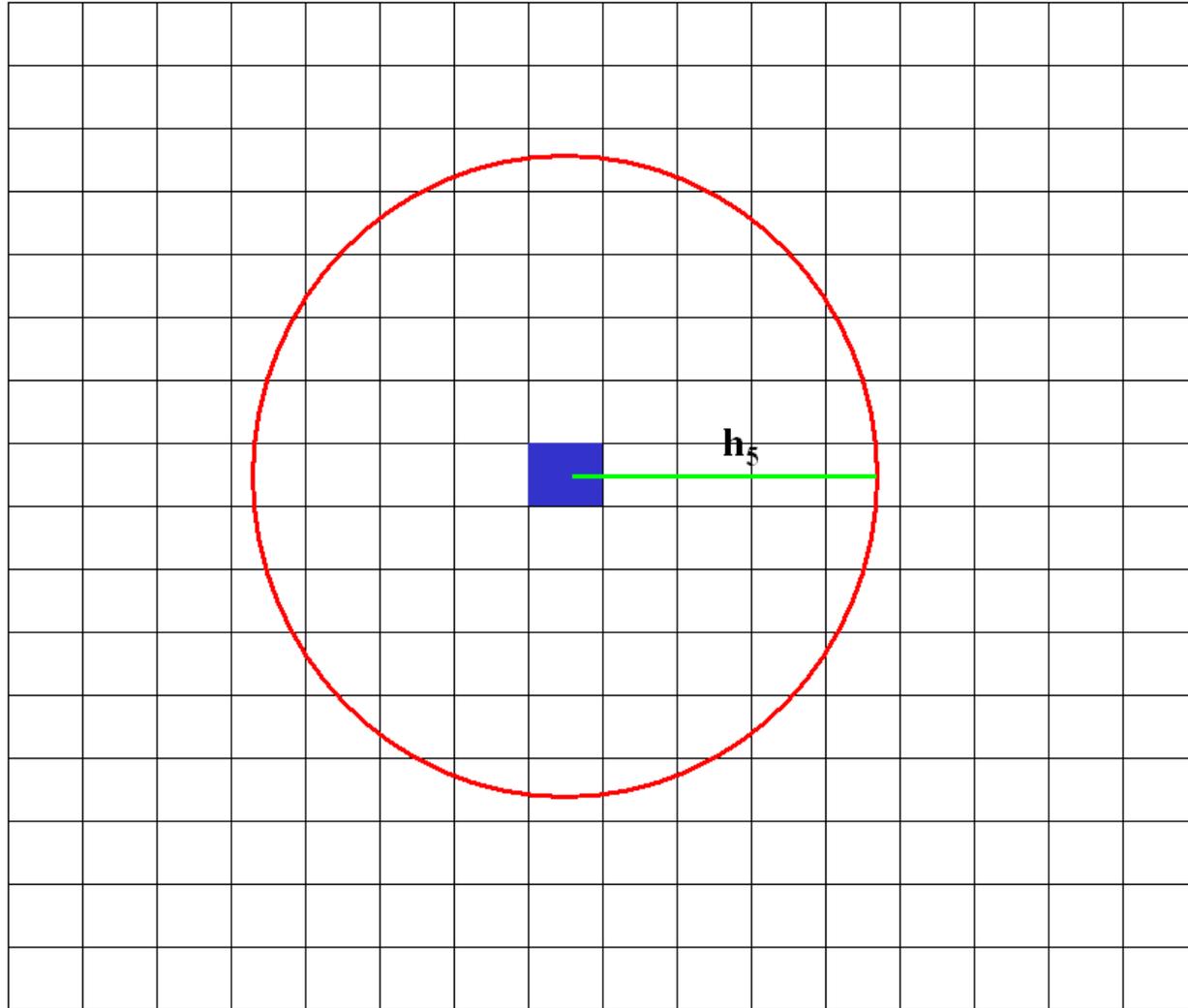


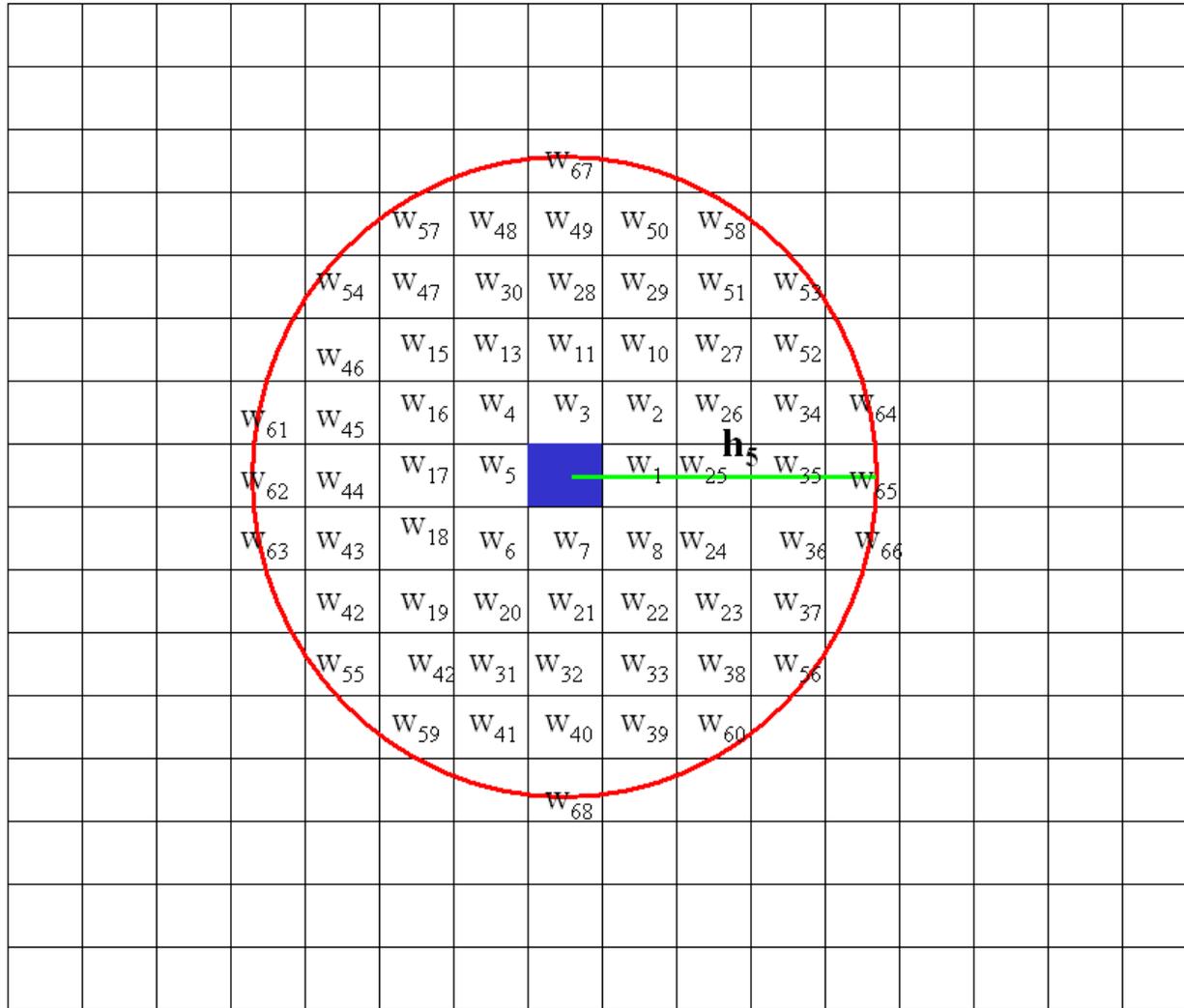


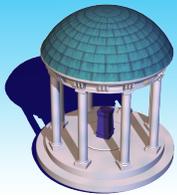






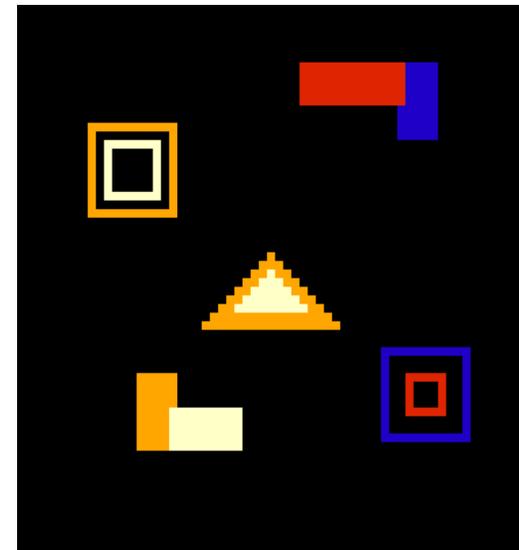






Simulation Studies

64x64 phantom image



Model $y_i(d) = x_i^T \beta(d) + \varepsilon_i(d)$

Error $\varepsilon_i(d) \sim N(0,1)$ $\varepsilon_i(d) \sim \chi^2(3) - 3$

$n = 60$ or $n = 80$

Covariates $x_i = (1, x_{i2}, x_{i3})^T$

$x_{i2} \sim \text{Bernoulli}(0.5)$

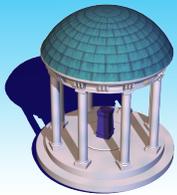
$x_{i3} \sim \text{Uniform}[1,2]$

Coefficients $\beta(d) = (\beta_1(d), \beta_2(d), \beta_3(d))^T$

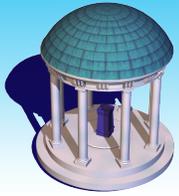
$\beta_1(d) = \beta_2(d) = 0$

ROIs

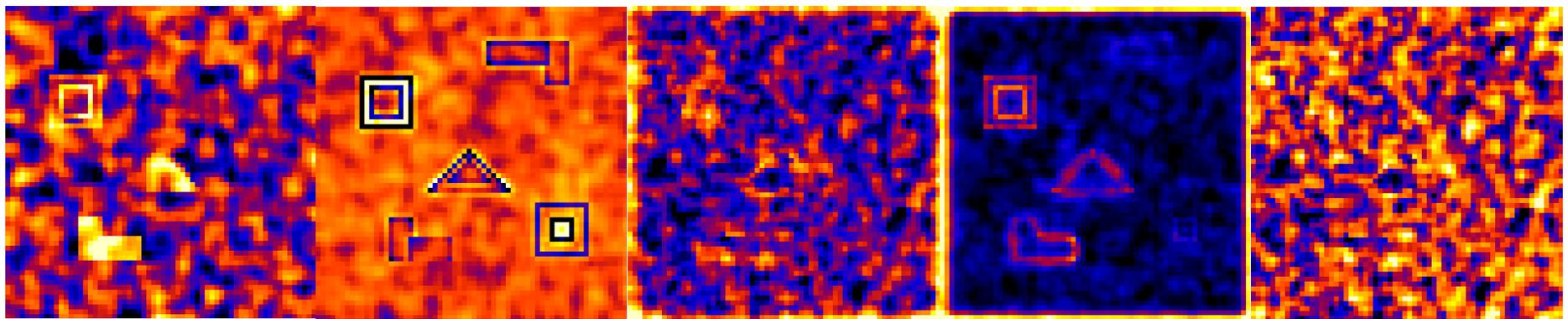
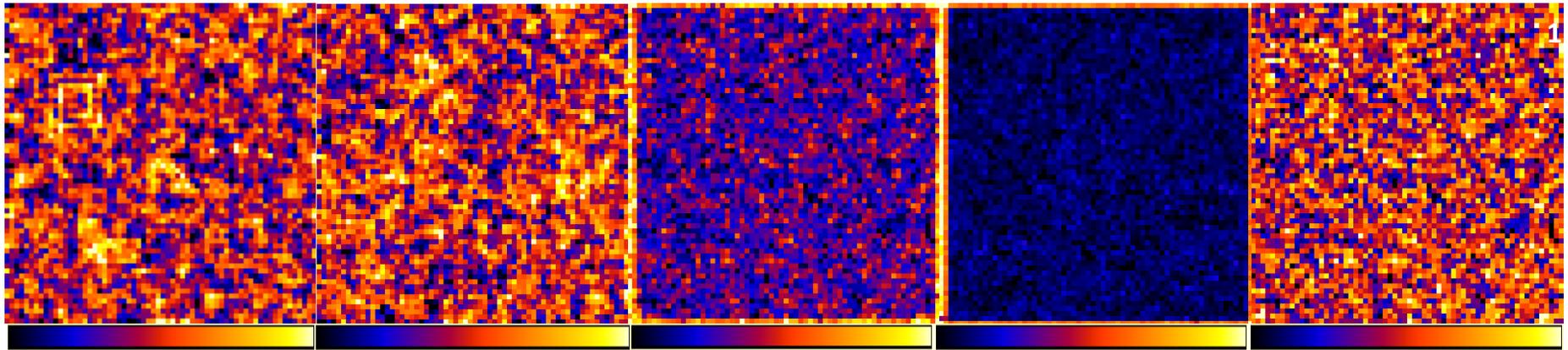
ROI	<i>black</i>	<i>blue</i>	<i>red</i>	<i>yellow</i>	<i>white</i>
$\beta_3(d)$	0.0	0.2	0.4	0.6	0.8

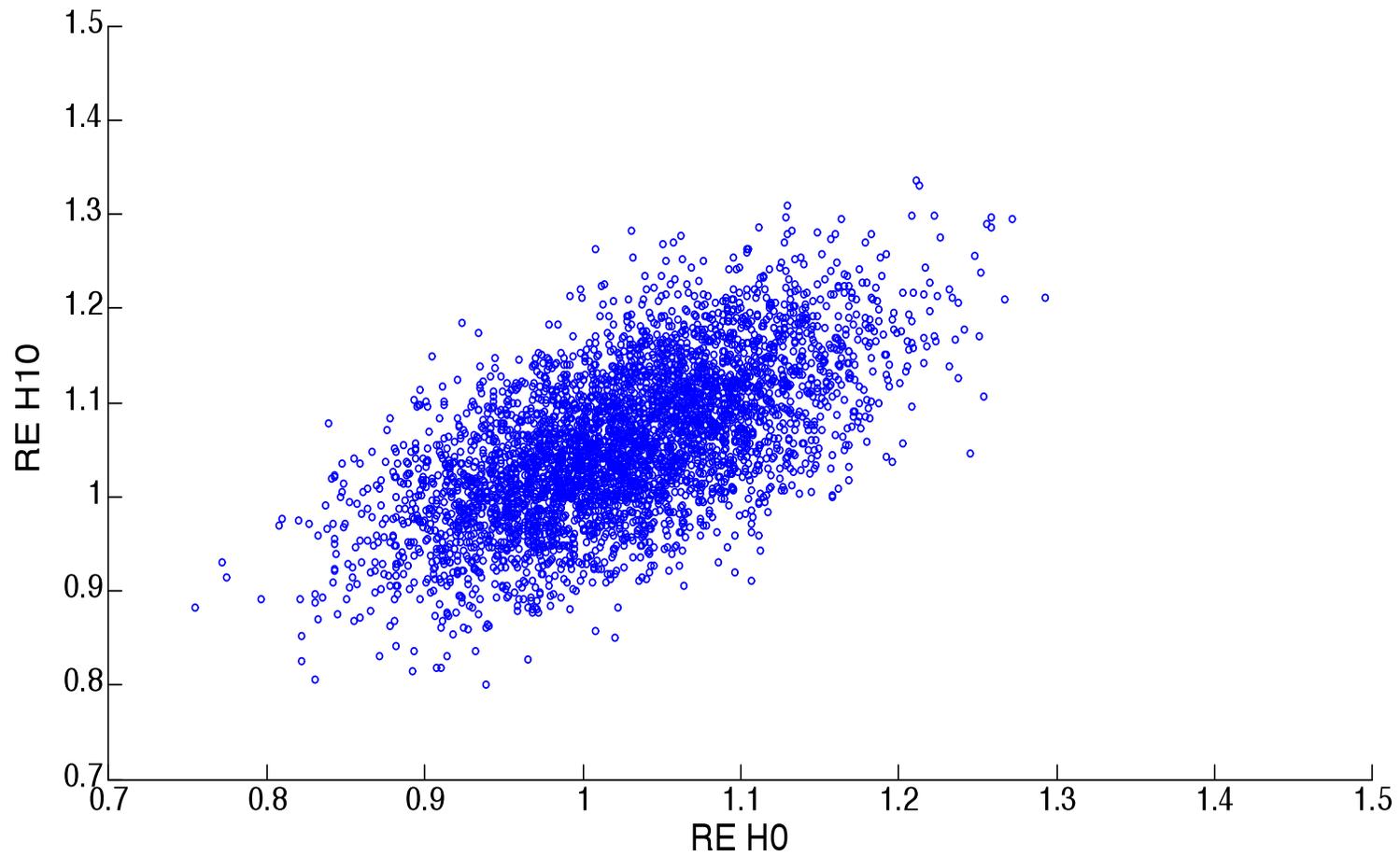
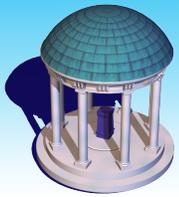


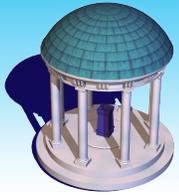
$\beta_2(d)$	$\chi^2(3) - 3$							$N(0, 1)$					
	$n = 60$			$n = 80$				$n = 60$			$n = 80$		
		h_0	h_5	h_{10}	h_0	h_5	h_{10}	h_0	h_5	h_{10}	h_0	h_5	h_{10}
0.0	BIAS	0.00	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
	RMS	0.48	0.35	0.26	0.41	0.31	0.22	0.20	0.15	0.11	0.17	0.13	0.09
	SD	0.47	0.34	0.24	0.41	0.30	0.21	0.19	0.14	0.10	0.17	0.12	0.09
	RE	1.03	1.05	1.06	1.02	1.03	1.04	1.03	1.05	1.06	1.02	1.03	1.04
0.2	BIAS	0.00	-0.03	-0.07	0.01	-0.02	-0.06	0.00	-0.03	-0.05	0.00	-0.02	-0.05
	RMS	0.46	0.34	0.24	0.39	0.29	0.21	0.19	0.14	0.11	0.16	0.12	0.09
	SD	0.46	0.33	0.24	0.40	0.29	0.21	0.19	0.14	0.10	0.16	0.12	0.09
	RE	1.01	1.01	1.01	0.99	1.00	1.01	1.02	1.04	1.06	1.02	1.02	1.03
0.4	BIAS	-0.01	-0.05	-0.09	0.01	-0.02	-0.06	0.00	0.00	-0.01	0.00	0.00	0.00
	RMS	0.46	0.34	0.25	0.40	0.30	0.22	0.19	0.15	0.12	0.16	0.13	0.10
	SD	0.46	0.33	0.24	0.40	0.29	0.21	0.19	0.14	0.11	0.16	0.12	0.09
	RE	1.01	1.02	1.03	1.01	1.02	1.03	1.03	1.05	1.07	1.00	1.01	1.02
0.6	BIAS	0.00	-0.05	-0.09	0.00	-0.04	-0.07	0.00	0.01	0.02	0.00	0.00	0.01
	RMS	0.46	0.35	0.26	0.40	0.30	0.23	0.19	0.15	0.12	0.16	0.13	0.10
	SD	0.46	0.34	0.25	0.40	0.30	0.22	0.19	0.14	0.11	0.16	0.13	0.10
	RE	1.01	1.03	1.04	1.01	1.02	1.03	1.02	1.04	1.06	1.01	1.03	1.04
0.8	BIAS	0.00	-0.04	-0.06	0.00	-0.02	-0.05	0.00	-0.01	-0.02	0.00	0.00	-0.01
	RMS	0.47	0.35	0.26	0.40	0.30	0.23	0.19	0.15	0.11	0.17	0.13	0.10
	SD	0.46	0.34	0.25	0.40	0.30	0.22	0.19	0.14	0.11	0.16	0.12	0.09
	RE	1.02	1.03	1.04	1.01	1.02	1.03	1.02	1.04	1.05	1.03	1.05	1.06



$$\hat{\beta}_3(d, h_0)$$



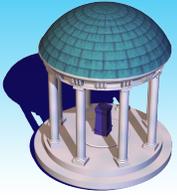




Simulation Studies

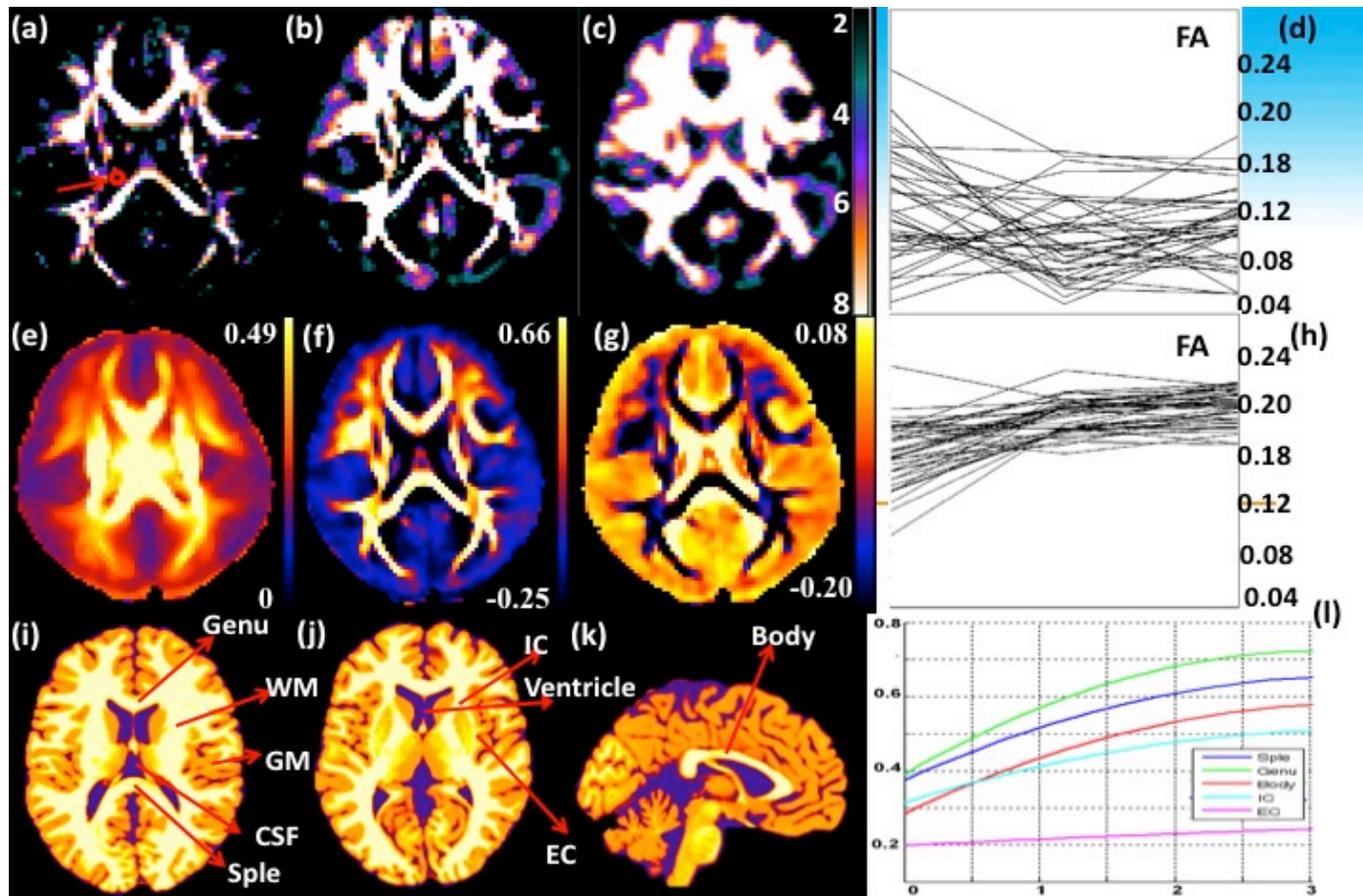
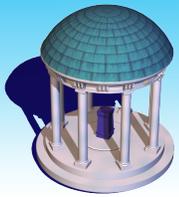
Table 2. Simulation study for $W_\mu(d, h)$: estimates (ES) and standard errors (SE) of rejection rates for pixels inside the five ROIs were reported at 2 different scales (h_0, h_{10}), 2 different distributions ($N(0, 1)$ and $\chi^2(3) - 3$), and 2 different sample sizes ($n = 60, 80$) at $\alpha = 5\%$. For each case, 1,000 simulated data sets were used.

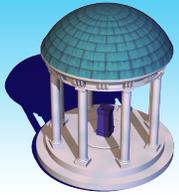
$\beta_2(d)$	s	$N(0, 1)$				$\chi^2(3) - 3$			
		$n = 60$		$n = 80$		$n = 60$		$n = 80$	
		ES	SE	ES	SE	ES	SE	ES	SE
0.2	h_0	0.20	0.066	0.24	0.070	0.08	0.038	0.08	0.037
	h_{10}	0.30	0.126	0.38	0.121	0.10	0.069	0.18	0.081
0.4	h_0	0.56	0.090	0.67	0.079	0.15	0.065	0.18	0.070
	h_{10}	0.93	0.051	0.98	0.030	0.26	0.129	0.35	0.159
0.6	h_0	0.88	0.039	0.95	0.024	0.27	0.057	0.33	0.050
	h_{10}	1.00	0.004	1.00	0.004	0.51	0.091	0.63	0.083
0.8	h_0	0.99	0.015	1.00	0.005	0.43	0.080	0.52	0.080
	h_{10}	0.99	0.010	0.99	0.011	0.78	0.099	0.90	0.006
0.0	h_0	0.07	0.006	0.07	0.006	0.06	0.007	0.07	0.006
	h_{10}	0.08	0.011	0.07	0.011	0.07	0.012	0.08	0.012



Infant Brain Development Data

- **Objective:** We want to assess the brain structure change in the early brain development.
- **Subject:** 38 infants.
- **Image:** Diffusion-weighted images and T1 weighted images were acquired for each subject at 2 weeks, 1 and 2 years old.
- **Method:** Voxel-wise imaging analysis and MARM.





New Developments

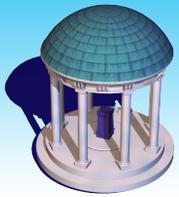
Adaptive Neighborhoods

Adaptive Weights

**Cross-sectional, longitudinal,
twin and family studies**

Robust Procedure

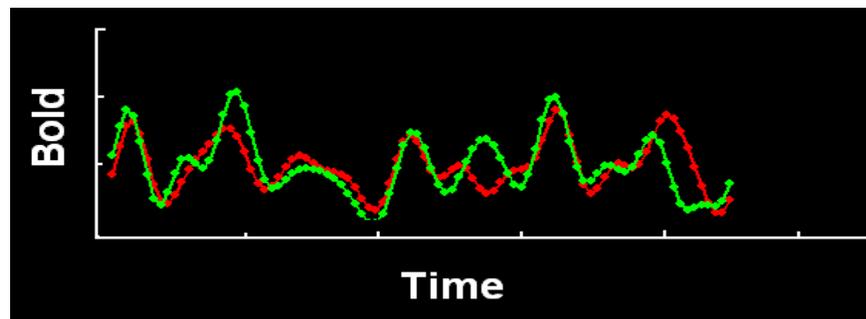
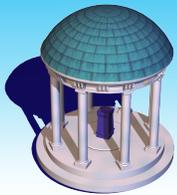
**Parametric and Nonparametric
Components**



Brain Connectivity Analysis

**Penalized Methods, Multivariate Analysis,
and Time Series Analysis**

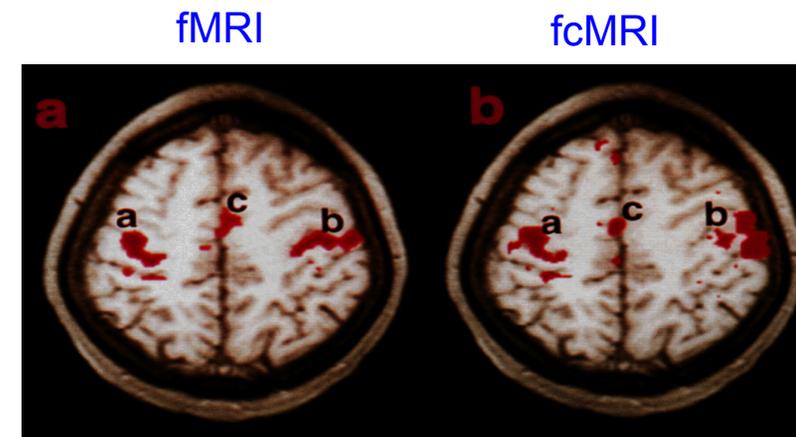
**How to *spatially and temporally* quantify
the dynamic association among
different functional regions?**



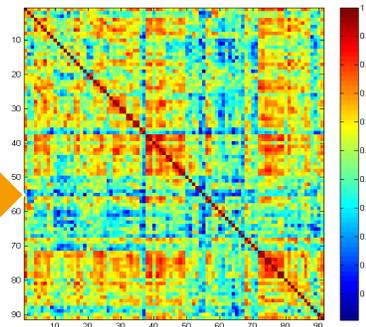
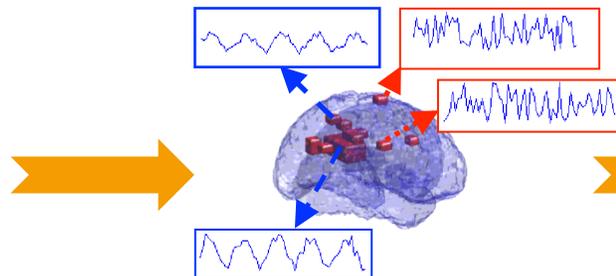
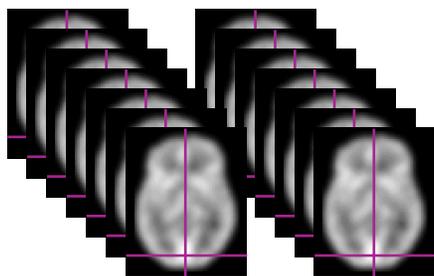
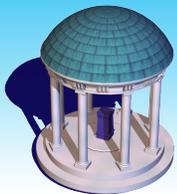
Functional connectivity is the mechanism for the coordination of activity between different neural assemblies in order to achieve a complex cognitive task or perceptual process. (Fingelkurts, Fingelkurts, Seppo Kahkonen, Fingelkurts, 2005)

Resting-State Network:

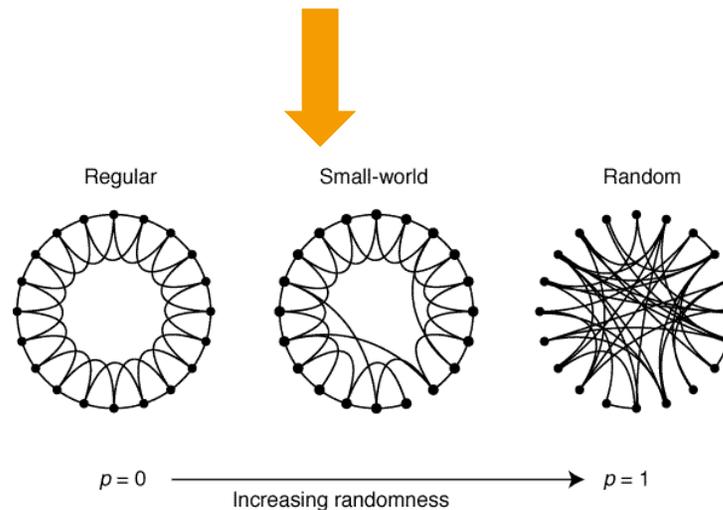
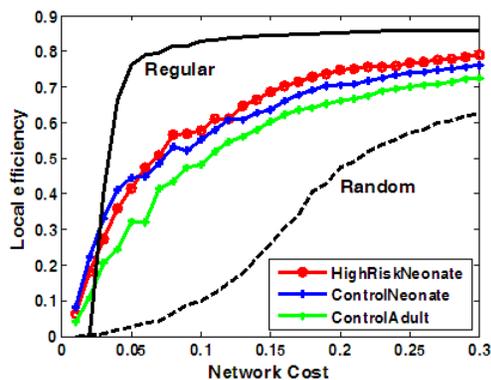
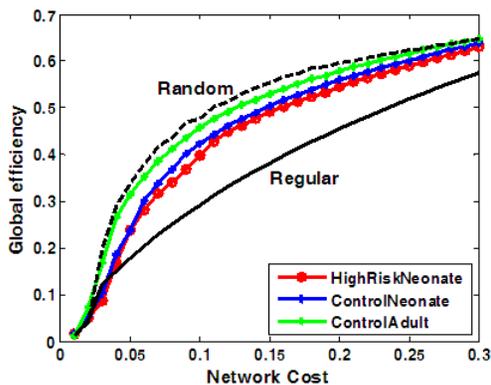
fMRI for finger tapping task;
fcMRI: contralateral motor cortex showed activation and low frequency (<0.1 Hz) fluctuations in the signal of the resting brain, revealing a high degree of temporal correlation.

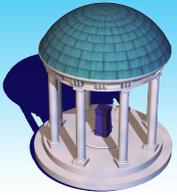


Biswal et al, JCBFM, 17:301-308, 1997

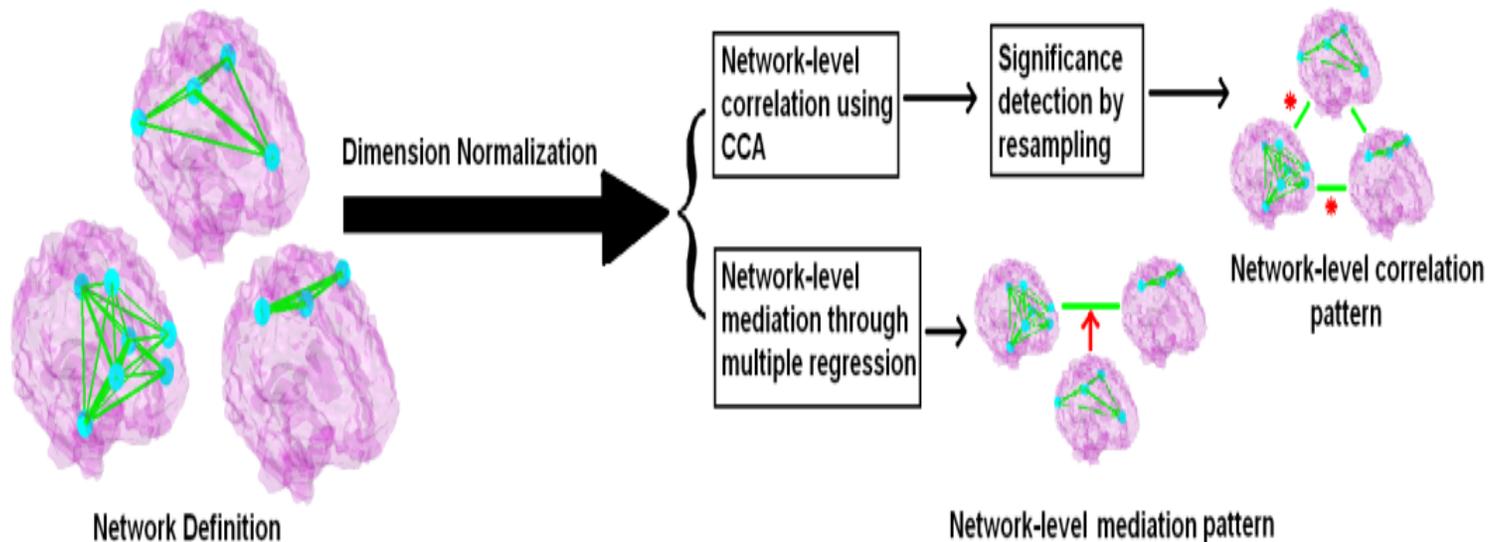


Conventional Analysis

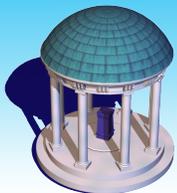




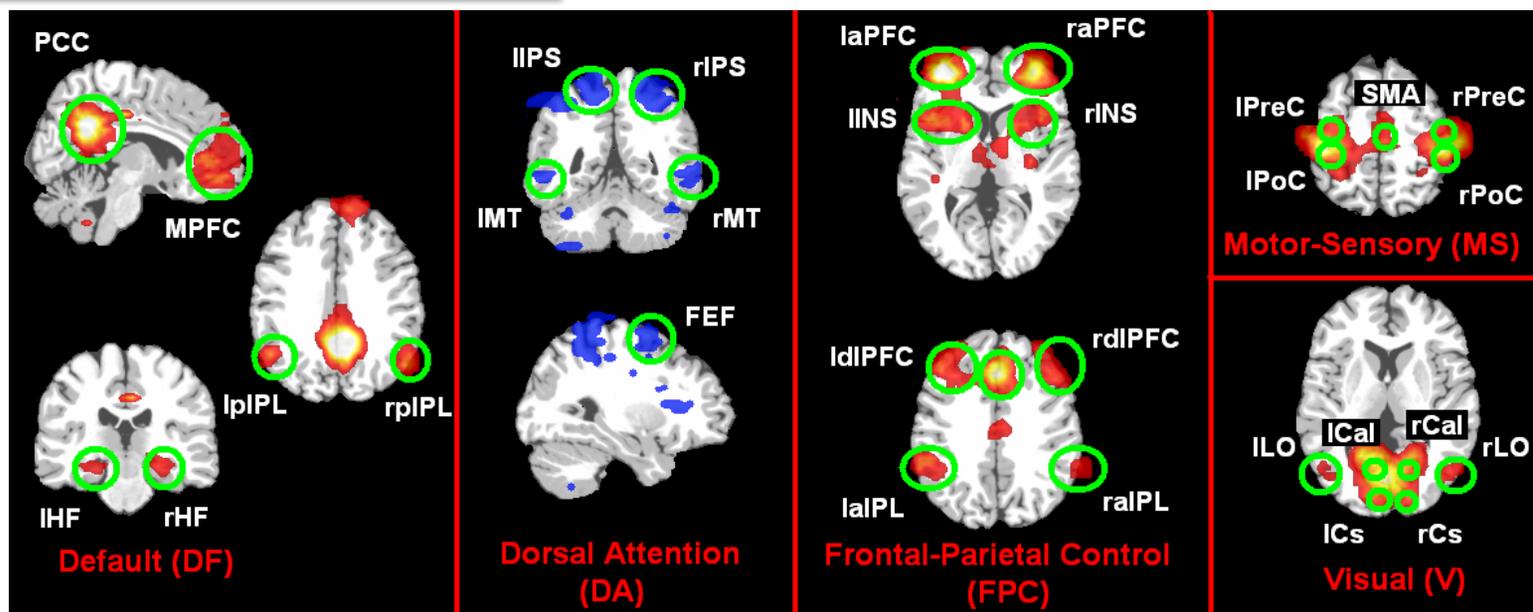
A multivariate network-level approach

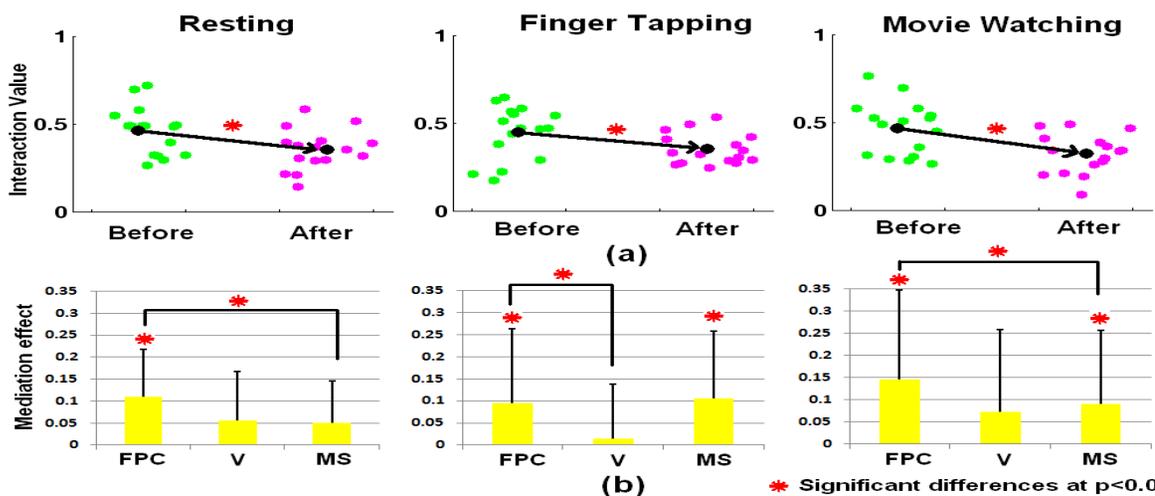
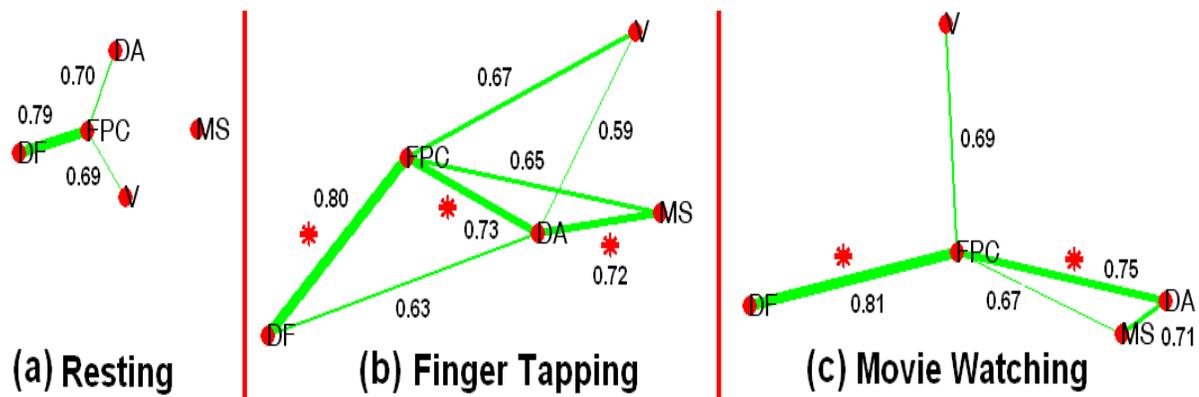
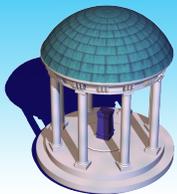


- (i) network-level correlation using CCA;
- (ii) network-level mediation analysis;
- (iii) significance detection by resampling methods;
- (iv) Network-level correlation pattern.

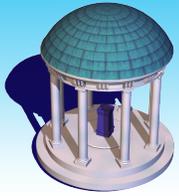


Network Definition

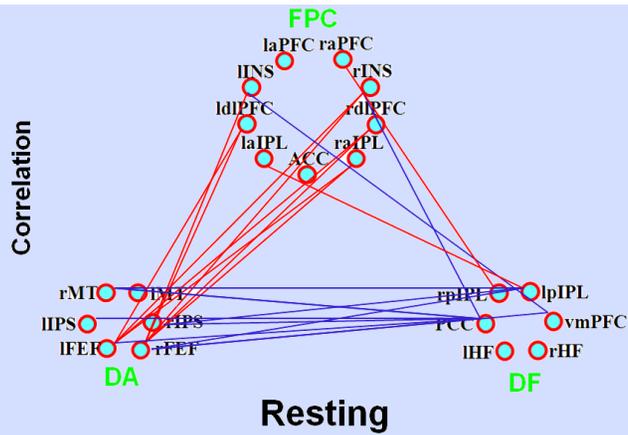


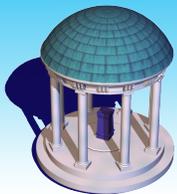


* Significant differences at p < 0.05



Selective regulation of the two opposing networks during different tasks





Bayesian Covariance Lasso

Flow Cytometry Data
11 proteins
7466 cells

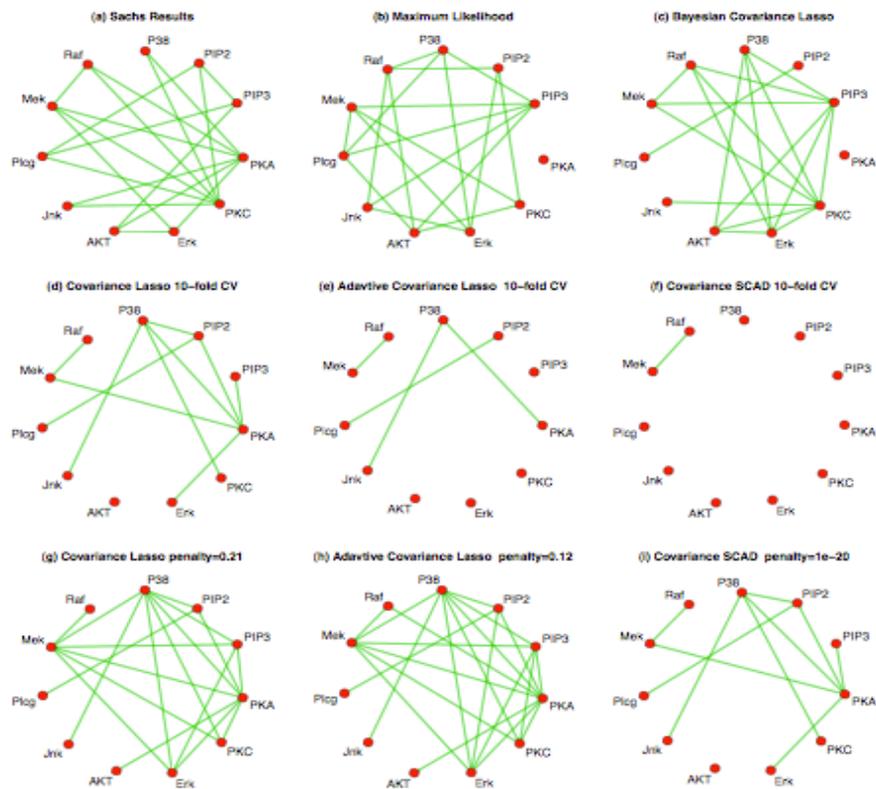
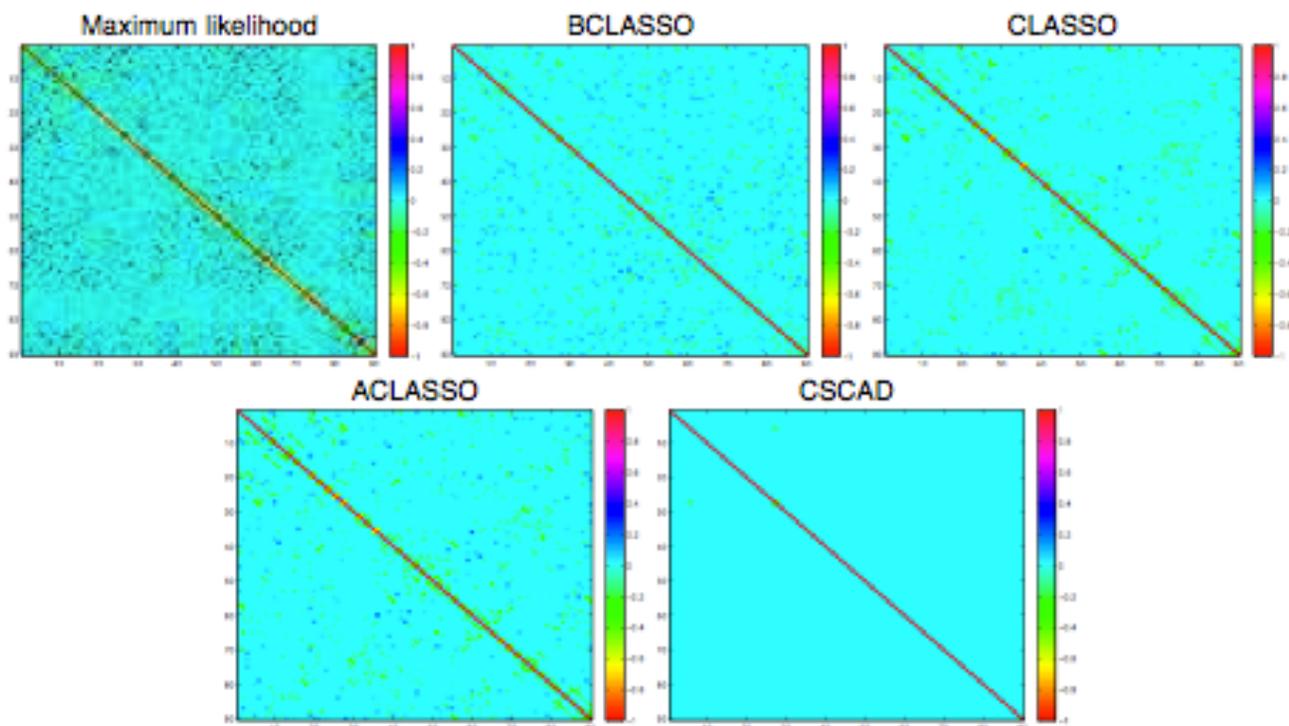
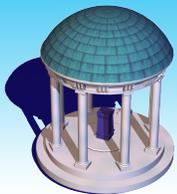
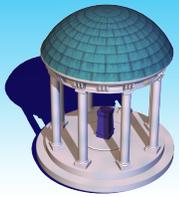


Fig. 5: Networks for 11 proteins from Sachs et al. (2003).



90 ROIs
30 subjects
2-rd fcMRI

Fig. 6: Image plots of the partial correlation matrices for 90 regions of 2-year old children's brains using the different methods



Acknowledgements

